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(54) Title: NUCLEIC ACID-ASSOCIATED PROTEINS

(57) Abstract: Various embodiments of the invention provide human nucleic acid-associated proteins (NAAP) and polynucleotides which identify and encode NAAP. Embodiments of the invention also provide expression vectors, host cells, antibodies, agonists, and antagonists. Other embodiments provide methods for diagnosing, treating, or preventing disorders associated with aberrant expression of NAAP.





NUCLEIC ACID-ASSOCIATED PROTEINS

TECHNICAL FIELD

The invention relates to novel nucleic acids, nucleic acid-associated proteins encoded by these nucleic acids, and to the use of these nucleic acids and proteins in the diagnosis, treatment, and prevention of cell proliferative, neurological, developmental, and autoimmune/inflammatory disorders, and infections. The invention also relates to the assessment of the effects of exogenous compounds on the expression of nucleic acids and nucleic acid-associated proteins.

BACKGROUND OF THE INVENTION

Multicellular organisms are comprised of diverse cell types that differ dramatically both in structure and function. The identity of a cell is determined by its characteristic pattern of gene expression, and different cell types express overlapping but distinctive sets of genes throughout development. Spatial and temporal regulation of gene expression is critical for the control of cell proliferation, cell differentiation, apoptosis, and other processes that contribute to organismal development. Furthermore, gene expression is regulated in response to extracellular signals that mediate cell-cell communication and coordinate the activities of different cell types. Appropriate gene regulation also ensures that cells function efficiently by expressing only those genes whose functions are required at a given time.

20 Transcription Factors

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Transcriptional regulatory proteins are essential for the control of gene expression. Some of these proteins function as transcription factors that initiate, activate, repress, or terminate gene transcription. Transcription factors generally bind to the promoter, enhancer, and upstream regulatory regions of a gene in a sequence-specific manner, although some factors bind regulatory elements within or downstream of a gene coding region. Transcription factors may bind to a specific region of DNA singly or as a complex with other accessory factors. (Reviewed in Lewin, B. (1990) Genes IV, Oxford University Press, New York, NY, and Cell Press, Cambridge, MA, pp. 554-570.)

The double helix structure and repeated sequences of DNA create topological and chemical features which can be recognized by transcription factors. These features are hydrogen bond donor and acceptor groups, hydrophobic patches, major and minor grooves, and regular, repeated stretches of sequence which induce distinct bends in the helix. Typically, transcription factors recognize specific DNA sequence motifs of about 20 nucleotides in length. Multiple, adjacent transcription factor-binding motifs may be required for gene regulation.

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Many transcription factors incorporate DNA-binding structural motifs which comprise either a helices or ß sheets that bind to the major groove of DNA. Four well-characterized structural motifs are helix-turn-helix, zinc finger, leucine zipper, and helix-loop-helix. Proteins containing these motifs may act alone as monomers, or they may form homo- or heterodimers that interact with DNA.

The helix-turn-helix motif consists of two α helices connected at a fixed angle by a short chain of amino acids. One of the helices binds to the major groove. Helix-turn-helix motifs are exemplified by the homeobox motif which is present in homeodomain proteins. These proteins are critical for specifying the anterior-posterior body axis during development and are conserved throughout the animal kingdom. The Antennapedia and Ultrabithorax proteins of *Drosophila melanogaster* are prototypical homeodomain proteins. (Pabo, C.O. and R.T. Sauer (1992) Annu. Rev. Biochem. 61:1053-1095.)

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The zinc finger motif, which binds zinc ions, generally contains tandem repeats of about 30 amino acids consisting of periodically spaced cysteine and histidine residues. Examples of this sequence pattern, designated C2H2 and C3HC4 ("RING" finger), have been described. (Lewin, supra.) Zinc finger proteins each contain an α helix and an antiparallel β sheet whose proximity and conformation are maintained by the zinc ion. Contact with DNA is made by the arginine preceding the α helix and by the second, third, and sixth residues of the α helix. Variants of the zinc finger motif include poorly defined cysteine-rich motifs which bind zinc or other metal ions. These motifs may not contain histidine residues and are generally nonrepetitive. The zinc finger motif may be repeated in a tandem array within a protein, such that the α helix of each zinc finger in the protein makes contact with the major groove of the DNA double helix. This repeated contact between the protein and the DNA produces a strong and specific DNA-protein interaction. The strength and specificity of the interaction can be regulated by the number of zinc finger motifs within the protein. Though originally identified in DNA-binding proteins as regions that interact directly with DNA, zinc fingers occur in a variety of proteins that do not bind DNA (Lodish, H. et al. (1995) Molecular Cell Biology, Scientific American Books, New York, NY, pp. 447-451). For example, Galcheva-Gargova, Z. et al. (1996) Science 272:1797-1802) have identified zinc finger proteins that interact with various cytokine receptors.

The C2H2-type zinc finger signature motif contains a 28 amino acid sequence, including 2 conserved Cys and 2 conserved His residues in a C-2-C-12-H-3-H type motif. The motif generally occurs in multiple tandem repeats. A cysteine-rich domain including the motif Asp-His-His-Cys (DHHC-CRD) has been identified as a distinct subgroup of zinc finger proteins. The DHHC-CRD region has been implicated in growth and development. One DHHC-CRD mutant shows defective

function of Ras, a small membrane-associated GTP-binding protein that regulates cell growth and differentiation, while other DHHC-CRD proteins probably function in pathways not involving Ras (Bartels, D.J. et al. (1999) Mol. Cell Biol. 19:6775-6787).

Zinc-finger transcription factors are often accompanied by modular sequence motifs such as the Kruppel-associated box (KRAB) and the SCAN domain. For example, the hypoalphalipoproteinemia susceptibility gene ZNF202 encodes a SCAN box and a KRAB domain followed by eight C2H2 zinc-finger motifs (Honer, C. et al. (2001) Biochim. Biophys. Acta 1517:441-448). The SCAN domain is a highly conserved, leucine-rich motif of approximately 60 amino acids found at the amino-terminal end of zinc finger transcription factors. SCAN domains are most often linked to C2H2 zinc finger motifs through their carboxyl-terminal end. Biochemical binding studies have established the SCAN domain as a selective hetero- and homotypic oligomerization domain. SCAN domain-mediated protein complexes may function to modulate the biological function of transcription factors (Schumacher, C. et al. (2000) J. Biol. Chem. 275:17173-17179).

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The KRAB (Kruppel-associated box) domain is a conserved amino acid sequence spanning approximately 75 amino acids and is found in almost one-third of the 300 to 700 genes encoding C2H2 zinc fingers. The KRAB domain is found N-terminally with respect to the finger repeats. The KRAB domain is generally encoded by two exons; the KRAB-A region or box is encoded by one exon and the KRAB-B region or box is encoded by a second exon. The function of the KRAB domain is the repression of transcription. Transcription repression is accomplished by recruitment of either the KRAB-associated protein-1, a transcriptional corepressor, or the KRAB-A interacting protein.

Proteins containing the KRAB domain are likely to play a regulatory role during development (Williams, A.J. et al. (1999) Mol. Cell Biol. 19:8526-8535). A subgroup of highly related human KRAB zinc finger proteins detectable in all human tissues is highly expressed in human T lymphoid cells (Bellefroid, E.J. et al. (1993) EMBO J. 12:1363-1374). The ZNF85 KRAB zinc finger gene, a member of the human ZNF91 family, is highly expressed in normal adult testis, in seminomas, and in the NT2/D1 teratocarcinoma cell line (Poncelet, D.A. et al. (1998) DNA Cell Biol.17:931-943).

The C4 motif is found in hormone-regulated proteins. The C4 motif generally includes only 2 repeats. A number of eukaryotic and viral proteins contain a conserved cysteine-rich domain of 40 to 60 residues (called C3HC4 zinc-finger or RING finger) that binds two atoms of zinc, and is probably involved in mediating protein-protein interactions. The 3D "cross-brace" structure of the zinc ligation system is unique to the RING domain. The spacing of the cysteines in such a domain is C-x(2)-C-x(9 to 39)-C-x(1 to 3)-H-x(2 to 3)-C-x(2)-C-x(4 to 48)-C-x(2)-C. The PHD finger is a C4HC3 zinc-finger-like motif found in nuclear proteins thought to be involved in chromatin-mediated

transcriptional regulation.

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GATA-type transcription factors contain one or two zinc finger domains which bind specifically to a region of DNA that contains the consecutive nucleotide sequence GATA. NMR studies indicate that the zinc finger comprises two irregular anti-parallel β sheets and an α helix, followed by a long loop to the C-terminal end of the finger (Ominchinski, J.G. (1993) Science 261:438-446). The helix and the loop connecting the two β-sheets contact the major groove of the DNA, while the C-terminal part, which determines the specificity of binding, wraps around into the minor groove.

The LIM motif consists of about 60 amino acid residues and contains seven conserved

cysteine residues and a histidine within a consensus sequence (Schmeichel, K.L. and Beckerle, M.C.

(1994) Cell 79:211-219). The LIM family includes transcription factors and cytoskeletal proteins

which may be involved in development, differentiation, and cell growth. One example is actin-binding

LIM protein, which may play roles in regulation of the cytoskeleton and cellular morphogenesis (Roof,

D.J. et al. (1997) J. Cell Biol. 138:575-588). The N-terminal domain of actin-binding LIM protein has

four double zinc finger motifs with the LIM consensus sequence. The C-terminal domain of actin
binding LIM protein shows sequence similarity to known actin-binding proteins such as dematin and

villin. Actin-binding LIM protein binds to F-actin through its dematin-like C-terminal domain. The

LIM domain may mediate protein-protein interactions with other LIM-binding proteins.

Myeloid cell development is controlled by tissue-specific transcription factors. Myeloid zinc finger proteins (MZF) include MZF-1 and MZF-2. MZF-1 functions in regulation of the development of neutrophilic granulocytes. A murine homolog MZF-2 is expressed in myeloid cells, particularly in the cells committed to the neutrophilic lineage. MZF-2 is down-regulated by G-CSF and appears to have a unique function in neutrophil development (Murai, K. et al. (1997) Genes Cells 2:581-591).

The leucine zipper motif comprises a stretch of amino acids rich in leucine which can form an amphipathic α helix. This structure provides the basis for dimerization of two leucine zipper proteins. The region adjacent to the leucine zipper is usually basic, and upon protein dimerization, is optimally positioned for binding to the major groove. Proteins containing such motifs are generally referred to as bZIP transcription factors. The leucine zipper motif is found in the proto-oncogenes Fos and Jun, which comprise the heterodimeric transcription factor AP1 involved in cell growth and the determination of cell lineage (Papavassiliou, A.G. (1995) N. Engl. J. Med. 332:45-47).

The helix-loop-helix motif (HLH) consists of a short α helix connected by a loop to a longer α helix. The loop is flexible and allows the two helices to fold back against each other and to bind to DNA. The transcription factor Myc contains a prototypical HLH motif.

The NF-kappa-B/Rel signature defines a family of eukaryotic transcription factors involved in oncogenesis, embryonic development, differentiation and immune response. Most transcription factors containing the Rel homology domain (RHD) bind as dimers to a consensus DNA sequence motif termed kappa-B. Members of the Rel family share a highly conserved 300 amino acid domain termed the Rel homology domain. The characteristic Rel C-terminal domain is involved in gene activation and cytoplasmic anchoring functions. Proteins known to contain the RHD domain include vertebrate nuclear factor NF-kappa-B, which is a heterodimer of a DNA-binding subunit and the transcription factor p65, mammalian transcription factor RelB, and vertebrate proto-oncogene c-rel, a protein associated with differentiation and lymphopoiesis (Kabrun, N. and Enrietto, P.J. (1994) Semin. Cancer Biol. 5:103-112).

A DNA binding motif termed ARID (AT-rich interactive domain) distinguishes an evolutionarily conserved family of proteins. The approximately 100-residue ARID sequence is present in a series of proteins strongly implicated in the regulation of cell growth, development, and tissue-specific gene expression. ARID proteins include Bright (a regulator of B-cell-specific gene expression), dead ringer (involved in development), and MRF-2 (which represses expression from the cytomegalovirus enhancer) (Dallas, P.B. et al. (2000) Mol. Cell Biol. 20:3137-3146).

The ELM2 (Egl-27 and MTA1 homology 2) domain is found in metastasis-associated protein MTA1 and protein ER1. The *Caenorhabditis elegans* gene egl-27 is required for embryonic patterning MTA1, a human gene with elevated expression in metastatic carcinomas, is a component of a protein complex with histone deacetylase and nucleosome remodelling activities (Solari, F. et al. (1999) Development 126:2483-2494). The ELM2 domain is usually found to the N terminus of a myb-like DNA binding domain. ELM2 is also found associated with an ARID DNA.

The Iroquois (Irx) family of genes are found in nematodes, insects and vertebrates. Irx genes usually occur in one or two genomic clusters of three genes each and encode transcriptional

25 controllers that possess a characteristic homeodomain. The Irx genes function early in development to specify the identity of diverse territories of the body. Later in development in both *Drosophila* and vertebrates, the Irx genes function again to subdivide those territories into smaller domains. (For a review of Iroquois genes, see Cavodeassi, F. et al. (2001) Development 128:2847-2855.) For example, mouse and human Irx4 proteins are 83% conserved and their 63-aa homeodomain is more than 93% identical to that of the *Drosophila* Iroquois patterning genes. Irx4 transcripts are predominantly expressed in the cardiac ventricles. The homeobox gene Irx4 mediates ventricular differentiation during cardiac development (Bruneau, B.G. et al. (2000) Dev. Biol. 217:266-77).

Histidine triad (HIT) proteins share residues in distinctive dimeric, 10-stranded half-barrel

structures that form two identical purine nucleotide-binding sites. Hint (histidine triad nucleotide-binding protein)-related proteins, found in all forms of life, and fragile histidine triad (Fhit)-related proteins, found in animals and fungi, represent the two main branches of the HIT superfamily. Fhit homologs bind and cleave diadenosine polyphosphates. Fhit-Ap(n)A complexes appear to function in a proapoptotic tumor suppression pathway in epithelial tissues (Brenner C. et al. (1999) J. Cell Physiol.181:179-187).

Most transcription factors contain characteristic DNA binding motifs, and variations on the above motifs and new motifs have been and are currently being characterized. (Faisst, S. and S. Meyer (1992) Nucleic Acids Res. 20:3-26.)

10 Chromatin Associated Proteins

In the nucleus, DNA is packaged into chromatin, the compact organization of which limits the accessibility of DNA to transcription factors and plays a key role in gene regulation. (Lewin, *supra*, pp. 409-410.) The compact structure of chromatin is determined and influenced by chromatin-associated proteins such as the histones, the high mobility group (HMG) proteins, and the chromodomain proteins. There are five classes of histones, H1, H2A, H2B, H3, and H4, all of which are highly basic, low molecular weight proteins. The fundamental unit of chromatin, the nucleosome, consists of 200 base pairs of DNA associated with two copies each of H2A, H2B, H3, and H4. H1 links adjacent nucleosomes. HMG proteins are low molecular weight, non-histone proteins that may play a role in unwinding DNA and stabilizing single-stranded DNA. Chromodomain proteins play a key role in the formation of highly compacted heterochromatin, which is transcriptionally silent.

Diseases and Disorders Related to Gene Regulation

Many neoplastic disorders in humans can be attributed to inappropriate gene expression. Malignant cell growth may result from either excessive expression of tumor promoting genes or insufficient expression of tumor suppressor genes. (Cleary, M.L. (1992) Cancer Surv. 15:89-104.)

The zinc finger-type transcriptional regulator WT1 is a tumor-suppressor protein that is inactivated in children with Wilm's tumor. The oncogene bcl-6, which plays an important role in large-cell lymphoma, is also a zinc-finger protein (Papavassiliou, A.G. (1995) N. Engl. J. Med. 332:45-47). Chromosomal translocations may also produce chimeric loci that fuse the coding sequence of one gene with the regulatory regions of a second unrelated gene. Such an arrangement likely results in inappropriate gene transcription, potentially contributing to malignancy. In Burkitt's lymphoma, for example, the transcription factor Myc is translocated to the immunoglobulin heavy chain locus, greatly enhancing Myc expression and resulting in rapid cell growth leading to leukemia (Latchman, D.S. (1996) N. Engl. J. Med. 334:28-33).

In addition, the immune system responds to infection or trauma by activating a cascade of events that coordinate the progressive selection, amplification, and mobilization of cellular defense mechanisms. A complex and balanced program of gene activation and repression is involved in this process. However, hyperactivity of the immune system as a result of improper or insufficient 5 regulation of gene expression may result in considerable tissue or organ damage. This damage is welldocumented in immunological responses associated with arthritis, allergens, heart attack, stroke, and infections. (Isselbacher et al. Harrison's Principles of Internal Medicine, 13/e, McGraw Hill, Inc. and Teton Data Systems Software, 1996.) The causative gene for autoimmune polyendocrinopathycandidiasis-ectodermal dystrophy (APECED) was recently isolated and found to encode a protein with two PHD-type zinc finger motifs (Bjorses, P. et al. (1998) Hum. Mol. Genet. 7:1547-1553).

Furthermore, the generation of multicellular organisms is based upon the induction and coordination of cell differentiation at the appropriate stages of development. Central to this process is differential gene expression, which confers the distinct identities of cells and tissues throughout the body. Failure to regulate gene expression during development could result in developmental disorders. Human developmental disorders caused by mutations in zinc finger-type transcriptional regulators include: urogenital developmental abnormalities associated with WT1; Greig cephalopolysyndactyly, Pallister-Hall syndrome, and postaxial polydactyly type A (GLI3), and Townes-Brocks syndrome, characterized by anal, renal, limb, and ear abnormalities (SALL1) (Engelkamp, D. and V. van: Heyningen (1996) Curr. Opin. Genet. Dev. 6:334-342; Kohlhase, J. et al. (1999) Am. J. Hum. Genet. 64:435-445).

Human acute leukemias involve reciprocal chromosome translocations that fuse the ALL-1 gene located at chromosome region 11q23 to a series of partner genes positioned on a variety of human chromosomes. The fused genes encode chimeric proteins. The AF17 gene encodes a protein of 1093 amino acids, containing a leucine-zipper dimerization motif located 3' of the fusion point and a 25 cysteine-rich domain at the N terminus that shows homology to a domain within the protein Br140 (peregrin) (Prasad R. et al. (1994) Proc. Natl. Acad. Sci. U S A 91:8107-8111).

SYNTHESIS OF NUCLEIC ACIDS

<u>Polymerases</u>

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DNA and RNA replication are critical processes for cell replication and function. DNA and RNA replication are mediated by the enzymes DNA and RNA polymerase, respectively, by a "templating" process in which the nucleotide sequence of a DNA or RNA strand is copied by complementary base-pairing into a complementary nucleic acid sequence of either DNA or RNA. However, there are fundamental differences between the two processes.

DNA polymerase catalyzes the stepwise addition of a deoxyribonucleotide to the 3'-OH end of a polynucleotide strand (the primer strand) that is paired to a second (template) strand. The new DNA strand therefore grows in the 5' to 3' direction (Alberts, B. et al. (1994) The Molecular Biology of the Cell, Garland Publishing Inc., New York, NY, pp 251-254). The substrates for the polymerization reaction are the corresponding deoxynucleotide triphosphates which must base-pair with the correct nucleotide on the template strand in order to be recognized by the polymerase. Because DNA exists as a double-stranded helix, each of the two strands may serve as a template for the formation of a new complementary strand. Each of the two daughter cells of a dividing cell therefore inherits a new DNA double helix containing one old and one new strand. Thus, DNA is said to be replicated "semiconservatively" by DNA polymerase. In addition to the synthesis of new DNA, DNA polymerase is also involved in the repair of damaged DNA as discussed below under "Ligases."

In contrast to DNA polymerase, RNA polymerase uses a DNA template strand to "transcribe" DNA into RNA using ribonucleotide triphosphates as substrates. Like DNA polymerization, RNA polymerization proceeds in a 5' to 3' direction by addition of a ribonucleoside monophosphate to the 3'-OH end of a growing RNA chain. DNA transcription generates messenger RNAs (mRNA) that carry information for protein synthesis, as well as the transfer, ribosomal, and other RNAs that have structural or catalytic functions. In eukaryotes, three discrete RNA polymerases synthesize the three different types of RNA (Alberts, *supra*, pp. 367-368). RNA polymerase I makes the large ribosomal RNAs, RNA polymerase II makes the mRNAs that will be translated into proteins, and RNA polymerase III makes a variety of small, stable RNAs, including 5S ribosomal RNA and the transfer RNAs (tRNA). In all cases, RNA synthesis is initiated by binding of the RNA polymerase to a promoter region on the DNA and synthesis begins at a start site within the promoter. Synthesis is completed at a stop (termination) signal in the DNA whereupon both the polymerase and the completed RNA chain are released.

25 Ligases

DNA repair is the process by which accidental base changes, such as those produced by oxidative damage, hydrolytic attack, or uncontrolled methylation of DNA, are corrected before replication or transcription of the DNA can occur. Because of the efficiency of the DNA repair process, fewer than one in a thousand accidental base changes causes a mutation (Alberts, *supra*, pp. 245-249). The three steps common to most types of DNA repair are (1) excision of the damaged or altered base or nucleotide by DNA nucleases, (2) insertion of the correct nucleotide in the gap left by the excised nucleotide by DNA polymerase using the complementary strand as the template and, (3) sealing the break left between the inserted nucleotide(s) and the existing DNA strand by DNA ligase.

In the last reaction, DNA ligase uses the energy from ATP hydrolysis to activate the 5' end of the broken phosphodiester bond before forming the new bond with the 3'-OH of the DNA strand. In Bloom's syndrome, an inherited human disease, individuals are partially deficient in DNA ligation and consequently have an increased incidence of cancer (Alberts, *supra* p. 247).

Nucleases

Nucleases comprise enzymes that hydrolyze both DNA (DNase) and RNA (Rnase). They serve different purposes in nucleic acid metabolism. Nucleases hydrolyze the phosphodiester bonds between adjacent nucleotides either at internal positions (endonucleases) or at the terminal 3' or 5' nucleotide positions (exonucleases). A DNA exonuclease activity in DNA polymerase, for example, serves to remove improperly paired nucleotides attached to the 3'-OH end of the growing DNA strand by the polymerase and thereby serves a "proofreading" function. As mentioned above, DNA endonuclease activity is involved in the excision step of the DNA repair process.

RNases also serve a variety of functions. For example, RNase P is a ribonucleoprotein enzyme which cleaves the 5' end of pre-tRNAs as part of their maturation process. RNase H digests the RNA strand of an RNA/DNA hybrid. Such hybrids occur in cells invaded by retroviruses, and RNase H is an important enzyme in the retroviral replication cycle. Pancreatic RNase secreted by the pancreas into the intestine hydrolyzes RNA present in ingested foods. RNase activity in serum and cell extracts is elevated in a variety of cancers and infectious diseases (Schein, C.H. (1997) Nat. Biotechnol. 15:529-536). Regulation of RNase activity is being investigated as a means to control tumor angiogenesis, allergic reactions, viral infection and replication, and fungal infections.

MODIFICATION OF NUCLEIC ACIDS

Methylases

Methylation of specific nucleotides occurs in both DNA and RNA, and serves different functions in the two macromolecules. Methylation of cytosine residues to form 5-methyl cytosine in DNA occurs specifically in CG sequences which are base-paired with one another in the DNA double-helix. The pattern of methylation is passed from generation to generation during DNA replication by an enzyme called "maintenance methylase" that acts preferentially on those CG sequences that are base-paired with a CG sequence that is already methylated. Such methylation appears to distinguish active from inactive genes by preventing the binding of regulatory proteins that "turn on" the gene, but permiting the binding of proteins that inactivate the gene (Alberts, supra pp. 448-451). In RNA metabolism, "tRNA methylase" produces one of several nucleotide modifications in tRNA that affect the conformation and base-pairing of the molecule and facilitate the recognition of the appropriate mRNA codons by specific tRNAs. The primary methylation pattern is the

dimethylation of guanine residues to form N,N-dimethyl guanine.

Helicases and Single-stranded Binding Proteins

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Helicases are enzymes that destabilize and unwind double helix structures in both DNA and RNA. Since DNA replication occurs more or less simultaneously on both strands, the two strands must first separate to generate a replication "fork" for DNA polymerase to act on. Two types of replication proteins contribute to this process, DNA helicases and single-stranded binding proteins. DNA helicases hydrolyze ATP and use the energy of hydrolysis to separate the DNA strands. Single-stranded binding proteins (SSBs) then bind to the exposed DNA strands, without covering the bases, thereby temporarily stabilizing them for templating by the DNA polymerase (Alberts, *supra*, pp. 255-256).

RNA helicases also alter and regulate RNA conformation and secondary structure. Like the DNA helicases, RNA helicases utilize energy derived from ATP hydrolysis to destabilize and unwind RNA duplexes. The most well-characterized and ubiquitous family of RNA helicases is the DEADbox family, so named for the conserved B-type ATP-binding motif which is diagnostic of proteins in this family. Over 40 DEAD-box helicases have been identified in organisms as diverse as bacteria, insects, yeast, amphibians, mammals, and plants. DEAD-box helicases function in diverse processes such as translation initiation, splicing, ribosome assembly, and RNA editing, transport, and stability. Examples of these RNA helicases include yeast Drs1 protein, which is involved in ribosomal RNA processing; yeast TIF1 and TIF2 and mammalian eIF-4A, which are essential to the initiation of RNA translation; and human p68 antigen, which regulates cell growth and division (Ripmaster, T.L. et al. (1992) Proc. Natl. Acad. Sci. USA 89:11131-11135; Chang, T.-H. et al. (1990) Proc. Natl. Acad. Sci. USA 87:1571-1575). These RNA helicases demonstrate strong sequence homology over a stretch of some 420 amino acids. Included among these conserved sequences are the consensus sequence for the A motif of an ATP binding protein; the "DEAD box" sequence, associated with ATPase activity; the sequence SAT, associated with the actual helicase unwinding region; and an octapeptide consensus sequence, required for RNA binding and ATP hydrolysis (Pause, A. et al. (1993) Mol. Cell Biol. 13:6789-6798). Differences outside of these conserved regions are believed to reflect differences in the functional roles of individual proteins (Chang, T.H. et al. (1990) Proc. Natl. Acad. Sci. USA 87:1571-1575).

Some DEAD-box helicases play tissue- and stage-specific roles in spermatogenesis and embryogenesis. Overexpression of the DEAD-box 1 protein (DDX1) may play a role in the progression of neuroblastoma (Nb) and retinoblastoma (Rb) tumors (Godbout, R. et al. (1998) J. Biol. Chem. 273:21161-21168). These observations suggest that DDX1 may promote or enhance tumor

progression by altering the normal secondary structure and expression levels of RNA in cancer cells. Other DEAD-box helicases have been implicated either directly or indirectly in tumorigenesis. (Discussed in Godbout, supra.) For example, murine p68 is mutated in ultraviolet light-induced tumors, and human DDX6 is located at a chromosomal breakpoint associated with B-cell lymphoma. Similarly, a chimeric protein comprised of DDX10 and NUP98, a nucleoporin protein, may be involved in the pathogenesis of certain myeloid malignancies.

Topoisomerases

Besides the need to separate DNA strands prior to replication, the two strands must be "unwound" from one another prior to their separation by DNA helicases. This function is performed by proteins known as DNA topoisomerases. DNA topoisomerase effectively acts as a reversible nuclease that hydrolyzes a phosphodiesterase bond in a DNA strand, permits the two strands to rotate freely about one another to remove the strain of the helix, and then rejoins the original phosphodiester bond between the two strands. Topoisomerases are essential enzymes responsible for the topological rearrangement of DNA brought about by transcription, replication, chromatin formation, recombination, and chromosome segregation. Superhelical coils are introduced into DNA by the passage of processive enzymes such as RNA polymerase, or by the separation of DNA strands by a helicase prior to replication. Knotting and concatenation can occur in the process of DNA synthesis, storage, and repair. All topoisomerases work by breaking a phosphodiester bond in the ribosephosphate backbone of DNA. A catalytic tyrosine residue on the enzyme makes a nucleophilic attack on the scissile phosphodiester bond, resulting in a reaction intermediate in which a covalent bond is formed between the enzyme and one end of the broken strand. A tyrosine-DNA phosphodiesterase functions in DNA repair by hydrolyzing this bond in occasional dead-end topoisomerase I-DNA intermediates (Pouliot, J.J. et al. (1999) Science 286:552-555).

Two types of DNA topoisomerase exist, types I and II. Type I topoisomerases work as monomers, making a break in a single strand of DNA while type II topoisomerases, working as homodimers, cleave both strands. DNA Topoisomerase I causes a single-strand break in a DNA helix to allow the rotation of the two strands of the helix about the remaining phosphodiester bond in the opposite strand. DNA topoisomerase II causes a transient break in both strands of a DNA helix where two double helices cross over one another. This type of topoisomerase can efficiently separate two interlocked DNA circles (Alberts, *supra*, pp.260-262). Type II topoisomerases are largely confined to proliferating cells in eukaryotes, such as cancer cells. For this reason they are targets for anticancer drugs. Topoisomerase II has been implicated in multi-drug resistance (MDR) as it appears to aid in the repair of DNA damage inflicted by DNA binding agents such as doxorubicin and

vincristine.

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The topoisomerase I family includes topoisomerases I and III (topo I and topo III). The crystal structure of human topoisomerase I suggests that rotation about the intact DNA strand is partially controlled by the enzyme. In this "controlled rotation" model, protein-DNA interactions limit the rotation, which is driven by torsional strain in the DNA (Stewart, L. et al. (1998) Science 379:1534-1541). Structurally, topo I can be recognized by its catalytic tyrosine residue and a number of other conserved residues in the active site region. Topo I is thought to function during transcription. Two topo IIIs are known in humans, and they are homologous to prokaryotic topoisomerase I, with a conserved tyrosine and active site signature specific to this family. Topo III has been suggested to play a role in meiotic recombination. A mouse topo III is highly expressed in testis tissue and its expression increases with the increase in the number of cells in pachytene (Seki, T. et al. (1998) J. Biol. Chem. 273:28553-28556).

The topoisomerase II family includes two isozymes (IIα and IIβ) encoded by different genes. Topo II cleaves double stranded DNA in a reproducible, nonrandom fashion, preferentially in an AT rich region, but the basis of cleavage site selectivity is not known. Structurally, topo II is made up of four domains, the first two of which are structurally similar and probably distantly homologous to similar domains in eukaryotic topo I. The second domain bears the catalytic tyrosine, as well as a highly conserved pentapeptide. The IIα isoform appears to be responsible for unlinking DNA during chromosome segregation. Cell lines expressing IIα but not IIβ suggest that IIβ is dispensable in cellular processes; however, IIβ knockout mice died perinatally due to a failure in neural development. That the major abnormalities occurred in predominantly late developmental events (neurogenesis) suggests that IIβ is needed not at mitosis, but rather during DNA repair (Yang, X. et al. (2000) Science 287:131-134).

Topoisomerases have been implicated in a number of disease states, and topoisomerase poisons have proven to be effective anti-tumor drugs for some human malignancies. Topo I is mislocalized in Fanconi's anemia, and may be involved in the chromosomal breakage seen in this disorder (Wunder, E. (1984) Hum. Genet. 68:276-281). Overexpression of a truncated topo III in ataxia-telangiectasia (A-T) cells partially suppresses the A-T phenotype, probably through a dominant negative mechanism. This suggests that topo III is deregulated in A-T (Fritz, E. et al. (1997) Proc. Natl. Acad. Sci. USA 94:4538-4542). Topo III also interacts with the Bloom's Syndrome gene product, and has been suggested to have a role as a tumor suppressor (Wu, L. et al. (2000) J. Biol. Chem. 275:9636-9644). Aberrant topo II activity is often associated with cancer or increased cancer risk. Greatly lowered topo II activity has been found in some, but not all A-T cell lines (Mohamed, R.

et al. (1987) Biochem. Biophys. Res. Commun. 149:233-238). On the other hand, topo II can break DNA in the region of the A-T gene (ATM), which controls all DNA damage-responsive cell cycle checkpoints (Kaufmann, W.K. (1998) Proc. Soc. Exp. Biol. Med. 217:327-334). The ability of topoisomerases to break DNA has been used as the basis of antitumor drugs. Topoisomerase poisons 5 act by increasing the number of dead-end covalent DNA-enzyme complexes in the cell, ultimately triggering cell death pathways (Fortune, J.M. and N. Osheroff (2000) Prog. Nucleic Acid Res. Mol. Biol. 64:221-253; Guichard, S.M. and M.K. Danks (1999) Curr. Opin. Oncol. 11:482-489). Antibodies against topo I are found in the serum of systemic sclerosis patients, and the levels of the antibody may be used as a marker of pulmonary involvement in the disease (Diot, E. et al. (1999) Chest 116:715-720). Finally, the DNA binding region of human topo I has been used as a DNA delivery vehicle for gene therapy (Chen, T.Y. et al. (2000) Appl. Microbiol. Biotechnol. 53:558-567).

Recombinases

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Genetic recombination is the process of rearranging DNA sequences within an organism's genome to provide genetic variation for the organism in response to changes in the environment. DNA recombination allows variation in the particular combination of genes present in an individual's genome, as well as the timing and level of expression of these genes. (See Alberts, supra pp. 263-273.) Two broad classes of genetic recombination are commonly recognized, general recombination and site-specific recombination. General recombination involves genetic exchange between any homologous pair of DNA sequences usually located on two copies of the same chromosome. The process is aided by enzymes, recombinases, that "nick" one strand of a DNA duplex more or less randomly and permit exchange with a complementary strand on another duplex. The process does not normally change the arrangement of genes in a chromosome. In site-specific recombination, the recombinase recognizes specific nucleotide sequences present in one or both of the recombining molecules. Base-pairing is not involved in this form of recombination and therefore it does not require DNA homology between the recombining molecules. Unlike general recombination, this form of recombination can alter the relative positions of nucleotide sequences in chromosomes.

RNA METABOLISM

Ribonucleic acid (RNA) is a linear single-stranded polymer of four nucleotides, ATP, CTP, UTP, and GTP. In most organisms, RNA is transcribed as a copy of deoxyribonucleic acid (DNA), the genetic material of the organism. In retroviruses RNA rather than DNA serves as the genetic material. RNA copies of the genetic material encode proteins or serve various structural, catalytic, or regulatory roles in organisms. RNA is classified according to its cellular localization and function. Messenger RNAs (mRNAs) encode polypeptides. Ribosomal RNAs (rRNAs) are assembled, along

with ribosomal proteins, into ribosomes, which are cytoplasmic particles that translate mRNA into polypeptides. Transfer RNAs (tRNAs) are cytosolic adaptor molecules that function in mRNA translation by recognizing both an mRNA codon and the amino acid that matches that codon. Heterogeneous nuclear RNAs (hnRNAs) include mRNA precursors and other nuclear RNAs of various sizes. Small nuclear RNAs (snRNAs) are a part of the nuclear spliceosome complex that removes intervening, non-coding sequences (introns) and rejoins exons in pre-mRNAs.

Proteins are associated with RNA during its transcription from DNA, RNA processing, and translation of mRNA into protein. Proteins are also associated with RNA as it is used for structural, catalytic, and regulatory purposes.

10 RNA Processing

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Ribosomal RNAs (rRNAs) are assembled, along with ribosomal proteins, into ribosomes, which are cytoplasmic particles that translate messenger RNA (mRNA) into polypeptides. The eukaryotic ribosome is composed of a 60S (large) subunit and a 40S (small) subunit, which together form the 80S ribosome. In addition to the 18S, 28S, 5S, and 5.8S rRNAs, ribosomes contain from 50 to over 80 different ribosomal proteins, depending on the organism. Ribosomal proteins are classified according to which subunit they belong (i.e., L, if associated with the large 60S large subunit or S if associated with the small 40S subunit). *E. coli* ribosomes have been the most thoroughly studied and contain 50 proteins, many of which are conserved in all life forms. The structures of nine ribosomal proteins have been solved to less than 3.0D resolution (i.e., S5, S6, S17, L1, L6, L9, L12, L14, L30), revealing common motifs, such as b-a-b protein folds in addition to acidic and basic RNA-binding motifs positioned between b-strands. Most ribosomal proteins are believed to contact rRNA directly (reviewed in Liljas, A. and Garber, M. (1995) Curr. Opin. Struct. Biol. 5:721–727; see also Woodson, S.A. and Leontis, N.B. (1998) Curr. Opin. Struct. Biol. 8:294-300; Ramakrishnan, V. and White, S.W. (1998) Trends Biochem. Sci. 23:208-212).

Ribosomal proteins may undergo post-translational modifications or interact with other ribosome-associated proteins to regulate translation. For example, the highly homologous 40S ribosomal protein S6 kinases (S6K1 and S6K2) play a key role in the regulation of cell growth by controlling the biosynthesis of translational components which make up the protein synthetic apparatus (including the ribosomal proteins). In the case of S6K1, at least eight phosphorylation sites are believed to mediate kinase activation in a hierarchical fashion (Dufner and Thomas (1999) Exp. Cell. Res. 253:100-109). Some of the ribosomal proteins, including L1, also function as translational repressors by binding to polycistronic mRNAs encoding ribosomal proteins (reviewed in Liljas, supra and Garber, supra).

Recent evidence suggests that a number of ribosomal proteins have secondary functions independent of their involvement in protein biosynthesis. These proteins function as regulators of cell proliferation and, in some instances, as inducers of cell death. For example, the expression of human ribosomal protein L13a has been shown to induce apoptosis by arresting cell growth in the G2/M phase of the cell cycle. Inhibition of expression of L13a induces apoptosis in target cells, which suggests that this protein is necessary, in the appropriate amount, for cell survival. Similar results have been obtained in yeast where inactivation of yeast homologues of L13a, rp22 and rp23, results in severe growth retardation and death. A closely related ribosomal protein, L7, arrests cells in G1 and also induces apoptosis. Thus, it appears that a subset of ribosomal proteins may function as cell cycle checkpoints and compose a new family of cell proliferation regulators.

Mapping of individual ribosomal proteins on the surface of intact ribosomes is accomplished using 3D immunocryoelectronmicroscopy, whereby antibodies raised against specific ribosomal proteins are visualized. Progress has been made toward the mapping of L1, L7, and L12 while the structure of the intact ribosome has been solved to only 20-25D resolution and inconsistencies exist among different crude structures (Frank, J. (1997) Curr. Opin. Struct. Biol. 7:266-272).

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Three distinct sites have been identified on the ribosome. The aminoacyl-tRNA acceptor site (A site) receives charged tRNAs (with the exception of the initiator-tRNA). The peptidyl-tRNA site (P site) binds the nascent polypeptide as the amino acid from the A site is added to the elongating chain. Deacylated tRNAs bind in the exit site (E site) prior to their release from the ribosome. The structure of the ribosome is reviewed in Stryer, L. (1995) <u>Biochemistry</u>, W.H. Freeman and Company, New York NY, pp. 888-9081; Lodish, H. et al. (1995) <u>Molecular Cell Biology</u>, Scientific American Books, New York NY, pp. 119-138; and Lewin, B (1997) <u>Genes VI</u>, Oxford University Press, Inc. New York, NY).

Various proteins are necessary for processing of transcribed RNAs in the nucleus. Pre-mRNA processing steps include capping at the 5' end with methylguanosine, polyadenylating the 3' end, and splicing to remove introns. The primary RNA transript from DNA is a faithful copy of the gene containing both exon and intron sequences, and the latter sequences must be cut out of the RNA transcript to produce a mRNA that codes for a protein. This "splicing" of the mRNA sequence takes place in the nucleus with the aid of a large, multicomponent ribonucleoprotein complex known as a spliceosome. The spliceosomal complex is comprised of five small nuclear ribonucleoprotein particles (snRNPs) designated U1, U2, U4, U5, and U6. Each snRNP contains a single species of snRNA and about ten proteins. The RNA components of some snRNPs recognize and base-pair with intron consensus sequences. The protein components mediate spliceosome assembly and the splicing

reaction. Autoantibodies to snRNP proteins are found in the blood of patients with systemic lupus erythematosus (Stryer, L. (1995) <u>Biochemistry</u>, W.H. Freeman and Company, New York NY, p. 863).

Several splicing regulatory proteins have been identified in <u>Drosophila</u>. Human (HsSWAP) and mouse (MmSWAP) homologs of the suppressor-of-white-apricot (su(wa)) gene have been cloned and characterized. HsSWAP and MmSWAP both have five highly homologous regions to su(wa), including an arginine/serine-rich domain and two repeated modules that are homologous to regions in the constitutive splicing factor, SPP91/PRP21. Mammalian SWAP mRNAs are alternatively spliced at the same splice sites as in <u>Drosophila</u>. The splice junctions of the <u>Drosophila</u> and mammalian regulated introns are conserved. Thus, research suggests that the mammalian SWAP gene functions as a vertebrate alternative splicing regulator (Denhez, F. and Lafyatis, R. (1994) Biol. Chem. 269:16170-16179).

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Serine- and arginine-rich pre-mRNA splicing factors (SR proteins) are phosphorylated before they regulate splicing events. SRrp86 (SR-related protein of 86 kDa) is a novel SR protein containing a single amino-terminal RNA recognition motif and two carboxy-terminal domains rich in serine-arginine (SR) dipeptides. SRrp86 activates splicing in the presence of SRp20. However, it inhibits the in vitro and in vivo activation of specific splice sites by SR proteins, including ASF/SF2, SC35, and SRp55. Research suggests that pairwise combination of SRrp86 with specific SR proteins leads to altered splicing efficiency and differential splice site selection (Barnard, D.C. and Patton, J.G. (2000) Mol. Cell. Biol. 20:3049-3057).

Heterogeneous nuclear ribonucleoproteins (hnRNPs) have been identified that have roles in splicing, exporting of the mature RNAs to the cytoplasm, and mRNA translation (Biamonti, G. et al. (1998) Clin. Exp. Rheumatol. 16:317-326). Some examples of hnRNPs include the yeast proteins Hrp1p, involved in cleavage and polyadenylation at the 3'end of the RNA; Cbp80p, involved in capping the 5'end of the RNA; and Npl3p, a homolog of mammalian hnRNP A1, involved in export of mRNA from the nucleus (Shen, E.C. et al. (1998) Genes Dev. 12:679-691). HnRNPs have been shown to be important targets of the autoimmune response in rheumatic diseases (Biamonti, supra).

Many snRNP and hnRNP proteins are characterized by an RNA recognition motif (RRM). (Reviewed in Birney, E. et al. (1993) Nucleic Acids Res. 21:5803-5816.) The RRM is about 80 amino acids in length and forms four β -strands and two α -helices arranged in an α / β sandwich. The RRM contains a core RNP-1 octapeptide motif along with surrounding conserved sequences. In addition to snRNP proteins, examples of RNA-binding proteins which contain the above motifs include heteronuclear ribonucleoproteins which stabilize nascent RNA and factors which regulate alternative

splicing. Alternative splicing factors include developmentally regulated proteins, specific examples of which have been identified in lower eukaryotes such as *Drosophila melanogaster* and *Caenorhabditis elegans*. These proteins play key roles in developmental processes such as pattern formation and sex determination, respectively. (See, for example, Hodgkin, J. et al. (1994) Development 120:3681-3689.)

The 3'ends of most eukaryote mRNAs are also posttranscriptionally modified by polyadenylation. Polyadenylation proceeds through two enzymatically distinct steps: (i) the endonucleolytic cleavage of nascent mRNAs at cis-acting polyadenylation signals in the 3'-untranslated (non-coding) region and (ii) the addition of a poly(A) tract to the 5'mRNA fragment. The presence of cis-acting RNA sequences is necessary for both steps. These sequences include 5'-AAUAAA-3' located 10-30 nucleotides upstream of the cleavage site and a less well-conserved GU-or U-rich sequence element located 10-30 nucleotides downstream of the cleavage site. Cleavage stimulation factor (CstF), cleavage factor I (CF I), and cleavage factor II (CF II) are involved in the cleavage reaction while cleavage and polyadenylation specificity factor (CPSF) and poly(A) polymerase (PAP) are necessary for both cleavage and polyadenylation. An additional enzyme, poly(A)-binding protein II (PAB II), promotes poly(A) tract elongation (Rüegsegger, U. et al. (1996) J. Biol. Chem. 271:6107-6113; and references within).

TRANSLATION

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Correct translation of the genetic code depends upon each amino acid forming a linkage with the appropriate transfer RNA (tRNA). The aminoacyl-tRNA synthetases (aaRSs) are essential proteins found in all living organisms. The aaRSs are responsible for the activation and correct attachment of an amino acid with its cognate tRNA, as the first step in protein biosynthesis. Prokaryotic organisms have at least twenty different types of aaRSs, one for each different amino acid, while eukaryotes usually have two aaRSs, a cytosolic form and a mitochondrial form, for each different amino acid. The 20 aaRS enzymes can be divided into two structural classes. Class I enzymes add amino acids to the 2' hydroxyl at the 3' end of tRNAs while Class II enzymes add amino acids to the 3' hydroxyl at the 3' end of tRNAs. Each class is characterized by a distinctive topology of the catalytic domain. Class I enzymes contain a catalytic domain based on the nucleotide-binding Rossman 'fold'. In particular, a consensus tetrapeptide motif is highly conserved (Prosite Document PDOC00161, Aminoacyl-transfer RNA synthetases class-I signature). Class I enzymes are specific for arginine, cysteine, glutamic acid, glutamine, isoleucine, leucine, methionine, tyrosine, tryptophan, and valine. Class II enzymes contain a central catalytic domain, which consists of a seven-stranded antiparallel \(\textit{B} \) sheet domain, as well as N- and C- terminal regulatory domains. Class II enzymes are

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separated into two groups based on the heterodimeric or homodimeric structure of the enzyme; the latter group is further subdivided by the structure of the N- and C-terminal regulatory domains (Hartlein, M. and Cusack, S. (1995) J. Mol. Evol. 40:519-530). Class II enzymes are specific for alanine, asparagine, aspartic acid, glycine, histidine, lysine, phenylalanine, proline, serine, and threonine.

Certain aaRSs also have editing functions. IleRS, for example, can misactivate valine to form Val-tRNA^{ne}, but this product is cleared by a hydrolytic activity that destroys the mischarged product. This editing activity is located within a second catalytic site found in the connective polypeptide 1 region (CP1), a long insertion sequence within the Rossman fold domain of Class I enzymes (Schimmel, P. et al. (1998) FASEB J. 12:1599-1609). AaRSs also play a role in tRNA processing. It has been shown that mature tRNAs are charged with their respective amino acids in the nucleus before export to the cytoplasm, and charging may serve as a quality control mechanism to insure the tRNAs are functional (Martinis, S.A. et al. (1999) EMBO J. 18:4591-4596).

Under optimal conditions, polypeptide synthesis proceeds at a rate of approximately 40 amino acid residues per second. The rate of misincorporation during translation in on the order of 10⁻⁴ and is primarily the result of aminoacyl-t-RNAs being charged with the incorrect amino acid. Incorrectly charged tRNA are toxic to cells as they result in the incorporation of incorrect amino acid residues into an elongating polypeptide. The rate of translation is presumed to be a compromise between the optimal rate of elongation and the need for translational fidelity. Mathematical calculations predict that 10⁴ is indeed the maximum acceptable error rate for protein synthesis in a biological system (reviewed in Stryer, supra; and Watson, J. et al. (1987) The Benjamin/Cummings Publishing Co., Inc. Menlo Park, CA). A particularly error prone aminoacyl-tRNA charging event is the charging of tRNA^{Gh} with Gln. A mechanism exits for the correction of this mischarging event which likely has its origins in evolution. Gln was among the last of the 20 naturally occurring amino acids used in polypeptide synthesis to appear in nature. Gram positive eubacteria, cyanobacteria, Archeae, and eukaryotic organelles possess a noncanonical pathway for the synthesis of Gln-tRNA^{Gln} based on the transformation of Glu-tRNA^{Gln} (synthesized by Glu-tRNA synthetase, GluRS) using the enzyme GlutRNA^{Gh} amidotransferase (Glu-AdT). The reactions involved in the transamidation pathway are as follows (Curnow, A.W. et al. (1997) Nucleic Acids Symposium 36:2-4):

30 GluRS $tRNA^{Gln} + Glu + ATP \rightarrow Glu - tRNA^{Gln} + AMP + PP;$

Glu-AdT

 $Glu-tRNA^{Gln} + Gln + ATP \rightarrow Gln-tRNA^{Gln} + Glu + ADP + P$

A similar enzyme, Asp-tRNA^{Asn} amidotransferase, exists in Archaea, which transforms Asp-tRNA^{Asn} to Asn-tRNA^{Asn}. Formylase, the enzyme that transforms Met-tRNA^{fMet} to fMet-tRNA^{fMet} in eubacteria, is likely to be a related enzyme. A hydrolytic activity has also been identified that destroys mischarged Val-tRNA^{fle} (Schimmel, P. et al. (1998) FASEB J. 12:1599-1609). One likely scenario for the evolution of Glu-AdT in primitive life forms is the absence of a specific glutaminyl-tRNA synthetase (GlnRS), requiring an alternative pathway for the synthesis of Gln-tRNA^{Gln}. In fact, deletion of the Glu-AdT operon in Gram positive bacteria is lethal (Curnow, A.W. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11819-11826). The existence of GluRS activity in other organisms has been inferred by the high degree of conservation in translation machinery in nature; however, GluRS has not been identified in all organisms, including *Homo sapiens*. Such an enzyme would be responsible for ensuring translational fidelity and reducing the synthesis of defective polypeptides.

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In addition to their function in protein synthesis, specific aminoacyl tRNA synthetases also play roles in cellular fidelity, RNA splicing, RNA trafficking, apoptosis, and transcriptional and translational regulation. For example, human tyrosyl-tRNA synthetase can be proteolytically cleaved into two fragments with distinct cytokine activities. The carboxy-terminal domain exhibits monocyte and leukocyte chemotaxis activity as well as stimulating production of myeloperoxidase, tumor necrosis factor-α, and tissue factor. The N-terminal domain binds to the interleukin-8 type A receptor and functions as an interleukin-8-like cytokine. Human tyrosyl-tRNA synthetase is secreted from apoptotic tumor cells and may accelerate apoptosis (Wakasugi, K., and Schimmel, P. (1999) Science 284:147-151). Mitochondrial Neurospora crassa TyrRS and S. cerevisiae LeuRS are essential factors for certain group I intron splicing activities, and human mitochondrial LeuRS can substitute for the yeast LeuRS in a yeast null strain. Certain bacterial aaRSs are involved in regulating their own transcription or translation (Martinis, supra). Several aaRSs are able to synthesize diadenosine oligophosphates, a class of signalling molecules with roles in cell proliferation, differentiation, and apoptosis (Kisselev, L.L et al. (1998) FEBS Lett. 427:157-163; Vartanian, A. et al. (1999) FEBS Lett. 456:175-180).

Autoantibodies against aminoacyl-tRNAs are generated by patients with autoimmune diseases such as rheumatic arthritis, dermatomyositis and polymyositis, and correlate strongly with complicating interstitial lung disease (ILD) (Freist, W. et al. (1999) Biol. Chem. 380:623-646; Freist, W. et al. (1996) Biol. Chem. Hoppe Seyler 377:343-356). These antibodies appear to be generated in response to viral infection, and coxsackie virus has been used to induce experimental viral myositis in animals.

Comparison of aaRS structures between humans and pathogens has been useful in the design

of novel antibiotics (Schimmel, *supra*). Genetically engineered aaRSs have been utilized to allow site-specific incorporation of unnatural amino acids into proteins *in vivo* (Liu, D.R. et al. (1997) Proc. Natl. Acad. Sci. USA 94:10092-10097).

tRNA Modifications

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The modified ribonucleoside, pseudouridine (Ψ) , is present ubiquitously in the anticodon regions of transfer RNAs (tRNAs), large and small ribosomal RNAs (rRNAs), and small nuclear RNAs (snRNAs), y is the most common of the modified nucleosides (i.e., other than G, A, U, and C) present in tRNAs. Only a few yeast tRNAs that are not involved in protein synthesis do not contain \(\psi \) (Cortese, R. et al. (1974) J. Biol. Chem. 249:1103-1108). The enzyme responsible for the conversion of uridine to w, pseudouridine synthase (pseudouridylate synthase), was first isolated from Salmonella typhimurium (Arena, F. et al. (1978) Nucleic Acids Res. 5:4523-4536). The enzyme has since been isolated from a number of mammals, including steer and mice (Green, C.J. et al. (1982) J. Biol. Chem. 257:3045-52; and Chen, J. and Patton, J.R. (1999) RNA 5:409-419). tRNA pseudouridine synthases have been the most extensively studied members of the family. They require a thiol donor (e.g., cysteine) and a monovalent cation (e.g., ammonia or potassium) for optimal activity. Additional cofactors or high energy molecules (e.g., ATP or GTP) are not required (Green, supra). Other eukaryotic pseudouridine synthases have been identified that appear to be specific for rRNA (reviewed in Smith, C.M. and Steitz, J.A. (1997) Cell 89:669-672) and a dual-specificity enzyme has been identified that uses both tRNA and rRNA substrates (Wrzesinski, J. et al. (1995) RNA 1: 437-448). The absence of w in the anticodon loop of tRNAs results in reduced growth in both bacteria (Singer, C.E. et al. (1972) Nature New Biol. 238:72-74) and yeast (Lecointe, F. (1998) J. Biol. Chem. 273:1316-1323), although the genetic defect is not lethal.

Another ribonucleoside modification that occurs primarily in eukaryotic cells is the conversion of guanosine to N²,N²-dimethylguanosine (m²₂G) at position 26 or 10 at the base of the D-stem of cytosolic and mitochondrial tRNAs. This posttranscriptional modification is believed to stabilize tRNA structure by preventing the formation of alternative tRNA secondary and tertiary structures. Yeast tRNA^{Asp} is unusual in that it does not contain this modification. The modification does not occur in eubacteria, presumably because the structure of tRNAs in these cells and organelles is sequence constrained and does not require posttranscriptional modification to prevent the formation of alternative structures (Steinberg, S. and Cedergren, R. (1995) RNA 1:886-891, and references within). The enzyme responsible for the conversion of guanosine to m²₂G is a 63 kDa S-adenosylmethionine (SAM)-dependent tRNA N²,N²-dimethyl-guanosine methyltransferase (also referred to as the TRM1 gene product and herein referred to as TRM) (Edqvist, J. (1995) Biochimie 77:54-61). The enzyme

localizes to both the nucleus and the mitochondria (Li, J-M. et al. (1989) J. Cell Biol. 109:1411-1419). Based on studies with TRM from Xenopus laevis, there appears to be a requirement for base pairing at positions C11-G24 and G10-C25 immediately preceding the G26 to be modified, with other structural features of the tRNA also being required for the proper presentation of the G26 substrate (Edqvist. J. et al. (1992) Nucleic Acids Res. 20:6575-6581). Studies in yeast suggest that cells carrying a weak ochre tRNA suppressor (sup3-i) are unable to suppress translation termination in the absence of TRM activity, suggesting a role for TRM in modifying the frequency of suppression in eukaryotic cells (Niederberger, C. et al. (1999) FEBS Lett. 464:67-70), in addition to the more general function of ensuring the proper three-dimensional structures for tRNA.

10 Translation Initiation

Initiation of translation can be divided into three stages. The first stage brings an initiator transfer RNA (Met-tRNA_f) together with the 40S ribosomal subunit to form the 43S preinitiation complex. The second stage binds the 43S preinitiation complex to the mRNA, followed by migration of the complex to the correct AUG initiation codon. The third stage brings the 60S ribosomal subunit to the 40S subunit to generate an 80S ribosome at the initiation codon. Regulation of translation primarily involves the first and second stage in the initiation process (V.M. Pain (1996) Eur. J. Biochem. 236:747-771).

Several initiation factors, many of which contain multiple subunits, are involved in bringing an initiator tRNA and the 40S ribosomal subunit together. eIF2, a guanine nucleotide binding protein, recruits the initiator tRNA to the 40S ribosomal subunit. Only when eIF2 is bound to GTP does it associate with the initiator tRNA. eIF2B, a guanine nucleotide exchange protein, is responsible for converting eIF2 from the GDP-bound inactive form to the GTP-bound active form. Two other factors, eIF1A and eIF3 bind and stabilize the 40S subunit by interacting with the 18S ribosomal RNA and specific ribosomal structural proteins. eIF3 is also involved in association of the 40S ribosomal subunit with mRNA. The Met-tRNA_p eIF1A, eIF3, and 40S ribosomal subunit together make up the 43S preinitiation complex (Pain, supra).

Additional factors are required for binding of the 43S preinitiation complex to an mRNA molecule, and the process is regulated at several levels. eIF4F is a complex consisting of three proteins: eIF4E, eIF4A, and eIF4G. eIF4E recognizes and binds to the mRNA 5'-terminal m'GTP cap, eIF4A is a bidirectional RNA-dependent helicase, and eIF4G is a scaffolding polypeptide. eIF4G has three binding domains. The N-terminal third of eIF4G interacts with eIF4E, the central third interacts with eIF4A, and the C-terminal third interacts with eIF3 bound to the 43S preinitiation complex. Thus, eIF4G acts as a bridge between the 40S ribosomal subunit and the mRNA (M.W.

Hentze (1997) Science 275:500-501).

The ability of eIF4F to initiate binding of the 43S preinitiation complex is regulated by structural features of the mRNA. The mRNA molecule has an untranslated region (UTR) between the 5' cap and the AUG start codon. In some mRNAs this region forms secondary structures that impede binding of the 43S preinitiation complex. The helicase activity of eIF4A is thought to function in removing this secondary structure to facilitate binding of the 43S preinitiation complex (Pain, supra).

Translation Elongation

Elongation is the process whereby additional amino acids are joined to the initiator methionine to form the complete polypeptide chain. The elongation factors EF1 α , EF1 β γ , and EF2 are involved in elongating the polypeptide chain following initiation. EF1 α is a GTP-binding protein. In EF1 α 's GTP-bound form, it brings an aminoacyl-tRNA to the ribosome's A site. The amino acid attached to the newly arrived aminoacyl-tRNA forms a peptide bond with the initiatior methionine. The GTP on EF1 α is hydrolyzed to GDP, and EF1 α -GDP dissociates from the ribosome. EF1 β γ binds EF1 α -GDP and induces the dissociation of GDP from EF1 α , allowing EF1 α to bind GTP and a new cycle to begin.

As subsequent aminoacyl-tRNAs are brought to the ribosome, EF-G, another GTP-binding protein, catalyzes the translocation of tRNAs from the A site to the P site and finally to the E site of the ribosome. This allows the ribosome and the mRNA to remain attached during translation.

20 Translation Termination

The release factor eRF carries out termination of translation. eRF recognizes stop codons in the mRNA, leading to the release of the polypeptide chain from the ribosome.

Expression profiling

Microarrays are analytical tools used in bioanalysis. A microarray has a plurality of molecules spatially distributed over, and stably associated with, the surface of a solid support. Microarrays of polypeptides, polynucleotides, and/or antibodies have been developed and find use in a variety of applications, such as gene sequencing, monitoring gene expression, gene mapping, bacterial identification, drug discovery, and combinatorial chemistry.

One area in particular in which microarrays find use is in gene expression analysis. Array technology can provide a simple way to explore the expression of a single polymorphic gene or the expression profile of a large number of related or unrelated genes. When the expression of a single gene is examined, arrays are employed to detect the expression of a specific gene or its variants. When an expression profile is examined, arrays provide a platform for identifying genes that are tissue

specific, are affected by a substance being tested in a toxicology assay, are part of a signaling cascade, carry out housekeeping functions, or are specifically related to a particular genetic predisposition, condition, disease, or disorder.

RNA Expression

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Array technology can provide a simple way to explore the expression of a single polymorphic gene or the expression profile of a large number of related or unrelated genes. When the expression of a single gene is examined, arrays are employed to detect the expression of a specific gene or its variants. When an expression profile is examined, arrays provide a platform for examining which genes are tissue specific, carrying out housekeeping functions, parts of a signaling cascade, or specifically related to a particular genetic predisposition, condition, disease, or disorder.

The potential application of gene expression profiling is particularly relevant to measuring the toxic response to potential therapeutic compounds and of the metabolic response to therapeutic agents. Diseases treated with steroids and disorders caused by the metabolic response to treatment with steroids include adenomatosis, cholestasis, cirrhosis, hemangioma, Henoch-Schonlein purpura, hepatitis, hepatocellular and metastatic carcinomas, idiopathic thrombocytopenic purpura, porphyria, sarcoidosis, and Wilson disease. Response may be measured by comparing both the levels and sequences expressed in tissues from subjects exposed to or treated with steroid compounds such as mifepristone, progesterone, beclomethasone, medroxyprogesterone, budesonide, prednisone, dexamethasone, betamethasone, or danazol with the levels and sequences expressed in normal untreated tissue.

Steroids are a class of lipid-soluble molecules, including cholesterol, bile acids, vitamin D, and hormones, that share a common four-ring structure based on cyclopentanoperhydrophenanthrene and that carrry out a wide variety of functions. Cholesterol, for example, is a component of cell membranes that controls membrane fluidity. It is also a precursor for bile acids which solubilize lipids and facilitate absorption in the small intestine during digestion. Vitamin D regulates the absorption of calcium in the small intestine and controls the concentration of calcium in plasma. Steroid hormones, produced by the adrenal cortex, ovaries, and testes, include glucocorticoids, mineralocorticoids, androgens, and estrogens. They control various biological processes by binding to intracellular receptors that regulate transcription of specific genes in the nucleus. Glucocorticoids, for example, increase blood glucose concentrations by regulation of gluconeogenesis in the liver, increase blood concentrations of fatty acids by promoting lipolysis in adipose tissues, modulate sensitivity to catcholamines in the central nervous system, and reduce inflammation. The principal

mineralocorticoid, aldosterone, is produced by the adrenal cortex and acts on cells of the distal tubules of the kidney to enhance sodium ion reabsorption. Androgens, produced by the interstitial cells of Leydig in the testis, include the male sex hormone testosterone, which triggers changes at puberty, the production of sperm and maintenance of secondary sexual characteristics. Female sex hormones, estrogen and progesterone, are produced by the ovaries and also by the placenta and adrenal cortex of the fetus during pregnancy. Estrogen regulates female reproductive processes and secondary sexual characteristics. Progesterone regulates changes in the endometrium during the menstrual cycle and pregnancy.

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Steroid hormones are widely used for fertility control and in anti-inflammatory treatments for physical injuries and diseases such as arthritis, asthma, and auto-immune disorders. Progesterone, a naturally occurring progestin, is primarily used to treat amenorrhea, abnormal uterine bleeding, or as a contraceptive. Endogenous progesterone is responsible for inducing secretory activity in the endometrium of the estrogen-primed uterus in preparation for the implantation of a fertilized egg and for the maintenance of pregnancy. It is secreted from the corpus luteum in response to luteinizing hormone (LH). The primary contraceptive effect of exogenous progestins involves the suppression of the midcycle surge of LH. At the cellular level, progestins diffuse freely into target cells and bind to the progesterone receptor. Target cells include the female reproductive tract, the mammary gland, the hypothalamus, and the pituitary. Once bound to the receptor, progestins slow the frequency of release of gonadotropin releasing hormone from the hypothalamus and blunt the pre-ovulatory LH surge, thereby preventing follicular maturation and ovulation. Progesterone has minimal estrogenic and androgenic activity. Progesterone is metabolized hepatically to pregnanediol and conjugated with glucuronic acid.

Medroxyprogesterone (MAH), also known as 6α-methyl-17-hydroxyprogesterone, is a synthetic progestin with a pharmacological activity about 15 times greater than progesterone. MAH is used for the treatment of renal and endometrial carcinomas, amenorrhea, abnormal uterine bleeding, and endometriosis associated with hormonal imbalance. MAH has a stimulatory effect on respiratory centers and has been used in cases of low blood oxygenation caused by sleep apnea, chronic obstructive pulmonary disease, or hypercapnia.

Mifepristone, also known as RU-486, is an antiprogesterone drug that blocks receptors of progesterone. It counteracts the effects of progesterone, which is needed to sustain pregnancy. Mifepristone induces spontaneous abortion when administered in early pregnancy followed by treatment with the prostaglandin, misoprostol. Further, studies show that mifepristone at a

substantially lower dose can be highly effective as a postcoital contraceptive when administered within five days after unprotected intercourse, thus providing women with a "morning-after pill" in case of contraceptive failure or sexual assault. Mifepristone also has potential uses in the treatment of breast and ovarian cancers in cases in which tumors are progesterone-dependent. It interferes with steroid-dependent growth of brain meningiomas, and may be useful in treatment of endometriosis where it blocks the estrogen-dependent growth of endometrial tissues. It may also be useful in treatment of uterine fibroid tumors and Cushing's Syndrome. Mifepristone binds to glucocorticoid receptors and interferes with cortisol binding. Mifepristone also may act as an anti-glucocorticoid and be effective for treating conditions where cortisol levels are elevated such as AIDS, anorexia nervosa, ulcers, diabetes, Parkinson's disease, multiple sclerosis, and Alzheimer's disease.

Danazol is a synthetic steroid derived from ethinyl testosterone. Danazol indirectly reduces estrogen production by lowering pituitary synthesis of follicle-stimulating hormone and LH. Danazol also binds to sex hormone receptors in target tissues, thereby exhibiting anabolic, antiestrognic, and weakly androgenic activity. Danazol does not possess any progestogenic activity, and does not suppress normal pituitary release of corticotropin or release of cortisol by the adrenal glands. Danazol is used in the treatment of endometriosis to relieve pain and inhibit endometrial cell growth. It is also used to treat fibrocystic breast disease and hereditary angioedema.

Corticosteroids are used to relieve inflammation and to suppress the immune response. They inhibit eosinophil, basophil, and airway epithelial cell function by regulation of cytokines that mediate the inflammatory response. They inhibit leukocyte infiltration at the site of inflammation, interfere in the function of mediators of the inflammatory response, and suppress the humoral immune response. Corticosteroids are used to treat allergies, asthma, arthritis, and skin conditions. Beclomethasone is a synthetic glucocorticoid that is used to treat steroid-dependent asthma, to relieve symptoms associated with allergic or nonallergic (vasomotor) rhinitis, or to prevent recurrent nasal polyps following surgical removal. The anti-inflammatory and vasoconstrictive effects of intranasal beclomethasone are 5000 times greater than those produced by hydrocortisone. Budesonide is a corticosteroid used to control symptoms associated with allergic rhinitis or asthma. Budesonide has high topical anti-inflammatory activity but low systemic activity. Dexamethasone is a synthetic glucocorticoid used in anti-inflammatory or immunosuppressive compositions. It is also used in inhalants to prevent symptoms of asthma. Due to its greater ability to reach the central nervous system, dexamethasone is usually the treatment of choice to control cerebral edema. Dexamethasone is approximately 20-30 times more potent than hydrocortisone and 5-7 times more potent than prednisone. Prednisone is metabolized in

the liver to its active form, prednisolone, a glucocorticoid with anti-inflammatory properties. Prednisone is approximately 4 times more potent than hydrocortisone and the duration of action of prednisone is intermediate between hydrocortisone and dexamethasone. Prednisone is used to treat allograft rejection, asthma, systemic lupus erythematosus, arthritis, ulcerative colitis, and other inflammatory conditions. Betamethasone is a synthetic glucocorticoid with antiinflammatory and immunosuppressive activity and is used to treat psoriasis and fungal infections, such as athlete's foot and ringworm.

The anti-inflammatory actions of corticosteroids are thought to involve phospholipase A₂ inhibitory proteins, collectively called lipocortins. Lipocortins, in turn, control the biosynthesis of potent mediators of inflammation such as prostaglandins and leukotrienes by inhibiting the release of the precursor molecule arachidonic acid. Proposed mechanisms of action include decreased IgE synthesis, increased number of β-adrenergic receptors on leukocytes, and decreased arachidonic acid metabolism. During an immediate allergic reaction, such as in chronic bronchial asthma, allergens bridge the IgE antibodies on the surface of mast cells, which triggers these cells to release chemotactic substances. Mast cell influx and activation, therefore, is partially responsible for the inflammation and hyperirritability of the oral mucosa in asthmatic patients. This inflammation can be retarded by administration of corticosteroids.

CA3+ Steroids

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dexamethasone, betamethasone, or danazol with the levels and sequences expressed in normal untreated tissue.

The effects upon liver metabolism and hormone clearance mechanisms are important to understand the pharmacodynamics of a drug. The human C3A cell line is a clonal derivative of

HepG2/C3 (hepatoma cell line, isolated from a 15-year-old male with liver tumor), which was selected for strong contact inhibition of growth. The use of a clonal population enhances the reproducibility of the cells. C3A cells have many characteristics of primary human hepatocytes in culture: i) expression of insulin receptor and insulin-like growth factor II receptor; ii) secretion of a high ratio of serum albumin compared with α-fetoprotein iii) conversion of ammonia to urea and glutamine; iv)

metabolize aromatic amino acids; and v) proliferate in glucose-free and insulin-free medium. The C3A cell line is now well established as an in vitro model of the mature human liver (Mickelson et al. (1995) Hepatology 22:866-875; Nagendra et al. (1997) Am J Physiol 272:G408-G416).

Vascular endothelial activation

Activation of vascular endothelium is a central event in a wide range of physiological and disease processes such as vascular tone regulation, coagulation and thrombosis, atherosclerosis, inflammation and some infectious diseases. Blood vessel walls are composed of two tissue layers: an endothelial cell (EC) layer which comprises the lumenal surface of the vessel, and an underlying vascular smooth muscle cell (VSMC) layer. Through dynamic interactions with each other and with surrounding tissues, the vascular endothelium and smooth muscle tissues maintain vascular tone, control selective permeability of the vascular wall, direct vessel remodeling and angiogenesis, and modulate inflammatory and immune responses.

The inflammatory response is a complex vascular reaction mediated by numerous cytokines, chemokines, growth factors, and other signaling molecules expressed by activated ECs, VSMCs and leukocytes. Inflammation protects the organism during trauma and infection, but can also lead to pathological conditions such as atherosclerosis.

Human coronary artery endothelial cells (HCAECs) are primary cells derived from the endothelium of a human coronary artery. HCAECs are used as an experimental model for investigating the role of the endothelium in human vascular biology in vitro. Human umbilical artery endothelial cells (HUAECs) are primary cells derived from the endothelium of an umbilical artery. Human uterine myometrium microvascular endothelial cells (UtMVECs) are primary cells derived from the uterine myometrium microvasculature. Human Iliac Artery Endothelial Cells (HIAECs) are primary cells derived from the endothelium of an iliac artery. Human umbilical vein endothelial cells

(HUVECs) are a primary cell line derived from the endothe-lium of the human umbilical vein. ECV304 is a human endothelial line.

Breast cancer

There are more than 180,000 new cases of breast cancer diagnosed each year, and the

mortality rate for breast cancer approaches 10% of all deaths in females between the ages of 45-54

(Gish, K. (1999) AWIS Magazine 28:7-10). However the survival rate based on early diagnosis of localized breast cancer is extremely high (97%), compared with the advanced stage of the disease in which the tumor has spread beyond the breast (22%). Current procedures for clinical breast examination are lacking in sensitivity and specificity, and efforts are underway to develop comprehensive gene expression profiles for breast cancer that may be used in conjunction with conventional screening methods to improve diagnosis and prognosis of this disease (Perou, C.M. et al. (2000) Nature 406:747-752). Mutations in two genes, BRCA1 and BRCA2, are known to greatly predispose a woman to breast cancer and may be passed on from parents to children (Gish, supra). However, this type of hereditary breast cancer accounts for only about 5% to 9% of breast cancers, while the vast majority of breast cancer is due to non-inherited mutations that occur in breast epithelial cells.

Cell lines derived from human mammary epithelial cells at various stages of breast cancer provide a useful model to study the process of malignant transformation and tumor progression as it has been shown that these cell lines retain many of the properties of their parental tumors for lengthy culture periods (Wistuba, I.I. et al. (1998) Clin. Cancer Res. 4:2931-2938). Such a model is particularly useful for comparing phenotypic and molecular characteristics of human mammary epithelial cells at various stages of malignant transformation.

Estragen stimulation plays a critical role in the development of normal mammary epithelium. Estradiol has a direct mitogenic effect on breast cancer cells, causing them to divide more rapidly by shortening their cell cycle. Also, estradiol induces a large number of enzymes and other proteins involved in nucleic acid synthesis in isolated breast cancer cell lines. Estradiol may increase the expression of the EGF receptor in response to TGF-α and EGF. In addition, estrogens may promote proliferation of tumor cells by inducing the synthesis of TGF-α and EGF, and may block growth factors that would normally inhibit tumor cell growth. Estrogen receptor (ER) has been investigated extensively as a prognostic marker in breast cancer. Patients whose tumors display high levels of estrogen receptor have a significantly better prognosis than patients with receptor-negative tumors. When ER is lost or cells expressing the ER are selected against by therapeutic treatments, the tumor

becomes more aggressive.

Lung cancer

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Lung cancer is the leading cause of cancer death for men and the second leading cause of cancer death for women in the U.S. Lung cancers are divided into four histopathologically distinct groups. Three groups (squamous cell carcinoma, adenocarcinoma, and large cell carcinoma) are classified as non-small cell lung cancers (NSCLCs). The fourth group of cancers is referred to as small cell lung cancer (SCLC). Deletions on chromosome 3 are common in this disease and are thought to indicate the presence of a tumor suppressor gene in this region. Activating mutations in K-ras are commonly found in lung cancer and are the basis of one of the mouse models for the disease.

10 Ovarian Cancer

Ovarian cancer is the leading cause of death from a gynecologic cancer. The majority of ovarian cancers are derived from epithelial cells, and 70% of patients with epithelial ovarian cancers present with late-stage disease. As a result, the long-term survival rates for this disease is very low. Identification of early-stage markers for ovarian cancer would significantly increase the survival rate. The molecular events that lead to ovarian cancer are poorly understood. Some of the known aberrations include mutation of p53 and microsatellite instability. Since gene expression patterns are likely to vary when normal ovary is compared to ovarian tumors, examination of gene expression in these tissues to identify possible markers for ovarian cancer is particularly relevant to improving

20 Colon Cancer

diagnosis, prognosis, and treatment of this disease.

Colorectal cancer is the second leading cause of cancer deaths in the United States. Colon cancer is associated with aging, since 90% of the total cases occur in individuals over the age of 55. A widely accepted hypothesis is that several contributing genetic mutations must accumulate over time in an individual who develops the disease. To understand the nature of genetic alterations in colorectal cancer, a number of studies have focused on the inherited syndromes. The first known inherited syndrome, Familial Adenomatous Polyposis (FAP), is caused by mutations in the Adenomatous Polyposis Coli gene (APC), resulting in truncated or inactive forms of the protein. This tumor suppressor gene has been mapped to chromosome 5q. The second known inherited syndrome is hereditary nonpolyposis colorectal cancer (HNPCC), which is caused by mutations in mismatch repair genes.

Although hereditary colon cancer syndromes occur in a small percentage of the population and most colorectal cancers are considered sporadic, knowledge from studies of the hereditary

syndromes can be generally applied. For instance, somatic mutations in APC occur in at least 80% of indiscriminate colon tumors. APC mutations are thought to be the initiating event in the disease. Other mutations occur subsequently. Approximately 50% of colorectal cancers contain activating mutations in ras, while 85% contain inactivating mutations in p53. Changes in these genes lead to gene expression changes in colon cancer. Less is understood about downstream targets of these mutations and the role they may play in cancer development and progression.

Inflammatory/Immune Response

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Blood vessel walls are composed of two tissue layers: an endothelial cell (EC) layer which comprises the lumenal surface of the vessel, and an underlying vascular smooth muscle cell (VSMC) layer. Through dynamic interactions with each other and with surrounding tissues, the vascular endothelium and smooth muscle tissues maintain vascular tone, control selective permeability of the vascular wall, direct vessel remodeling and angiogenesis, and modulate inflammatory and immune responses.

The inflammatory response is a complex vascular reaction mediated by numerous cytokines, chemokines, growth factors, and other signaling molecules expressed by activated ECs, VSMCs and leukocytes. Inflammation protects the organism during trauma and infection, but can also lead to pathological conditions such as atherosclerosis. The pro-inflammatory cytokines, interleukin (IL)-1 and tumor necrosis factor (TNF), are secreted by a small number of activated macrophages or other cells and can set off a cascade of vascular changes, largely through their ability to alter gene expression patterns in ECs and VSMCs. These vascular changes include vasodilation and increased permeability of microvasculature, edema, and leukocyte extravasation and transmigration across the vessel wall. Ultimately, leukocytes, particularly neutrophils and monocytes/macrophages, accumulate in the extravascular space, where they remove injurious agents by phagocytosis and oxidative killing, a process accompanied by release of toxic factors, such as proteases and reactive oxygen species.

Tumor necrosis factor-α (TNF-α) is a pleiotropic factor that exerts a variety of effects, such as growth promotion, growth inhibition, angiogenesis, cytotoxicity, inflammation, and immunomodulation. This cytokine is synthesized mainly by macrophages in response to invasive stimuli as an active 26 kDa membrane-bound precursor that is cleaved proteolytically to a mature 17 kDa form with the prosequence polypeptide remaining associated to the membrane. The peptide is as bioactive as a 51 kDa trimer, which can be recognized by TNF-α receptors. TNF-α receptors are present in the majority of cell types. IL-1 and TNF induce pro-inflammatory, thrombotic, and antiapoptotic changes in gene expression by signaling through receptors on the surface of ECs and

VSMCs; these receptors activate transcription factors such as NFkB as well as AP-1, IRF-1, and NF-GMa, leading to alterations in gene expression. Genes known to be differentially regulated in EC by IL-1 and TNF include E selectin, VCAM-1, ICAM-1, PAF, IkBα, IAP-1, MCP-1, eotaxin, ENA-78, G-CSF, A20, ICE, and complement C3 component. A key event in inflammation, adhesion and transmigration of blood leukocytes across the vascular endothelium, for example, is mediated by increased expression of E selectin, P selectin, ICAM-1, and VCAM-1 on activated endothelium.

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Several investigators have examined changes in vascular cell gene expression associated with various inflammatory diseases or model systems. Examining human umbilical vein endothelial cells (HUVEC) activated by recombinant TNFα or conditioned medium from activated human primary monocytes, Horrevoets et al. (1999; Blood 93:3418-3431) identified 106 differentially regulated genes. In a similar approach, de Vries et al. (2000; JBC 275:23939-23947) identified 40 differentially regulated genes in umbilical cord artery-derived smooth muscle cells activated by conditioned media from cultured macrophages after stimulation with oxidized LDL particles. In both studies, many of the identified genes were already known to be involved in inflammation. Comparing expression profiles from inflammatory diseased tissues, cultured macrophages, chondrocyte cell lines, primary chondrocytes, and synoviocytes, Heller et al. (1997; Proc Natl Acad Sci USA 94:2150-2155) identified candidate genes involved in inflammatory responses, including TNF, IL-1 IL-6, IL-8 G-CSF, RANTES, and V-CAM. From this candidate gene set, tissue inhibitor of metalloproteinase 1, ferritin light chain, and manganese superoxide dismutase were found to be differentially expressed in rheumatoid arthritis (RA) relative to inflammatory bowel disease (IBD). Further, IL-3, chemokine Grog, and metalloproteinase matrix metallo-elastase were expressed in both RA and IBD. Most recently, in an analysis of cultured aortic smooth muscle cells treated with TNFa, Haley et al. (2000; Circulation 102:2185-2189) found a 20-fold increase in eotaxin, an eosinophil chemotactic factor. The overexpression of eotaxin and its receptor CCR3 in atherosclerotic lesions was confirmed by northern analysis.

Array technology can provide a simple way to explore the expression of a single polymorphic gene or the expression profile of a large number of related or unrelated genes. When the expression of a single gene is examined, arrays are employed to detect the expression of a specific gene or its variants. When an expression profile is examined, arrays provide a platform for identifying genes that are tissue specific, carry out housekeeping functions, are part of a signaling cascade, or are specifically related to a particular genetic predisposition, condition, disease, or disorder. The potential application of gene expression profiling is particularly relevant to improving diagnosis, prognosis, and

treatment of disease. For example, both the levels and sequences expressed in tissues from subjects with coronary artery disease may be compared with the levels and sequences expressed in normal vascular tissue.

DNA-based array technology is also useful as a method of analyzing cell signaling pathways. For example, protein kinase C (PKC) is a family of serine/threonine kinases which play a critical role in many signal transduction pathways in the cell (Kanashiro, C.A. and Khalil, R.A. (1998) Clin. Exp. Pharmacol. Physiol. 25:974-985). Phorbol 12-myristate 13-acetate (PMA) is a broad activator of the protein kinase C-dependent pathways. PMA promotes tumors in cells by over activating PKC pathways. PKC is also affected by intracellular calcium levels. Internal calcium flux has been shown have many effects on cells, particular in the areas of cell activation and proliferation (Cole, K, and Kohn, E. (1994) Cancer Metastasis Rev. 13:31-44). Ionomycin is a calcium ionophore that permits the entry of calcium into the cell, hence increasing the cytosolic calcium concentration. Thus the combination of PMA and ionomycin activates two of the major signaling pathways used by mammalian cells to interact with their environment. In T cells, for example, the combination of PMA and ionomycin mimics the type of secondary signaling events elicited during optimal T cell activation, while in endothelial cells, PMA and ionomycin reproduce the effect of inflammatory mediatiors.

Cell lines are widely used in experimental biology to model human cell behavior. Jurkat, an acute T-cell leukemia cell line that grows actively in the absence of external stimuli, is used to study signaling in human T cells. ECV304, a cell line derived from the endothelium of the human umbilical vein, is used to study the functional biology of human endothelial cells.

Neurological disorders

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Characterization of region-specific gene expression in the human brain provides a context and background for molecular neurobiology on a variety of neurological disorders. For example, Alzheimer's disease (AD) is a progressive, neurodestructive process of the human neocortex, characterized by the deterioration of memory and higher cognitive function. A progressive and irreversible brain disorder, AD is characterized by three major pathogenic episodes involving (a) an aberrant processing and deposition of beta-amyloid precursor protein (betaAPP) to form neurotoxic beta-amyloid (betaA) peptides and an aggregated insoluble polymer of betaA that forms the senile plaque, (b) the establishment of intraneuronal neuritic tau pathology yielding widespread deposits of agyrophilic neurofibrillary tangles (NFT) and (c) the initiation and proliferation of a brain-specific inflammatory response. These three seemingly disperse attributes of AD etiopathogenesis are linked by the fact that proinflammatory microglia, reactive astrocytes and their associated cytokines and

chemokines are associated with the biology of the microtubule associated protein tau, betaA speciation and aggregation. Missense mutations in the presentilin genes PS1 and PS2, implicated in early onset familial AD, cause abnormal betaAPP processing with resultant overproduction of betaA42 and related neurotoxic peptides. Specific betaA fragments such as betaA42 can further potentiate proinflammatory mechanisms. Expression of the inducible oxidoreductase cyclooxygenase-2 and cytosolic phospholipase A2 (cPLA2) are strongly activated during cerebral ischemia and trauma, epilepsy and AD, indicating the induction of proinflammatory gene pathways as a response to brain injury. Neurotoxic metals such as aluminum and zinc, both implicated in AD etiopathogenesis, and arachidonic acid, a major metabolite of brain cPLA2 activity, each polymerize hyperphosphorylated tau to form NFT-like bundles. Studies have identified a reduced risk for AD in patients aged over 70 years who were previously treated with non-steroidal anti-inflammatory drugs for non-CNS afflictions that include arthritis. (For a review of the interrelationships between the mechanisms of PS1, PS2 and betaAPP gene expression, tau and betaA deposition and the induction, regulation and proliferation in AD of the neuroinflammatory response, see Lukiw W.J, and Bazan N.G.(2000) Neurochem. Res. 2000 25:1173-1184).

The discovery of new nucleic acid-associated proteins, and the polynucleotides encoding them, satisfies a need in the art by providing new compositions which are useful in the diagnosis, prevention, and treatment of cell proliferative, neurological, developmental, and autoimmune/inflammatory disorders, and infections, and in the assessment of the effects of exogenous compounds on the expression of nucleic acid and amino acid sequences of nucleic acid-associated proteins.

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There is a need in the art for new compositions, including nucleic acids and proteins, for the diagnosis, prevention, and treatment of cell proliferative, neurological, developmental, and autoimmune/inflammatory disorders, and infections.

SUMMARY OF THE INVENTION

Various embodiments of the invention provide purified polypeptides, nucleic acid-associated proteins, referred to collectively as 'NAAP' and individually as 'NAAP-1,' 'NAAP-2,' 'NAAP-3,' 'NAAP-4,' 'NAAP-5,' 'NAAP-6,' 'NAAP-7,' 'NAAP-8,' 'NAAP-9,' 'NAAP-10,' 'NAAP-11,' 'NAAP-12,' 'NAAP-13,' 'NAAP-14,' 'NAAP-15,' 'NAAP-16,' 'NAAP-17,' 'NAAP-18,' 'NAAP-19,' 'NAAP-20,' 'NAAP-21,' 'NAAP-22,' 'NAAP-23,' 'NAAP-24,' 'NAAP-25,' 'NAAP-26,' 'NAAP-27,' 'NAAP-28,' 'NAAP-29,' 'NAAP-30,' 'NAAP-31,' 'NAAP-32,' 'NAAP-33,' 'NAAP-34,' 'NAAP-35,' 'NAAP-36,' 'NAAP-37,' 'NAAP-38,' 'NAAP-39,' 'NAAP-40,' 'NAAP-41,'

'NAAP-42,' 'NAAP-43,' 'NAAP-44,' 'NAAP-45,' 'NAAP-46,' 'NAAP-47,' 'NAAP-48,' 'NAAP-49,' 'NAAP-50,' 'NAAP-51,' 'NAAP-52,' 'NAAP-53,' 'NAAP-54,' 'NAAP-55,' 'NAAP-56,' and 'NAAP-57' and methods for using these proteins and their encoding polynucleotides for the detection, diagnosis, and treatment of diseases and medical conditions. Embodiments also provide methods for utilizing the purified nucleic acid-associated proteins and/or their encoding polynucleotides for facilitating the drug discovery process, including determination of efficacy, dosage, toxicity, and pharmacology. Related embodiments provide methods for utilizing the purified nucleic acid-associated proteins and/or their encoding polynucleotides for investigating the pathogenesis of diseases and medical conditions.

An embodiment provides an isolated polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. Another embodiment provides an isolated polypeptide comprising an amino acid sequence of SEQ ID NO:1-57.

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Still another embodiment provides an isolated polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. In another embodiment, the polynucleotide encodes a polypeptide selected from the group consisting of SEQ ID NO:1-57. In an alternative embodiment, the polynucleotide is selected from the group consisting of SEQ ID NO:58-114.

Still another embodiment provides a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group

consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. Another embodiment provides a cell transformed with the recombinant polynucleotide. Yet another embodiment provides a transgenic organism comprising the recombinant polynucleotide.

Another embodiment provides a method for producing a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. The method comprises a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide comprising a promoter sequence operably linked to a polynucleotide encoding the polypeptide, and b) recovering the polypeptide so expressed.

Yet another embodiment provides an isolated antibody which specifically binds to a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57.

Still yet another embodiment provides an isolated polynucleotide selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). In other embodiments, the polynucleotide can comprise at least about 20, 30, 40, 60, 80, or 100 contiguous nucleotides.

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Yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and b) detecting the presence or absence of said hybridization complex. In a related embodiment, the method can include detecting the amount of the hybridization complex. In still other embodiments, the probe can comprise at least about 20, 30, 40, 60, 80, or 100 contiguous nucleotides.

Still yet another embodiment provides a method for detecting a target polynucleotide in a sample, said target polynucleotide being selected from the group consisting of a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, b) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, c) a polynucleotide complementary to the polynucleotide of a), d) a polynucleotide complementary to the polynucleotide of b), and e) an RNA equivalent of a)-d). The method comprises a) amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof. In a related embodiment, the method can include detecting the amount of the amplified target polynucleotide or fragment thereof.

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Another embodiment provides a composition comprising an effective amount of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the

group consisting of SEQ ID NO:1-57, and a pharmaceutically acceptable excipient. In one embodiment, the composition can comprise an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. Other embodiments provide a method of treating a disease or condition associated with decreased or abnormal expression of functional NAAP, comprising administering to a patient in need of such treatment the composition.

Yet another embodiment provides a method for screening a compound for effectiveness as an agonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting agonist activity in the sample. Another embodiment provides a composition comprising an agonist compound identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment provides a method of treating a disease or condition associated with decreased expression of functional NAAP, comprising administering to a patient in need of such treatment the composition.

Still yet another embodiment provides a method for screening a compound for effectiveness as an antagonist of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. The method comprises a) exposing a sample comprising the polypeptide to a compound, and b) detecting antagonist activity in the sample. Another embodiment provides a composition comprising an antagonist compound identified by the method and a pharmaceutically acceptable excipient. Yet another embodiment provides a method of treating a disease or condition associated with overexpression of functional NAAP, comprising administering to a patient in need of such treatment the composition.

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Another embodiment provides a method of screening for a compound that specifically binds to

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a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active 5 fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. The method comprises a) combining the polypeptide with at least one test compound under suitable conditions, and b) detecting binding of the polypeptide to the test compound, thereby identifying a compound that specifically binds to the polypeptide.

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Yet another embodiment provides a method of screening for a compound that modulates the activity of a polypeptide selected from the group consisting of a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical or at least about 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, c) a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and d) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57. The method comprises a) combining the polypeptide with at least one test compound under conditions permissive for the activity of the polypeptide, b) assessing the activity of the polypeptide in the presence of the test compound, and c) comparing the activity of the polypeptide in the presence of the test compound with the activity of the polypeptide in the absence of the test compound, wherein a change in the activity of the polypeptide in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide.

Still yet another embodiment provides a method for screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, the method comprising a) exposing a sample comprising the target polynucleotide to a compound, b) detecting altered expression of the target polynucleotide, and c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.

Another embodiment provides a method for assessing toxicity of a test compound, said

method comprising a) treating a biological sample containing nucleic acids with the test compound; b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, iii) a polynucleotide having a sequence complementary to i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Hybridization occurs under conditions whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide selected from the group consisting of i) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, ii) a polynucleotide comprising a naturally occurring polynucleotide sequence at least 90% identical or at least about 90% identical to a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114, iii) a polynucleotide complementary to the polynucleotide of i), iv) a polynucleotide complementary to the polynucleotide of ii), and v) an RNA equivalent of i)-iv). Alternatively, the target polynucleotide can comprise a fragment of a polynucleotide selected from the group consisting of i)-v) above; c) quantifying the amount of hybridization complex; and d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.

BRIEF DESCRIPTION OF THE TABLES

Table 1 summarizes the nomenclature for full length polynucleotide and polypeptide embodiments of the invention.

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Table 2 shows the GenBank identification number and annotation of the nearest GenBank homolog, and the PROTEOME database identification numbers and annotations of PROTEOME database homologs, for polypeptide embodiments of the invention. The probability scores for the matches between each polypeptide and its homolog(s) are also shown.

Table 3 shows structural features of polypeptide embodiments, including predicted motifs and domains, along with the methods, algorithms, and searchable databases used for analysis of the polypeptides.

Table 4 lists the cDNA and/or genomic DNA fragments which were used to assemble

polynucleotide embodiments, along with selected fragments of the polynucleotides.

Table 5 shows representative cDNA libraries for polynucleotide embodiments.

Table 6 provides an appendix which describes the tissues and vectors used for construction of the cDNA libraries shown in Table 5.

Table 7 shows the tools, programs, and algorithms used to analyze polynucleotides and polypeptides, along with applicable descriptions, references, and threshold parameters.

Table 8 shows single nucleotide polymorphisms found in polynucleotide sequences of the invention, along with allele frequencies in different human populations.

DESCRIPTION OF THE INVENTION

Before the present proteins, nucleic acids, and methods are described, it is understood that embodiments of the invention are not limited to the particular machines, instruments, materials, and methods described, as these may vary. It is also to be understood that the terminology used herein is for the purpose of describing particular embodiments only, and is not intended to limit the scope of the invention.

As used herein and in the appended claims, the singular forms "a," "an," and "the" include plural reference unless the context clearly dictates otherwise. Thus, for example, a reference to "a host cell" includes a plurality of such host cells, and a reference to "an antibody" is a reference to one or more antibodies and equivalents thereof known to those skilled in the art, and so forth.

Unless defined otherwise, all technical and scientific terms used herein have the same meanings as commonly understood by one of ordinary skill in the art to which this invention belongs. Although any machines, materials, and methods similar or equivalent to those described herein can be used to practice or test the present invention, the preferred machines, materials and methods are now described. All publications mentioned herein are cited for the purpose of describing and disclosing the cell lines, protocols, reagents and vectors which are reported in the publications and which might be used in connection with various embodiments of the invention. Nothing herein is to be construed as an admission that the invention is not entitled to antedate such disclosure by virtue of prior invention.

DEFINITIONS

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"NAAP" refers to the amino acid sequences of substantially purified NAAP obtained from any species, particularly a mammalian species, including bovine, ovine, porcine, murine, equine, and human, and from any source, whether natural, synthetic, semi-synthetic, or recombinant.

The term "agonist" refers to a molecule which intensifies or mimics the biological activity of

NAAP. Agonists may include proteins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of NAAP either by directly interacting with NAAP or by acting on components of the biological pathway in which NAAP participates.

An "allelic variant" is an alternative form of the gene encoding NAAP. Allelic variants may result from at least one mutation in the nucleic acid sequence and may result in altered mRNAs or in polypeptides whose structure or function may or may not be altered. A gene may have none, one, or many allelic variants of its naturally occurring form. Common mutational changes which give rise to allelic variants are generally ascribed to natural deletions, additions, or substitutions of nucleotides. Each of these types of changes may occur alone, or in combination with the others, one or more times in a given sequence.

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"Altered" nucleic acid sequences encoding NAAP include those sequences with deletions, insertions, or substitutions of different nucleotides, resulting in a polypeptide the same as NAAP or a polypeptide with at least one functional characteristic of NAAP. Included within this definition are polymorphisms which may or may not be readily detectable using a particular oligonucleotide probe of the polynucleotide encoding NAAP, and improper or unexpected hybridization to allelic variants, with a locus other than the normal chromosomal locus for the polynucleotide encoding NAAP. The encoded protein may also be "altered," and may contain deletions, insertions, or substitutions of amino acid residues which produce a silent change and result in a functionally equivalent NAAP. Deliberate amino acid substitutions may be made on the basis of one or more similarities in polarity, charge, solubility, hydrophobicity, hydrophilicity, and/or the amphipathic nature of the residues, as long as the biological or immunological activity of NAAP is retained. For example, negatively charged amino acids may include aspartic acid and glutamic acid, and positively charged amino acids may include lysine and arginine. Amino acids with uncharged polar side chains having similar hydrophilicity values may include: asparagine and glutamine; and serine and threonine. Amino acids with uncharged side chains having similar hydrophilicity values may include: leucine, isoleucine, and valine; glycine and alanine; and phenylalanine and tyrosine.

The terms "amino acid" and "amino acid sequence" can refer to an oligopeptide, a peptide, a polypeptide, or a protein sequence, or a fragment of any of these, and to naturally occurring or synthetic molecules. Where "amino acid sequence" is recited to refer to a sequence of a naturally occurring protein molecule, "amino acid sequence" and like terms are not meant to limit the amino acid sequence to the complete native amino acid sequence associated with the recited protein molecule.

"Amplification" relates to the production of additional copies of a nucleic acid. Amplification

may be carried out using polymerase chain reaction (PCR) technologies or other nucleic acid amplification technologies well known in the art.

The term "antagonist" refers to a molecule which inhibits or attenuates the biological activity of NAAP. Antagonists may include proteins such as antibodies, anticalins, nucleic acids, carbohydrates, small molecules, or any other compound or composition which modulates the activity of NAAP either by directly interacting with NAAP or by acting on components of the biological pathway in which NAAP participates.

The term "antibody" refers to intact immunoglobulin molecules as well as to fragments thereof, such as Fab, F(ab')₂, and Fv fragments, which are capable of binding an epitopic determinant.

Antibodies that bind NAAP polypeptides can be prepared using intact polypeptides or using fragments containing small peptides of interest as the immunizing antigen. The polypeptide or oligopeptide used to immunize an animal (e.g., a mouse, a rat, or a rabbit) can be derived from the translation of RNA, or synthesized chemically, and can be conjugated to a carrier protein if desired. Commonly used carriers that are chemically coupled to peptides include bovine serum albumin, thyroglobulin, and keyhole limpet hemocyanin (KLH). The coupled peptide is then used to immunize the animal.

The term "antigenic determinant" refers to that region of a molecule (i.e., an epitope) that makes contact with a particular antibody. When a protein or a fragment of a protein is used to immunize a host animal, numerous regions of the protein may induce the production of antibodies which bind specifically to antigenic determinants (particular regions or three-dimensional structures on the protein). An antigenic determinant may compete with the intact antigen (i.e., the immunogen used to elicit the immune response) for binding to an antibody.

The term "aptamer" refers to a nucleic acid or oligonucleotide molecule that binds to a specific molecular target. Aptamers are derived from an *in vitro* evolutionary process (e.g., SELEX (Systematic Evolution of Ligands by EXponential Enrichment), described in U.S. Patent No.

5,270,163), which selects for target-specific aptamer sequences from large combinatorial libraries. Aptamer compositions may be double-stranded or single-stranded, and may include deoxyribonucleotides, ribonucleotides, nucleotide derivatives, or other nucleotide-like molecules. The nucleotide components of an aptamer may have modified sugar groups (e.g., the 2'-OH group of a ribonucleotide may be replaced by 2'-F or 2'-NH₂), which may improve a desired property, e.g.,

resistance to nucleases or longer lifetime in blood. Aptamers may be conjugated to other molecules, e.g., a high molecular weight carrier to slow clearance of the aptamer from the circulatory system. Aptamers may be specifically cross-linked to their cognate ligands, e.g., by photo-activation of a

cross-linker (Brody, E.N. and L. Gold (2000) J. Biotechnol. 74:5-13).

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The term "intramer" refers to an aptamer which is expressed in vivo. For example, a vaccinia virus-based RNA expression system has been used to express specific RNA aptamers at high levels in the cytoplasm of leukocytes (Blind, M. et al. (1999) Proc. Natl. Acad. Sci. USA 96:3606-3610).

The term "spiegelmer" refers to an aptamer which includes L-DNA, L-RNA, or other left-handed nucleotide derivatives or nucleotide-like molecules. Aptamers containing left-handed nucleotides are resistant to degradation by naturally occurring enzymes, which normally act on substrates containing right-handed nucleotides.

The term "antisense" refers to any composition capable of base-pairing with the "sense" (coding) strand of a polynucleotide having a specific nucleic acid sequence. Antisense compositions may include DNA; RNA; peptide nucleic acid (PNA); oligonucleotides having modified backbone linkages such as phosphorothioates, methylphosphonates, or benzylphosphonates; oligonucleotides having modified sugar groups such as 2'-methoxyethyl sugars or 2'-methoxyethoxy sugars; or oligonucleotides having modified bases such as 5-methyl cytosine, 2'-deoxyuracil, or 7-deaza-2'-deoxyguanosine. Antisense molecules may be produced by any method including chemical synthesis or transcription. Once introduced into a cell, the complementary antisense molecule base-pairs with a naturally occurring nucleic acid sequence produced by the cell to form duplexes which block either transcription or translation. The designation "negative" or "minus" can refer to the antisense strand, and the designation "positive" or "plus" can refer to the sense strand of a reference DNA molecule.

The term "biologically active" refers to a protein having structural, regulatory, or biochemical functions of a naturally occurring molecule. Likewise, "immunologically active" or "immunogenic" refers to the capability of the natural, recombinant, or synthetic NAAP, or of any oligopeptide thereof, to induce a specific immune response in appropriate animals or cells and to bind with specific antibodies.

"Complementary" describes the relationship between two single-stranded nucleic acid sequences that annual by base-pairing. For example, 5'-AGT-3' pairs with its complement, 3'-TCA-5'.

A "composition comprising a given polynucleotide" and a "composition comprising a given polypeptide" can refer to any composition containing the given polynucleotide or polypeptide. The composition may comprise a dry formulation or an aqueous solution. Compositions comprising polynucleotides encoding NAAP or fragments of NAAP may be employed as hybridization probes.

The probes may be stored in freeze-dried form and may be associated with a stabilizing agent such as a carbohydrate. In hybridizations, the probe may be deployed in an aqueous solution containing salts (e.g., NaCl), detergents (e.g., sodium dodecyl sulfate; SDS), and other components (e.g., Denhardt's solution, dry milk, salmon sperm DNA, etc.).

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"Consensus sequence" refers to a nucleic acid sequence which has been subjected to repeated DNA sequence analysis to resolve uncalled bases, extended using the XL-PCR kit (Applied Biosystems, Foster City CA) in the 5' and/or the 3' direction, and resequenced, or which has been assembled from one or more overlapping cDNA, EST, or genomic DNA fragments using a computer program for fragment assembly, such as the GELVIEW fragment assembly system (Accelrys, Burlington MA) or Phrap (University of Washington, Seattle WA). Some sequences have been both extended and assembled to produce the consensus sequence.

"Conservative amino acid substitutions" are those substitutions that are predicted to least interfere with the properties of the original protein, i.e., the structure and especially the function of the protein is conserved and not significantly changed by such substitutions. The table below shows amino acids which may be substituted for an original amino acid in a protein and which are regarded as conservative amino acid substitutions.

	Original Residue	Conservative Substitution
	Ala	Gly, Ser
	Arg	His, Lys
25	Asn	Asp, Gln, His
	Asp	Asn, Glu
	Cys	Ala, Ser
	Gln	Asn, Glu, His
	Glu	Asp, Gln, His
	Gly	Ala
	His	Asn, Arg, Gln, Glu
	· Ile	Leu, Val
	Leu	Ile, Val
	Lys	Arg, Gln, Glu
30	Met	Leu, Ile
	Phe	His, Met, Leu, Trp, Tyr
	Ser	Cys, Thr
	Thr	Ser, Val
	Тгр	Phe, Tyr
35	Туг	His, Phe, Trp
	Val	Ile, Leu, Thr

Conservative amino acid substitutions generally maintain (a) the structure of the polypeptide backbone in the area of the substitution, for example, as a beta sheet or alpha helical conformation,

(b) the charge or hydrophobicity of the molecule at the site of the substitution, and/or (c) the bulk of the side chain.

A "deletion" refers to a change in the amino acid or nucleotide sequence that results in the absence of one or more amino acid residues or nucleotides.

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The term "derivative" refers to a chemically modified polynucleotide or polypeptide.

Chemical modifications of a polynucleotide can include, for example, replacement of hydrogen by an alkyl, acyl, hydroxyl, or amino group. A derivative polynucleotide encodes a polypeptide which retains at least one biological or immunological function of the natural molecule. A derivative polypeptide is one modified by glycosylation, pegylation, or any similar process that retains at least one biological or immunological function of the polypeptide from which it was derived.

A "detectable label" refers to a reporter molecule or enzyme that is capable of generating a measurable signal and is covalently or noncovalently joined to a polynucleotide or polypeptide.

"Differential expression" refers to increased or upregulated; or decreased, downregulated, or absent gene or protein expression, determined by comparing at least two different samples. Such comparisons may be carried out between, for example, a treated and an untreated sample, or a diseased and a normal sample.

"Exon shuffling" refers to the recombination of different coding regions (exons). Since an exon may represent a structural or functional domain of the encoded protein, new proteins may be assembled through the novel reassortment of stable substructures, thus allowing acceleration of the evolution of new protein functions.

A "fragment" is a unique portion of NAAP or a polynucleotide encoding NAAP which can be identical in sequence to, but shorter in length than, the parent sequence. A fragment may comprise up to the entire length of the defined sequence, minus one nucleotide/amino acid residue. For example, a fragment may comprise from about 5 to about 1000 contiguous nucleotides or amino acid residues. A fragment used as a probe, primer, antigen, therapeutic molecule, or for other purposes, may be at least 5, 10, 15, 16, 20, 25, 30, 40, 50, 60, 75, 100, 150, 250 or at least 500 contiguous nucleotides or amino acid residues in length. Fragments may be preferentially selected from certain regions of a molecule. For example, a polypeptide fragment may comprise a certain length of contiguous amino acids selected from the first 250 or 500 amino acids (or first 25% or 50%) of a polypeptide as shown in a certain defined sequence. Clearly these lengths are exemplary, and any length that is supported by the specification, including the Sequence Listing, tables, and figures, may be encompassed by the present embodiments.

A fragment of SEQ ID NO:58-114 can comprise a region of unique polynucleotide sequence that specifically identifies SEQ ID NO:58-114, for example, as distinct from any other sequence in the genome from which the fragment was obtained. A fragment of SEQ ID NO:58-114 can be employed in one or more embodiments of methods of the invention, for example, in hybridization and amplification technologies and in analogous methods that distinguish SEQ ID NO:58-114 from related polynucleotides. The precise length of a fragment of SEQ ID NO:58-114 and the region of SEQ ID NO:58-114 to which the fragment corresponds are routinely determinable by one of ordinary skill in the art based on the intended purpose for the fragment.

A fragment of SEQ ID NO:1-57 is encoded by a fragment of SEQ ID NO:58-114. A fragment of SEQ ID NO:1-57 can comprise a region of unique amino acid sequence that specifically identifies SEQ ID NO:1-57. For example, a fragment of SEQ ID NO:1-57 can be used as an immunogenic peptide for the development of antibodies that specifically recognize SEQ ID NO:1-57. The precise length of a fragment of SEQ ID NO:1-57 and the region of SEQ ID NO:1-57 to which the fragment corresponds can be determined based on the intended purpose for the fragment using one or more analytical methods described herein or otherwise known in the art.

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A "full length" polynucleotide is one containing at least a translation initiation codon (e.g., methionine) followed by an open reading frame and a translation termination codon. A "full length" polynucleotide sequence encodes a "full length" polypeptide sequence.

"Homology" refers to sequence similarity or, alternatively, sequence identity, between two or more polynucleotide sequences or two or more polypeptide sequences.

The terms "percent identity" and "% identity," as applied to polynucleotide sequences, refer to the percentage of identical residue matches between at least two polynucleotide sequences aligned using a standardized algorithm. Such an algorithm may insert, in a standardized and reproducible way, gaps in the sequences being compared in order to optimize alignment between two sequences, and therefore achieve a more meaningful comparison of the two sequences.

Percent identity between polynucleotide sequences may be determined using one or more computer algorithms or programs known in the art or described herein. For example, percent identity can be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program. This program is part of the LASERGENE software package, a suite of molecular biological analysis programs (DNASTAR, Madison WI). CLUSTAL V is described in Higgins, D.G. and P.M. Sharp (1989; CABIOS 5:151-153) and in Higgins, D.G. et al. (1992; CABIOS 8:189-191). For pairwise alignments of

polynucleotide sequences, the default parameters are set as follows: Ktuple=2, gap penalty=5, window=4, and "diagonals saved"=4. The "weighted" residue weight table is selected as the default.

Alternatively, a suite of commonly used and freely available sequence comparison algorithms which can be used is provided by the National Center for Biotechnology Information (NCBI) Basic

Local Alignment Search Tool (BLAST) (Altschul, S.F. et al. (1990) J. Mol. Biol. 215:403-410), which is available from several sources, including the NCBI, Bethesda, MD, and on the Internet at http://www.ncbi.nlm.nih.gov/BLAST/. The BLAST software suite includes various sequence analysis programs including "blastn," that is used to align a known polynucleotide sequence with other polynucleotide sequences from a variety of databases. Also available is a tool called "BLAST 2

Sequences" that is used for direct pairwise comparison of two nucleotide sequences. "BLAST 2

Sequences" can be accessed and used interactively at http://www.ncbi.nlm.nih.gov/gorf/bl2.html. The "BLAST 2 Sequences" tool can be used for both blastn and blastp (discussed below). BLAST programs are commonly used with gap and other parameters set to default settings. For example, to compare two nucleotide sequences, one may use blastn with the "BLAST 2 Sequences" tool Version

2.0.12 (April-21-2000) set at default parameters. Such default parameters may be, for example:

Matrix: BLOSUM62

Reward for match: 1

Penalty for mismatch: -2

Open Gap: 5 and Extension Gap: 2 penalties

Gap x drop-off: 50

Expect: 10

Word Size: 11

Filter: on

Percent identity may be measured over the length of an entire defined sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined sequence, for instance, a fragment of at least 20, at least 30, at least 40, at least 50, at least 70, at least 100, or at least 200 contiguous nucleotides. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures, or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

Nucleic acid sequences that do not show a high degree of identity may nevertheless encode similar amino acid sequences due to the degeneracy of the genetic code. It is understood that changes

in a nucleic acid sequence can be made using this degeneracy to produce multiple nucleic acid sequences that all encode substantially the same protein.

The phrases "percent identity" and "% identity," as applied to polypeptide sequences, refer to the percentage of identical residue matches between at least two polypeptide sequences aligned using a standardized algorithm. Methods of polypeptide sequence alignment are well-known. Some alignment methods take into account conservative amino acid substitutions. Such conservative substitutions, explained in more detail above, generally preserve the charge and hydrophobicity at the site of substitution, thus preserving the structure (and therefore function) of the polypeptide. The phrases "percent similarity" and "% similarity," as applied to polypeptide sequences, refer to the percentage of residue matches, including identical residue matches and conservative substitutions, between at least two polypeptide sequences aligned using a standardized algorithm. In contrast, conservative substitutions are not included in the calculation of percent identity between polypeptide sequences.

Percent identity between polypeptide sequences may be determined using the default parameters of the CLUSTAL V algorithm as incorporated into the MEGALIGN version 3.12e sequence alignment program (described and referenced above). For pairwise alignments of polypeptide sequences using CLUSTAL V, the default parameters are set as follows: Ktuple=1, gap penalty=3, window=5, and "diagonals saved"=5. The PAM250 matrix is selected as the default residue weight table.

Alternatively the NCBI BLAST software suite may be used. For example, for a pairwise comparison of two polypeptide sequences, one may use the "BLAST 2 Sequences" tool Version 2.0.12 (April-21-2000) with blastp set at default parameters. Such default parameters may be, for example:

. Matrix: BLOSUM62

Open Gap: 11 and Extension Gap: 1 penalties

Gap x drop-off: 50

Expect: 10
Word Size: 3

Filter: on

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Percent identity may be measured over the length of an entire defined polypeptide sequence, for example, as defined by a particular SEQ ID number, or may be measured over a shorter length, for example, over the length of a fragment taken from a larger, defined polypeptide sequence, for

instance, a fragment of at least 15, at least 20, at least 30, at least 40, at least 50, at least 70 or at least 150 contiguous residues. Such lengths are exemplary only, and it is understood that any fragment length supported by the sequences shown herein, in the tables, figures or Sequence Listing, may be used to describe a length over which percentage identity may be measured.

"Human artificial chromosomes" (HACs) are linear microchromosomes which may contain DNA sequences of about 6 kb to 10 Mb in size and which contain all of the elements required for chromosome replication, segregation and maintenance.

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The term "humanized antibody" refers to an antibody molecule in which the amino acid sequence in the non-antigen binding regions has been altered so that the antibody more closely resembles a human antibody, and still retains its original binding ability.

"Hybridization" refers to the process by which a polynucleotide strand anneals with a complementary strand through base pairing under defined hybridization conditions. Specific hybridization is an indication that two nucleic acid sequences share a high degree of complementarity. Specific hybridization complexes form under permissive annealing conditions and remain hybridized after the "washing" step(s). The washing step(s) is particularly important in determining the stringency of the hybridization process, with more stringent conditions allowing less non-specific binding, i.e., binding between pairs of nucleic acid strands that are not perfectly matched. Permissive conditions for annealing of nucleic acid sequences are routinely determinable by one of ordinary skill in the art and may be consistent among hybridization experiments, whereas wash conditions may be varied among experiments to achieve the desired stringency, and therefore hybridization specificity. Permissive annealing conditions occur, for example, at 68°C in the presence of about 6 x SSC, about 1% (w/v) SDS, and about 100 μg/ml sheared, denatured salmon sperm DNA.

Generally, stringency of hybridization is expressed, in part, with reference to the temperature under which the wash step is carried out. Such wash temperatures are typically selected to be about 5°C to 20°C lower than the thermal melting point (T_m) for the specific sequence at a defined ionic strength and pH. The T_m is the temperature (under defined ionic strength and pH) at which 50% of the target sequence hybridizes to a perfectly matched probe. An equation for calculating T_m and conditions for nucleic acid hybridization are well known and can be found in Sambrook, J. and D.W. Russell (2001; Molecular Cloning: A Laboratory Manual, 3rd ed., vol. 1-3, Cold Spring Harbor Press, Cold Spring Harbor NY, ch. 9).

High stringency conditions for hybridization between polynucleotides of the present invention include wash conditions of 68°C in the presence of about 0.2 x SSC and about 0.1% SDS, for 1 hour.

Alternatively, temperatures of about 65°C, 60°C, 55°C, or 42°C may be used. SSC concentration may be varied from about 0.1 to 2 x SSC, with SDS being present at about 0.1%. Typically, blocking reagents are used to block non-specific hybridization. Such blocking reagents include, for instance, sheared and denatured salmon sperm DNA at about 100-200 μ g/ml. Organic solvent, such as formamide at a concentration of about 35-50% v/v, may also be used under particular circumstances, such as for RNA:DNA hybridizations. Useful variations on these wash conditions will be readily apparent to those of ordinary skill in the art. Hybridization, particularly under high stringency conditions, may be suggestive of evolutionary similarity between the nucleotides. Such similarity is strongly indicative of a similar role for the nucleotides and their encoded polypeptides.

The term "hybridization complex" refers to a complex formed between two nucleic acids by virtue of the formation of hydrogen bonds between complementary bases. A hybridization complex may be formed in solution (e.g., C₀t or R₀t analysis) or formed between one nucleic acid present in solution and another nucleic acid immobilized on a solid support (e.g., paper, membranes, filters, chips, pins or glass slides, or any other appropriate substrate to which cells or their nucleic acids have been fixed).

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The words "insertion" and "addition" refer to changes in an amino acid or polynucleotide sequence resulting in the addition of one or more amino acid residues or nucleotides, respectively.

"Immune response" can refer to conditions associated with inflammation, trauma, immune disorders, or infectious or genetic disease, etc. These conditions can be characterized by expression of various factors, e.g., cytokines, chemokines, and other signaling molecules, which may affect cellular and systemic defense systems.

An "immunogenic fragment" is a polypeptide or oligopeptide fragment of NAAP which is capable of eliciting an immune response when introduced into a living organism, for example, a mammal. The term "immunogenic fragment" also includes any polypeptide or oligopeptide fragment of NAAP which is useful in any of the antibody production methods disclosed herein or known in the art.

The term "microarray" refers to an arrangement of a plurality of polynucleotides, polypeptides, antibodies, or other chemical compounds on a substrate.

The terms "element" and "array element" refer to a polynucleotide, polypeptide, antibody, or other chemical compound having a unique and defined position on a microarray.

The term "modulate" refers to a change in the activity of NAAP. For example, modulation may cause an increase or a decrease in protein activity, binding characteristics, or any other biological,

functional, or immunological properties of NAAP.

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The phrases "nucleic acid" and "nucleic acid sequence" refer to a nucleotide, oligonucleotide, polynucleotide, or any fragment thereof. These phrases also refer to DNA or RNA of genomic or synthetic origin which may be single-stranded or double-stranded and may represent the sense or the antisense strand, to peptide nucleic acid (PNA), or to any DNA-like or RNA-like material.

"Operably linked" refers to the situation in which a first nucleic acid sequence is placed in a functional relationship with a second nucleic acid sequence. For instance, a promoter is operably linked to a coding sequence if the promoter affects the transcription or expression of the coding sequence. Operably linked DNA sequences may be in close proximity or contiguous and, where necessary to join two protein coding regions, in the same reading frame.

"Peptide nucleic acid" (PNA) refers to an antisense molecule or anti-gene agent which comprises an oligonucleotide of at least about 5 nucleotides in length linked to a peptide backbone of amino acid residues ending in lysine. The terminal lysine confers solubility to the composition. PNAs preferentially bind complementary single stranded DNA or RNA and stop transcript elongation, and may be pegylated to extend their lifespan in the cell.

"Post-translational modification" of an NAAP may involve lipidation, glycosylation, phosphorylation, acetylation, racemization, proteolytic cleavage, and other modifications known in the art. These processes may occur synthetically or biochemically. Biochemical modifications will vary by cell type depending on the enzymatic milieu of NAAP.

"Probe" refers to nucleic acids encoding NAAP, their complements, or fragments thereof, which are used to detect identical, allelic or related nucleic acids. Probes are isolated oligonucleotides or polynucleotides attached to a detectable label or reporter molecule. Typical labels include radioactive isotopes, ligands, chemiluminescent agents, and enzymes. "Primers" are short nucleic acids, usually DNA oligonucleotides, which may be annealed to a target polynucleotide by 25 complementary base-pairing. The primer may then be extended along the target DNA strand by a DNA polymerase enzyme. Primer pairs can be used for amplification (and identification) of a nucleic acid, e.g., by the polymerase chain reaction (PCR).

Probes and primers as used in the present invention typically comprise at least 15 contiguous nucleotides of a known sequence. In order to enhance specificity, longer probes and primers may also be employed, such as probes and primers that comprise at least 20, 25, 30, 40, 50, 60, 70, 80, 90, 100, or at least 150 consecutive nucleotides of the disclosed nucleic acid sequences. Probes and primers may be considerably longer than these examples, and it is understood that any length supported by the

specification, including the tables, figures, and Sequence Listing, may be used.

Methods for preparing and using probes and primers are described in, for example, Sambrook, J. and D.W. Russell (2001; Molecular Cloning: A Laboratory Manual, 3rd ed., vol. 1-3, Cold Spring Harbor Press, Cold Spring Harbor NY), Ausubel, F.M. et al. (1999; Short Protocols in Molecular Biology, 4th ed., John Wiley & Sons, New York NY), and Innis, M. et al. (1990; PCR Protocols, A Guide to Methods and Applications, Academic Press, San Diego CA). PCR primer pairs can be derived from a known sequence, for example, by using computer programs intended for that purpose such as Primer (Version 0.5, 1991, Whitehead Institute for Biomedical Research, Cambridge MA).

Oligonucleotides for use as primers are selected using software known in the art for such purpose. For example, OLIGO 4.06 software is useful for the selection of PCR primer pairs of up to 100 nucleotides each, and for the analysis of oligonucleotides and larger polynucleotides of up to 5,000 nucleotides from an input polynucleotide sequence of up to 32 kilobases. Similar primer selection programs have incorporated additional features for expanded capabilities. For example, the PrimOU primer selection program (available to the public from the Genome Center at University of Texas South West Medical Center, Dallas TX) is capable of choosing specific primers from megabase sequences and is thus useful for designing primers on a genome-wide scope. The Primer3 primer selection program (available to the public from the Whitehead Institute/MIT Center for Genome Research, Cambridge MA) allows the user to input a "mispriming library," in which sequences to avoid as primer binding sites are user-specified. Primer3 is useful, in particular, for the selection of oligonucleotides for microarrays. (The source code for the latter two primer selection programs may also be obtained from their respective sources and modified to meet the user's specific needs.) The PrimeGen program (available to the public from the UK Human Genome Mapping Project Resource Centre, Cambridge UK) designs primers based on multiple sequence alignments, thereby allowing selection of primers that hybridize to either the most conserved or least conserved regions of aligned nucleic acid sequences. Hence, this program is useful for identification of both unique and conserved oligonucleotides and polynucleotide fragments. The oligonucleotides and polynucleotide fragments identified by any of the above selection methods are useful in hybridization technologies, for example, as PCR or sequencing primers, microarray elements, or specific probes to identify fully or partially complementary polynucleotides in a sample of nucleic acids. Methods of oligonucleotide selection are not limited to those described above.

A "recombinant nucleic acid" is a nucleic acid that is not naturally occurring or has a sequence that is made by an artificial combination of two or more otherwise separated segments of

sequence. This artificial combination is often accomplished by chemical synthesis or, more commonly, by the artificial manipulation of isolated segments of nucleic acids, e.g., by genetic engineering techniques such as those described in Sambrook and Russell (supra). The term recombinant includes nucleic acids that have been altered solely by addition, substitution, or deletion of a portion of the nucleic acid. Frequently, a recombinant nucleic acid may include a nucleic acid sequence operably linked to a promoter sequence. Such a recombinant nucleic acid may be part of a vector that is used, for example, to transform a cell.

Alternatively, such recombinant nucleic acids may be part of a viral vector, e.g., based on a vaccinia virus, that could be use to vaccinate a mammal wherein the recombinant nucleic acid is expressed, inducing a protective immunological response in the mammal.

A "regulatory element" refers to a nucleic acid sequence usually derived from untranslated regions of a gene and includes enhancers, promoters, introns, and 5' and 3' untranslated regions (UTRs). Regulatory elements interact with host or viral proteins which control transcription, translation, or RNA stability.

"Reporter molecules" are chemical or biochemical moieties used for labeling a nucleic acid, amino acid, or antibody. Reporter molecules include radionuclides; enzymes; fluorescent, chemiluminescent, or chromogenic agents; substrates; cofactors; inhibitors; magnetic particles; and other moieties known in the art.

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An "RNA equivalent," in reference to a DNA molecule, is composed of the same linear sequence of nucleotides as the reference DNA molecule with the exception that all occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The term "sample" is used in its broadest sense. A sample suspected of containing NAAP, nucleic acids encoding NAAP, or fragments thereof may comprise a bodily fluid; an extract from a cell, chromosome, organelle, or membrane isolated from a cell; a cell; genomic DNA, RNA, or cDNA, in solution or bound to a substrate; a tissue; a tissue print; etc.

The terms "specific binding" and "specifically binding" refer to that interaction between a protein or peptide and an agonist, an antibody, an antagonist, a small molecule, or any natural or synthetic binding composition. The interaction is dependent upon the presence of a particular structure of the protein, e.g., the antigenic determinant or epitope, recognized by the binding molecule. For example, if an antibody is specific for epitope "A," the presence of a polypeptide comprising the epitope A, or the presence of free unlabeled A, in a reaction containing free labeled A and the

antibody will reduce the amount of labeled A that binds to the antibody.

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The term "substantially purified" refers to nucleic acid or amino acid sequences that are removed from their natural environment and are isolated or separated, and are at least about 60% free, preferably at least about 75% free, and most preferably at least about 90% free from other components with which they are naturally associated.

A "substitution" refers to the replacement of one or more amino acid residues or nucleotides by different amino acid residues or nucleotides, respectively.

"Substrate" refers to any suitable rigid or semi-rigid support including membranes, filters, chips, slides, wafers, fibers, magnetic or nonmagnetic beads, gels, tubing, plates, polymers, microparticles and capillaries. The substrate can have a variety of surface forms, such as wells, trenches, pins, channels and pores, to which polynucleotides or polypeptides are bound.

A "transcript image" or "expression profile" refers to the collective pattern of gene expression by a particular cell type or tissue under given conditions at a given time.

"Transformation" describes a process by which exogenous DNA is introduced into a recipient cell. Transformation may occur under natural or artificial conditions according to various methods well known in the art, and may rely on any known method for the insertion of foreign nucleic acid sequences into a prokaryotic or eukaryotic host cell. The method for transformation is selected based on the type of host cell being transformed and may include, but is not limited to, bacteriophage or viral infection, electroporation, heat shock, lipofection, and particle bombardment. The term "transformed cells" includes stably transformed cells in which the inserted DNA is capable of replication either as an autonomously replicating plasmid or as part of the host chromosome, as well as transiently transformed cells which express the inserted DNA or RNA for limited periods of time.

A "transgenic organism," as used herein, is any organism, including but not limited to animals and plants, in which one or more of the cells of the organism contains heterologous nucleic acid introduced by way of human intervention, such as by transgenic techniques well known in the art. The nucleic acid is introduced into the cell, directly or indirectly by introduction into a precursor of the cell, by way of deliberate genetic manipulation, such as by microinjection or by infection with a recombinant virus. In another embodiment, the nucleic acid can be introduced by infection with a recombinant viral vector, such as a lentiviral vector (Lois, C. et al. (2002) Science 295:868-872). The term genetic manipulation does not include classical cross-breeding, or *in vitro* fertilization, but rather is directed to the introduction of a recombinant DNA molecule. The transgenic organisms contemplated in accordance with the present invention include bacteria, cyanobacteria, fungi, plants

and animals. The isolated DNA of the present invention can be introduced into the host by methods known in the art, for example infection, transfection, transformation or transconjugation. Techniques for transferring the DNA of the present invention into such organisms are widely known and provided in references such as Sambrook and Russell (supra).

A "variant" of a particular nucleic acid sequence is defined as a nucleic acid sequence having at least 40% sequence identity to the particular nucleic acid sequence over a certain length of one of the nucleic acid sequences using blastn with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of nucleic acids may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity over a certain defined length. A variant may be described as, for example, an "allelic" (as defined above), "splice," "species," or "polymorphic" variant. A splice variant may have significant identity to a reference molecule, but will generally have a greater or lesser number of polynucleotides due to alternate splicing of exons during mRNA processing. The corresponding polypeptide may possess additional functional domains or lack domains that are present in the reference molecule. Species variants are polynucleotides that vary from one species to another. The resulting polypeptides will generally have significant amino acid identity relative to each other. A polymorphic variant is a variation in the polynucleotide sequence of a particular gene between individuals of a given species. Polymorphic variants also may encompass "single nucleotide polymorphisms" (SNPs) in which the polynucleotide sequence varies by one nucleotide base. The presence of SNPs may be indicative of, for example, a certain population, a disease state, or a propensity for a disease state.

A "variant" of a particular polypeptide sequence is defined as a polypeptide sequence having at least 40% sequence identity or sequence similarity to the particular polypeptide sequence over a certain length of one of the polypeptide sequences using blastp with the "BLAST 2 Sequences" tool Version 2.0.9 (May-07-1999) set at default parameters. Such a pair of polypeptides may show, for example, at least 50%, at least 60%, at least 70%, at least 80%, at least 85%, at least 90%, at least 91%, at least 92%, at least 93%, at least 94%, at least 95%, at least 96%, at least 97%, at least 98%, or at least 99% or greater sequence identity or sequence similarity over a certain defined length of one of the polypeptides.

THE INVENTION

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Various embodiments of the invention include new human nucleic acid-associated proteins (NAAP), the polynucleotides encoding NAAP, and the use of these compositions for the diagnosis, treatment, or prevention of cell proliferative, neurological, developmental, and autoimmune/inflammatory disorders, and infections.

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Table 1 summarizes the nomenclature for the full length polynucleotide and polypeptide embodiments of the invention. Each polynucleotide and its corresponding polypeptide are correlated to a single Incyte project identification number (Incyte Project ID). Each polypeptide sequence is denoted by both a polypeptide sequence identification number (Polypeptide SEQ ID NO:) and an Incyte polypeptide sequence number (Incyte Polypeptide ID) as shown. Each polynucleotide sequence is denoted by both a polynucleotide sequence identification number (Polynucleotide SEQ ID NO:) and an Incyte polynucleotide consensus sequence number (Incyte Polynucleotide ID) as shown. Column 6 shows the Incyte ID numbers of physical, full length clones corresponding to the polypeptide and polynucleotide sequences of the invention. The full length clones encode polypeptides which have at least 95% sequence identity to the polypeptide sequences shown in column 3.

Table 2 shows sequences with homology to polypeptide embodiments of the invention as identified by BLAST analysis against the GenBank protein (genpept) database and the PROTEOME database. Columns 1 and 2 show the polypeptide sequence identification number (Polypeptide SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for polypeptides of the invention. Column 3 shows the GenBank identification number (GenBank ID NO:) of the nearest GenBank homolog and the PROTEOME database identification numbers (PROTEOME ID NO:) of the nearest PROTEOME database homologs. Column 4 shows the probability scores for the matches between each polypeptide and its homolog(s). Column 5 shows the annotation of the GenBank and PROTEOME database homolog(s) along with relevant citations where applicable, all of which are expressly incorporated by reference herein.

Table 3 shows various structural features of the polypeptides of the invention. Columns 1 and 2 show the polypeptide sequence identification number (SEQ ID NO:) and the corresponding Incyte polypeptide sequence number (Incyte Polypeptide ID) for each polypeptide of the invention. Column 3 shows the number of amino acid residues in each polypeptide. Column 4 shows potential phosphorylation sites, and column 5 shows potential glycosylation sites, as determined by the MOTIFS program of the GCG sequence analysis software package (Accelrys, Burlington MA). Column 6 shows amino acid residues comprising signature sequences, domains, and motifs. Column 7 shows analytical methods for protein structure/function analysis and in some cases, searchable databases to

which the analytical methods were applied.

Together, Tables 2 and 3 summarize the properties of polypeptides of the invention, and these properties establish that the claimed polypeptides are nucleic acid-associated proteins. For example, SEQ ID NO:11 is 100% identical, from residue M1 to residue L174 and from residue Q175 to residue Q488, to a human protein similar to interferon regulatory factor 5 (GenBank ID g13278720) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.8e-273, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:11 also has homology to human and mouse interferon regulatory factor 5, a transcription factor that induces expression of members of the interferon A family in response to NDV viral infection, as determined by BLAST analysis using the PROTEOME database. SEQ ID NO:11 also contains an interferon regulatory factor transcription factor domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, MOTIFS, and PROFILESCAN analyses provide further corroborative evidence that SEQ ID NO:11 is a member of the interferon regulatory factor (IRF) family of transcription factors.

In an alternative example, SEQ ID NO:24 is 96% identical, from residue M1 to residue R133, to human hMBF1alpha (GenBank ID g6526355) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 5.0e-63, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:24 also has homology to endothelial differentiation-related factor-1, a putative transcriptional coactivator that binds to calmodulin (CALM1), in a calcium-dependent manner, and to the TATA-binding protein (TBP), has transcription factor function, and are EDF1 transcription coactivators, as determined by BLAST analysis using the PROTEOME database. SEQ ID NO:24 also contains a helix-turn-helix domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from additional BLAST analyses provide further corroborative evidence that SEQ ID NO:24 is a transcription factor regulator.

In an alternative example, SEQ ID NO:35 is 84% identical, from residue M932 to residue G1638, to human ras-responsive element binding protein (GenBank ID g1654112) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 0.0, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:35 also has homology to nuclear transcription factors, as determined by BLAST

PCT/US02/41115 WO 03/054219

analysis using the PROTEOME database. SEQ ID NO:35 also contains a zinc finger domain as determined by searching for statistically significant matches in the hidden Markov model (HMM)based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS and MOTIFS analyses provide further corroborative evidence that SEQ ID NO:35 is a transcription factor.

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In an alternative example, SEQ ID NO:55 is 85% identical, from residue M1 to residue V508, to rat serine-arginine-rich splicing regulatory protein, SRRP86 (GenBank ID g7158880) as determined by the Basic Local Alignment Search Tool (BLAST). (See Table 2.) The BLAST probability score is 4.8e-225, which indicates the probability of obtaining the observed polypeptide sequence alignment by chance. SEQ ID NO:55 also has homology to proteins that are localized to the nucleus, function as RNA-binding proteins, and are serine-arginine-rich proteins, as determined by BLAST analysis using the PROTEOME database. SEQ ID NO:55 also contains an RNA recognition motif as determined by searching for statistically significant matches in the hidden Markov model (HMM)-based PFAM database of conserved protein family domains. (See Table 3.) Data from BLIMPS, and other BLAST analyses provide further corroborative evidence that SEQ ID NO:55 is a serine-arginine-rich splicing regulatory protein.

SEQ ID NO:1-10, SEQ ID NO:12-23, SEQ ID NO:25-34, SEQ ID NO:36-54 and SEQ ID NO:56-57 were analyzed and annotated in a similar manner. The algorithms and parameters for the analysis of SEQ ID NO:1-57 are described in Table 7.

As shown in Table 4, the full length polynucleotide embodiments were assembled using cDNA sequences or coding (exon) sequences derived from genomic DNA, or any combination of these two types of sequences. Column 1 lists the polynucleotide sequence identification number (Polynucleotide SEQ ID NO:), the corresponding Incyte polynucleotide consensus sequence number (Incyte ID) for each polynucleotide of the invention, and the length of each polynucleotide sequence in basepairs. 25 Column 2 shows the nucleotide start (5') and stop (3') positions of the cDNA and/or genomic sequences used to assemble the full length polynucleotide embodiments, and of fragments of the polynucleotides which are useful, for example, in hybridization or amplification technologies that identify SEQ ID NO:58-114 or that distinguish between SEQ ID NO:58-114 and related polynucleotides.

The polynucleotide fragments described in Column 2 of Table 4 may refer specifically, for example, to Incyte cDNAs derived from tissue-specific cDNA libraries or from pooled cDNA libraries. Alternatively, the polynucleotide fragments described in column 2 may refer to GenBank

cDNAs or ESTs which contributed to the assembly of the full length polynucleotides. In addition, the polynucleotide fragments described in column 2 may identify sequences derived from the ENSEMBL (The Sanger Centre, Cambridge, UK) database (i.e., those sequences including the designation "ENST"). Alternatively, the polynucleotide fragments described in column 2 may be derived from the NCBI RefSeq Nucleotide Sequence Records Database (i.e., those sequences including the designation "NM" or "NT") or the NCBI RefSeq Protein Sequence Records (i.e., those sequences including the designation "NP"). Alternatively, the polynucleotide fragments described in column 2 may refer to assemblages of both cDNA and Genscan-predicted exons brought together by an "exon stitching" algorithm. For example, a polynucleotide sequence identified as

FL_XXXXXX_N₁_N₂_YYYYY_N₃_N₄ represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the

FL_XXXXXX_ N_1 _ N_2 _YYYYY_ N_3 _ N_4 represents a "stitched" sequence in which XXXXXX is the identification number of the cluster of sequences to which the algorithm was applied, and YYYYY is the number of the prediction generated by the algorithm, and $N_{1,2,3...}$, if present, represent specific exons that may have been manually edited during analysis (See Example V). Alternatively, the polynucleotide fragments in column 2 may refer to assemblages of exons brought together by an "exon-stretching" algorithm. For example, a polynucleotide sequence identified as

FLXXXXXX_gAAAAA_gBBBBB_1_N is a "stretched" sequence, with XXXXXX being the Incyte project identification number, gAAAAA being the GenBank identification number of the human genomic sequence to which the "exon-stretching" algorithm was applied, gBBBBB being the GenBank identification number or NCBI RefSeq identification number of the nearest GenBank protein homolog, and N referring to specific exons (See Example V). In instances where a RefSeq sequence was used as a protein homolog for the "exon-stretching" algorithm, a RefSeq identifier (denoted by "NM,"

Alternatively, a prefix identifies component sequences that were hand-edited, predicted from genomic DNA sequences, or derived from a combination of sequence analysis methods. The

"NP," or "NT") may be used in place of the GenBank identifier (i.e., gBBBBB).

following Table lists examples of component sequence prefixes and corresponding sequence analysis methods associated with the prefixes (see Example IV and Example V).

Prefix	Type of analysis and/or examples of programs
GNN, GFG,	Exon prediction from genomic sequences using, for example,
ENST	GENSCAN (Stanford University, CA, USA) or FGENES
	(Computer Genomics Group, The Sanger Centre, Cambridge, UK).
GBI	Hand-edited analysis of genomic sequences.
FL	Stitched or stretched genomic sequences (see Example V).

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INCY	Full length transcript and exon prediction from mapping of EST	
	sequences to the genome. Genomic location and EST composition	
	data are combined to predict the exons and resulting transcript.	

In some cases, Incyte cDNA coverage redundant with the sequence coverage shown in Table 4 was obtained to confirm the final consensus polynucleotide sequence, but the relevant Incyte cDNA identification numbers are not shown.

Table 5 shows the representative cDNA libraries for those full length polynucleotides which were assembled using Incyte cDNA sequences. The representative cDNA library is the Incyte cDNA library which is most frequently represented by the Incyte cDNA sequences which were used to assemble and confirm the above polynucleotides. The tissues and vectors which were used to construct the cDNA libraries shown in Table 5 are described in Table 6.

Table 8 shows single nucleotide polymorphisms (SNPs) found in polynucleotide sequences of the invention, along with allele frequencies in different human populations. Columns 1 and 2 show the polynucleotide sequence identification number (SEQ ID NO:) and the corresponding Incyte project identification number (PID) for polynucleotides of the invention. Column 3 shows the Incyte identification number for the EST in which the SNP was detected (EST ID), and column 4 shows the identification number for the SNP (SNP ID). Column 5 shows the position within the EST sequence at which the SNP is located (EST SNP), and column 6 shows the position of the SNP within the full-length polynucleotide sequence (CB1 SNP). Column 7 shows the allele found in the EST sequence. Columns 8 and 9 show the two alleles found at the SNP site. Column 10 shows the amino acid encoded by the codon including the SNP site, based upon the allele found in the EST. Columns 11-14 show the frequency of allele 1 in four different human populations. An entry of n/d (not detected) indicates that the frequency of allele 1 in the population was too low to be detected, while n/a (not available) indicates that the allele frequency was not determined for the population.

The invention also encompasses NAAP variants. Various embodiments of NAAP variants can have at least about 80%, at least about 90%, or at least about 95% amino acid sequence identity to the NAAP amino acid sequence, and can contain at least one functional or structural characteristic of NAAP.

Various embodiments also encompass polynucleotides which encode NAAP. In a particular embodiment, the invention encompasses a polynucleotide sequence comprising a sequence selected from the group consisting of SEQ ID NO:58-114, which encodes NAAP. The polynucleotide

sequences of SEQ ID NO:58-114, as presented in the Sequence Listing, embrace the equivalent RNA sequences, wherein occurrences of the nitrogenous base thymine are replaced with uracil, and the sugar backbone is composed of ribose instead of deoxyribose.

The invention also encompasses variants of a polynucleotide encoding NAAP. In particular, such a variant polynucleotide will have at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a polynucleotide encoding NAAP. A particular aspect of the invention encompasses a variant of a polynucleotide comprising a sequence selected from the group consisting of SEQ ID NO:58-114 which has at least about 70%, or alternatively at least about 85%, or even at least about 95% polynucleotide sequence identity to a nucleic acid sequence selected from the group consisting of SEQ ID NO:58-114. Any one of the polynucleotide variants described above can encode a polypeptide which contains at least one functional or structural characteristic of NAAP.

In addition, or in the alternative, a polynucleotide variant of the invention is a splice variant of a polynucleotide encoding NAAP. A splice variant may have portions which have significant sequence identity to a polynucleotide encoding NAAP, but will generally have a greater or lesser number of polynucleotides due to additions or deletions of blocks of sequence arising from alternate splicing of exons during mRNA processing. A splice variant may have less than about 70%, or alternatively less than about 60%, or alternatively less than about 50% polynucleotide sequence identity to a polynucleotide encoding NAAP over its entire length; however, portions of the splice variant will have at least about 70%, or alternatively at least about 85%, or alternatively at least about 95%, or alternatively 100% polynucleotide sequence identity to portions of the polynucleotide encoding NAAP. Any one of the splice variants described above can encode a polypeptide which contains at least one functional or structural characteristic of NAAP.

It will be appreciated by those skilled in the art that as a result of the degeneracy of the genetic code, a multitude of polynucleotide sequences encoding NAAP, some bearing minimal similarity to the polynucleotide sequences of any known and naturally occurring gene, may be produced. Thus, the invention contemplates each and every possible variation of polynucleotide sequence that could be made by selecting combinations based on possible codon choices. These combinations are made in accordance with the standard triplet genetic code as applied to the polynucleotide sequence of naturally occurring NAAP, and all such variations are to be considered as being specifically disclosed.

Although polynucleotides which encode NAAP and its variants are generally capable of

hybridizing to polynucleotides encoding naturally occurring NAAP under appropriately selected conditions of stringency, it may be advantageous to produce polynucleotides encoding NAAP or its derivatives possessing a substantially different codon usage, e.g., inclusion of non-naturally occurring codons. Codons may be selected to increase the rate at which expression of the peptide occurs in a particular prokaryotic or eukaryotic host in accordance with the frequency with which particular codons are utilized by the host. Other reasons for substantially altering the nucleotide sequence encoding NAAP and its derivatives without altering the encoded amino acid sequences include the production of RNA transcripts having more desirable properties, such as a greater half-life, than transcripts produced from the naturally occurring sequence.

The invention also encompasses production of polynucleotides which encode NAAP and NAAP derivatives, or fragments thereof, entirely by synthetic chemistry. After production, the synthetic polynucleotide may be inserted into any of the many available expression vectors and cell systems using reagents well known in the art. Moreover, synthetic chemistry may be used to introduce mutations into a polynucleotide encoding NAAP or any fragment thereof.

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Embodiments of the invention can also include polynucleotides that are capable of hybridizing to the claimed polynucleotides, and, in particular, to those having the sequences shown in SEQ ID NO:58-114 and fragments thereof, under various conditions of stringency (Wahl, G.M. and S.L. Berger (1987) Methods Enzymol. 152:399-407; Kimmel, A.R. (1987) Methods Enzymol. 152:507-511). Hybridization conditions, including annealing and wash conditions, are described in "Definitions."

Methods for DNA sequencing are well known in the art and may be used to practice any of the embodiments of the invention. The methods may employ such enzymes as the Klenow fragment of DNA polymerase I, SEQUENASE (US Biochemical, Cleveland OH), Taq polymerase (Applied Biosystems), thermostable T7 polymerase (Amersham Biosciences, Piscataway NJ), or combinations of polymerases and proofreading exonucleases such as those found in the ELONGASE amplification system (Invitrogen, Carlsbad CA). Preferably, sequence preparation is automated with machines such as the MICROLAB 2200 liquid transfer system (Hamilton, Reno NV), PTC200 thermal cycler (MJ Research, Watertown MA) and ABI CATALYST 800 thermal cycler (Applied Biosystems). Sequencing is then carried out using either the ABI 373 or 377 DNA sequencing system (Applied Biosystems), the MEGABACE 1000 DNA sequencing system (Amersham Biosciences), or other systems known in the art. The resulting sequences are analyzed using a variety of algorithms which are well known in the art (Ausubel et al., supra, ch. 7; Meyers, R.A. (1995) Molecular Biology and Biotechnology, Wiley VCH, New York NY, pp. 856-853).

The nucleic acids encoding NAAP may be extended utilizing a partial nucleotide sequence and employing various PCR-based methods known in the art to detect upstream sequences, such as promoters and regulatory elements. For example, one method which may be employed, restriction-site PCR, uses universal and nested primers to amplify unknown sequence from genomic DNA within a cloning vector (Sarkar, G. (1993) PCR Methods Applic. 2:318-322). Another method, inverse PCR, uses primers that extend in divergent directions to amplify unknown sequence from a circularized template. The template is derived from restriction fragments comprising a known genomic locus and surrounding sequences (Triglia, T. et al. (1988) Nucleic Acids Res. 16:8186). A third method, capture PCR, involves PCR amplification of DNA fragments adjacent to known sequences in human and yeast artificial chromosome DNA (Lagerstrom, M. et al. (1991) PCR Methods Applic. 1:111-119). In this method, multiple restriction enzyme digestions and ligations may be used to insert an engineered double-stranded sequence into a region of unknown sequence before performing PCR. Other methods which may be used to retrieve unknown sequences are known in the art (Parker, J.D. et al. (1991) Nucleic Acids Res. 19:3055-3060). Additionally, one may use PCR, nested primers, and PROMOTERFINDER libraries (Clontech, Palo Alto CA) to walk genomic DNA. This procedure avoids the need to screen libraries and is useful in finding intron/exon junctions. For all PCR-based methods, primers may be designed using commercially available software, such as OLIGO 4.06 primer analysis software (National Biosciences, Plymouth MN) or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the template at temperatures of about 68°C to 72°C.

When screening for full length cDNAs, it is preferable to use libraries that have been size-selected to include larger cDNAs. In addition, random-primed libraries, which often include sequences containing the 5' regions of genes, are preferable for situations in which an oligo d(T) library does not yield a full-length cDNA. Genomic libraries may be useful for extension of sequence into 5' non-transcribed regulatory regions.

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Capillary electrophoresis systems which are commercially available may be used to analyze the size or confirm the nucleotide sequence of sequencing or PCR products. In particular, capillary sequencing may employ flowable polymers for electrophoretic separation, four different nucleotide-specific, laser-stimulated fluorescent dyes, and a charge coupled device camera for detection of the emitted wavelengths. Output/light intensity may be converted to electrical signal using appropriate software (e.g., GENOTYPER and SEQUENCE NAVIGATOR, Applied Biosystems), and the entire process from loading of samples to computer analysis and electronic data display may be computer

controlled. Capillary electrophoresis is especially preferable for sequencing small DNA fragments which may be present in limited amounts in a particular sample.

In another embodiment of the invention, polynucleotides or fragments thereof which encode NAAP may be cloned in recombinant DNA molecules that direct expression of NAAP, or fragments or functional equivalents thereof, in appropriate host cells. Due to the inherent degeneracy of the genetic code, other polynucleotides which encode substantially the same or a functionally equivalent polypeptides may be produced and used to express NAAP.

The polynucleotides of the invention can be engineered using methods generally known in the art in order to alter NAAP-encoding sequences for a variety of purposes including, but not limited to, modification of the cloning, processing, and/or expression of the gene product. DNA shuffling by random fragmentation and PCR reassembly of gene fragments and synthetic oligonucleotides may be used to engineer the nucleotide sequences. For example, oligonucleotide-mediated site-directed mutagenesis may be used to introduce mutations that create new restriction sites, alter glycosylation patterns, change codon preference, produce splice variants, and so forth.

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The nucleotides of the present invention may be subjected to DNA shuffling techniques such as MOLECULARBREEDING (Maxygen Inc., Santa Clara CA; described in U.S. Patent No. 5,837,458; Chang, C.-C. et al. (1999) Nat. Biotechnol. 17:793-797; Christians, F.C. et al. (1999) Nat. Biotechnol. 17:259-264; and Crameri, A. et al. (1996) Nat. Biotechnol. 14:315-319) to alter or improve the biological properties of NAAP, such as its biological or enzymatic activity or its ability to bind to other molecules or compounds. DNA shuffling is a process by which a library of gene variants is produced using PCR-mediated recombination of gene fragments. The library is then subjected to selection or screening procedures that identify those gene variants with the desired properties. These preferred variants may then be pooled and further subjected to recursive rounds of DNA shuffling and selection/screening. Thus, genetic diversity is created through "artificial" breeding and rapid molecular evolution. For example, fragments of a single gene containing random point mutations may be recombined, screened, and then reshuffled until the desired properties are optimized. Alternatively, fragments of a given gene may be recombined with fragments of homologous genes in the same gene family, either from the same or different species, thereby maximizing the genetic diversity of multiple naturally occurring genes in a directed and controllable manner.

In another embodiment, polynucleotides encoding NAAP may be synthesized, in whole or in part, using one or more chemical methods well known in the art (Caruthers, M.H. et al. (1980) Nucleic Acids Symp. Ser. 7:215-223; Horn, T. et al. (1980) Nucleic Acids Symp. Ser. 7:225-232).

Alternatively, NAAP itself or a fragment thereof may be synthesized using chemical methods known in the art. For example, peptide synthesis can be performed using various solution-phase or solid-phase techniques (Creighton, T. (1984) Proteins, Structures and Molecular Properties, WH Freeman, New York NY, pp. 55-60; Roberge, J.Y. et al. (1995) Science 269:202-204). Automated synthesis may be achieved using the ABI 431A peptide synthesizer (Applied Biosystems). Additionally, the amino acid sequence of NAAP, or any part thereof, may be altered during direct synthesis and/or combined with sequences from other proteins, or any part thereof, to produce a variant polypeptide or a polypeptide having a sequence of a naturally occurring polypeptide.

The peptide may be substantially purified by preparative high performance liquid chromatography (Chiez, R.M. and F.Z. Regnier (1990) Methods Enzymol. 182:392-421). The composition of the synthetic peptides may be confirmed by amino acid analysis or by sequencing (Creighton, *supra*, pp. 28-53).

In order to express a biologically active NAAP, the polynucleotides encoding NAAP or derivatives thereof may be inserted into an appropriate expression vector, i.e., a vector which contains the necessary elements for transcriptional and translational control of the inserted coding sequence in a suitable host. These elements include regulatory sequences, such as enhancers, constitutive and inducible promoters, and 5' and 3' untranslated regions in the vector and in polynucleotides encoding NAAP. Such elements may vary in their strength and specificity. Specific initiation signals may also be used to achieve more efficient translation of polynucleotides encoding NAAP. Such signals include the ATG initiation codon and adjacent sequences, e.g. the Kozak sequence. In cases where a polynucleotide sequence encoding NAAP and its initiation codon and upstream regulatory sequences are inserted into the appropriate expression vector, no additional transcriptional or translational control signals may be needed. However, in cases where only coding sequence, or a fragment thereof, is inserted, exogenous translational control signals including an in-frame ATG initiation codon should be provided by the vector. Exogenous translational elements and initiation codons may be of various origins, both natural and synthetic. The efficiency of expression may be enhanced by the inclusion of enhancers appropriate for the particular host cell system used (Scharf, D. et al. (1994) Results Probl. Cell Differ. 20:125-162).

Methods which are well known to those skilled in the art may be used to construct expression vectors containing polynucleotides encoding NAAP and appropriate transcriptional and translational control elements. These methods include *in vitro* recombinant DNA techniques, synthetic techniques, and *in vivo* genetic recombination (Sambrook and Russell, *supra*, ch. 1-4, and 8; Ausubel et al.,

supra, ch. 1, 3, and 15).

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A variety of expression vector/host systems may be utilized to contain and express polynucleotides encoding NAAP. These include, but are not limited to, microorganisms such as bacteria transformed with recombinant bacteriophage, plasmid, or cosmid DNA expression vectors; yeast transformed with yeast expression vectors; insect cell systems infected with viral expression vectors (e.g., baculovirus); plant cell systems transformed with viral expression vectors (e.g., cauliflower mosaic virus, CaMV, or tobacco mosaic virus, TMV) or with bacterial expression vectors (e.g., Ti or pBR322 plasmids); or animal cell systems (Sambrook and Russell, supra; Ausubel et al., supra; Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509; Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945; Takamatsu, N. (1987) EMBO J. 6:307-311; The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196; Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659; Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355). Expression vectors derived from retroviruses, adenoviruses, or herpes or vaccinia viruses, or from various bacterial plasmids, may be used for delivery of polynucleotides to the targeted organ, tissue, or cell population (Di Nicola, M. et al. (1998) Cancer Gen. Ther. 5:350-356; Yu, M. et al. (1993) Proc. Natl. Acad. Sci. USA 90:6340-6344; Buller, R.M. et al. (1985) Nature 317:813-815; McGregor, D.P. et al. (1994) Mol. Immunol. 31:219-226; Verma, I.M. and N. Somia (1997) Nature 389:239-242). The invention is not limited by the host cell employed.

In bacterial systems, a number of cloning and expression vectors may be selected depending upon the use intended for polynucleotides encoding NAAP. For example, routine cloning, subcloning, and propagation of polynucleotides encoding NAAP can be achieved using a multifunctional *E. coli* vector such as PBLUESCRIPT (Stratagene, La Jolla CA) or PSPORT1 plasmid (Invitrogen). Ligation of polynucleotides encoding NAAP into the vector's multiple cloning site disrupts the *lacZ* gene, allowing a colorimetric screening procedure for identification of transformed bacteria containing recombinant molecules. In addition, these vectors may be useful for *in vitro* transcription, dideoxy sequencing, single strand rescue with helper phage, and creation of nested deletions in the cloned sequence (Van Heeke, G. and S.M. Schuster (1989) J. Biol. Chem. 264:5503-5509). When large quantities of NAAP are needed, e.g. for the production of antibodies, vectors which direct high level expression of NAAP may be used. For example, vectors containing the strong, inducible SP6 or T7 bacteriophage promoter may be used.

Yeast expression systems may be used for production of NAAP. A number of vectors

containing constitutive or inducible promoters, such as alpha factor, alcohol oxidase, and PGH promoters, may be used in the yeast Saccharomyces cerevisiae or Pichia pastoris. In addition, such vectors direct either the secretion or intracellular retention of expressed proteins and enable integration of foreign polynucleotide sequences into the host genome for stable propagation (Ausubel et al., supra; Bitter, G.A. et al. (1987) Methods Enzymol. 153:516-544; Scorer, C.A. et al. (1994) Bio/Technology 12:181-184).

Plant systems may also be used for expression of NAAP. Transcription of polynucleotides encoding NAAP may be driven by viral promoters, e.g., the 35S and 19S promoters of CaMV used alone or in combination with the omega leader sequence from TMV (Takamatsu, N. (1987) EMBO J. 6:307-311). Alternatively, plant promoters such as the small subunit of RUBISCO or heat shock promoters may be used (Coruzzi, G. et al. (1984) EMBO J. 3:1671-1680; Broglie, R. et al. (1984) Science 224:838-843; Winter, J. et al. (1991) Results Probl. Cell Differ. 17:85-105). These constructs can be introduced into plant cells by direct DNA transformation or pathogen-mediated transfection (The McGraw Hill Yearbook of Science and Technology (1992) McGraw Hill, New York NY, pp. 191-196).

In mammalian cells, a number of viral-based expression systems may be utilized. In cases where an adenovirus is used as an expression vector, polynucleotides encoding NAAP may be ligated into an adenovirus transcription/translation complex consisting of the late promoter and tripartite leader sequence. Insertion in a non-essential E1 or E3 region of the viral genome may be used to obtain infective virus which expresses NAAP in host cells (Logan, J. and T. Shenk (1984) Proc. Natl. Acad. Sci. USA 81:3655-3659). In addition, transcription enhancers, such as the Rous sarcoma virus (RSV) enhancer, may be used to increase expression in mammalian host cells. SV40 or EBV-based vectors may also be used for high-level protein expression.

Human artificial chromosomes (HACs) may also be employed to deliver larger fragments of DNA than can be contained in and expressed from a plasmid. HACs of about 6 kb to 10 Mb are constructed and delivered via conventional delivery methods (liposomes, polycationic amino polymers, or vesicles) for therapeutic purposes (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355).

For long term production of recombinant proteins in mammalian systems, stable expression of NAAP in cell lines is preferred. For example, polynucleotides encoding NAAP can be transformed into cell lines using expression vectors which may contain viral origins of replication and/or endogenous expression elements and a selectable marker gene on the same or on a separate vector. Following the introduction of the vector, cells may be allowed to grow for about 1 to 2 days in enriched

media before being switched to selective media. The purpose of the selectable marker is to confer resistance to a selective agent, and its presence allows growth and recovery of cells which successfully express the introduced sequences. Resistant clones of stably transformed cells may be propagated using tissue culture techniques appropriate to the cell type.

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Any number of selection systems may be used to recover transformed cell lines. These include, but are not limited to, the herpes simplex virus thymidine kinase and adenine phosphoribosyltransferase genes, for use in *tk* and *apr* cells, respectively (Wigler, M. et al. (1977) Cell 11:223-232; Lowy, I. et al. (1980) Cell 22:817-823). Also, antimetabolite, antibiotic, or herbicide resistance can be used as the basis for selection. For example, *dhfr* confers resistance to methotrexate; *neo* confers resistance to the aminoglycosides neomycin and G-418; and *als* and *pat* confer resistance to chlorsulfuron and phosphinotricin acetyltransferase, respectively (Wigler, M. et al. (1980) Proc. Natl. Acad. Sci. USA 77:3567-3570; Colbere-Garapin, F. et al. (1981) J. Mol. Biol. 150:1-14). Additional selectable genes have been described, e.g., *trpB* and *hisD*, which alter cellular requirements for metabolites (Hartman, S.C. and R.C. Mulligan (1988) Proc. Natl. Acad. Sci. USA 85:8047-8051). Visible markers, e.g., anthocyanins, green fluorescent proteins (GFP; Clontech), β-glucuronidase and its substrate β-glucuronide, or luciferase and its substrate luciferin may be used. These markers can be used not only to identify transformants, but also to quantify the amount of transient or stable protein expression attributable to a specific vector system (Rhodes, C.A. (1995) Methods Mol. Biol. 55:121-131).

Although the presence/absence of marker gene expression suggests that the gene of interest is also present, the presence and expression of the gene may need to be confirmed. For example, if the sequence encoding NAAP is inserted within a marker gene sequence, transformed cells containing polynucleotides encoding NAAP can be identified by the absence of marker gene function.

Alternatively, a marker gene can be placed in tandem with a sequence encoding NAAP under the control of a single promoter. Expression of the marker gene in response to induction or selection usually indicates expression of the tandem gene as well.

In general, host cells that contain the polynucleotide encoding NAAP and that express NAAP may be identified by a variety of procedures known to those of skill in the art. These procedures include, but are not limited to, DNA-DNA or DNA-RNA hybridizations, PCR amplification, and protein bioassay or immunoassay techniques which include membrane, solution, or chip based technologies for the detection and/or quantification of nucleic acid or protein sequences.

Immunological methods for detecting and measuring the expression of NAAP using either

specific polyclonal or monoclonal antibodies are known in the art. Examples of such techniques include enzyme-linked immunosorbent assays (ELISAs), radioimmunoassays (RIAs), and fluorescence activated cell sorting (FACS). A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering epitopes on NAAP is preferred, but a competitive binding assay may be employed. These and other assays are well known in the art (Hampton, R. et al. (1990) Serological Methods, a Laboratory Manual, APS Press, St. Paul MN, Sect. IV; Coligan, J.E. et al. (1997) Current Protocols in Immunology, Greene Pub. Associates and Wiley-Interscience, New York NY; Pound, J.D. (1998) Immunochemical Protocols, Humana Press, Totowa NJ).

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A wide variety of labels and conjugation techniques are known by those skilled in the art and may be used in various nucleic acid and amino acid assays. Means for producing labeled hybridization or PCR probes for detecting sequences related to polynucleotides encoding NAAP include oligolabeling, nick translation, end-labeling, or PCR amplification using a labeled nucleotide.

Alternatively, polynucleotides encoding NAAP, or any fragments thereof, may be cloned into a vector for the production of an mRNA probe. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes *in vitro* by addition of an appropriate RNA polymerase such as T7, T3, or SP6 and labeled nucleotides. These procedures may be conducted using a variety of commercially available kits, such as those provided by Amersham Biosciences, Promega (Madison WI), and US Biochemical. Suitable reporter molecules or labels which may be used for ease of detection include radionuclides, enzymes, fluorescent, chemiluminescent, or chromogenic agents, as well as substrates, cofactors, inhibitors, magnetic particles, and the like.

Host cells transformed with polynucleotides encoding NAAP may be cultured under conditions suitable for the expression and recovery of the protein from cell culture. The protein produced by a transformed cell may be secreted or retained intracellularly depending on the sequence and/or the vector used. As will be understood by those of skill in the art, expression vectors containing polynucleotides which encode NAAP may be designed to contain signal sequences which direct secretion of NAAP through a prokaryotic or eukaryotic cell membrane.

In addition, a host cell strain may be chosen for its ability to modulate expression of the inserted polynucleotides or to process the expressed protein in the desired fashion. Such modifications of the polypeptide include, but are not limited to, acetylation, carboxylation, glycosylation, phosphorylation, lipidation, and acylation. Post-translational processing which cleaves a "prepro" or "pro" form of the protein may also be used to specify protein targeting, folding, and/or activity.

Different host cells which have specific cellular machinery and characteristic mechanisms for post-translational activities (e.g., CHO, HeLa, MDCK, HEK293, and WI38) are available from the American Type Culture Collection (ATCC, Manassas VA) and may be chosen to ensure the correct modification and processing of the foreign protein.

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In another embodiment of the invention, natural, modified, or recombinant polynucleotides encoding NAAP may be ligated to a heterologous sequence resulting in translation of a fusion protein in any of the aforementioned host systems. For example, a chimeric NAAP protein containing a heterologous moiety that can be recognized by a commercially available antibody may facilitate the screening of peptide libraries for inhibitors of NAAP activity. Heterologous protein and peptide moieties may also facilitate purification of fusion proteins using commercially available affinity matrices. Such moieties include, but are not limited to, glutathione S-transferase (GST), maltose binding protein (MBP), thioredoxin (Trx), calmodulin binding peptide (CBP), 6-His, FLAG, c-myc, and hemagglutinin (HA). GST, MBP, Trx, CBP, and 6-His enable purification of their cognate fusion proteins on immobilized glutathione, maltose, phenylarsine oxide, calmodulin, and metal-chelate resins, 15 respectively. FLAG, c-myc, and hemagglutinin (HA) enable immunoaffinity purification of fusion proteins using commercially available monoclonal and polyclonal antibodies that specifically recognize these epitope tags. A fusion protein may also be engineered to contain a proteolytic cleavage site located between the NAAP encoding sequence and the heterologous protein sequence, so that NAAP may be cleaved away from the heterologous moiety following purification. Methods for fusion protein expression and purification are discussed in Ausubel et al. (supra, ch. 10 and 16). A variety of commercially available kits may also be used to facilitate expression and purification of fusion proteins.

In another embodiment, synthesis of radiolabeled NAAP may be achieved in vitro using the TNT rabbit reticulocyte lysate or wheat germ extract system (Promega). These systems couple transcription and translation of protein-coding sequences operably associated with the T7, T3, or SP6 promoters. Translation takes place in the presence of a radiolabeled amino acid precursor, for example, ³⁵S-methionine.

NAAP, fragments of NAAP, or variants of NAAP may be used to screen for compounds that specifically bind to NAAP. One or more test compounds may be screened for specific binding to NAAP. In various embodiments, 1, 2, 3, 4, 5, 10, 20, 50, 100, or 200 test compounds can be screened for specific binding to NAAP. Examples of test compounds can include antibodies, anticalins, oligonucleotides, proteins (e.g., ligands or receptors), or small molecules.

In related embodiments, variants of NAAP can be used to screen for binding of test

compounds, such as antibodies, to NAAP, a variant of NAAP, or a combination of NAAP and/or one or more variants NAAP. In an embodiment, a variant of NAAP can be used to screen for compounds that bind to a variant of NAAP, but not to NAAP having the exact sequence of a sequence of SEQ ID NO:1-57. NAAP variants used to perform such screening can have a range of about 50% to about 99% sequence identity to NAAP, with various embodiments having 60%, 70%, 75%, 80%, 85%, 90%, and 95% sequence identity.

In an embodiment, a compound identified in a screen for specific binding to NAAP can be closely related to the natural ligand of NAAP, e.g., a ligand or fragment thereof, a natural substrate, a structural or functional mimetic, or a natural binding partner (Coligan, J.E. et al. (1991) <u>Current Protocols in Immunology</u> 1(2):Chapter 5). In another embodiment, the compound thus identified can be a natural ligand of a receptor NAAP (Howard, A.D. et al. (2001) Trends Pharmacol. Sci.22:132-140; Wise, A. et al. (2002) Drug Discovery Today 7:235-246).

In other embodiments, a compound identified in a screen for specific binding to NAAP can be closely related to the natural receptor to which NAAP binds, at least a fragment of the receptor, or a fragment of the receptor including all or a portion of the ligand binding site or binding pocket. For example, the compound may be a receptor for NAAP which is capable of propagating a signal, or a decoy receptor for NAAP which is not capable of propagating a signal (Ashkenazi, A. and V.M. Divit (1999) Curr. Opin. Cell Biol. 11:255-260; Mantovani, A. et al. (2001) Trends Immunol. 22:328-336). The compound can be rationally designed using known techniques. Examples of such techniques include those used to construct the compound etanercept (ENBREL; Amgen Inc., Thousand Oaks CA), which is efficacious for treating rheumatoid arthritis in humans. Etanercept is an engineered p75 tumor necrosis factor (TNF) receptor dimer linked to the Fc portion of human IgG₁ (Taylor, P.C. et al. (2001) Curr. Opin. Immunol. 13:611-616).

In one embodiment, two or more antibodies having similar or, alternatively, different specificities can be screened for specific binding to NAAP, fragments of NAAP, or variants of NAAP. The binding specificity of the antibodies thus screened can thereby be selected to identify particular fragments or variants of NAAP. In one embodiment, an antibody can be selected such that its binding specificity allows for preferential identification of specific fragments or variants of NAAP. In another embodiment, an antibody can be selected such that its binding specificity allows for preferential diagnosis of a specific disease or condition having increased, decreased, or otherwise abnormal production of NAAP.

In an embodiment, anticalins can be screened for specific binding to NAAP, fragments of

NAAP, or variants of NAAP. Anticalins are ligand-binding proteins that have been constructed based on a lipocalin scaffold (Weiss, G.A. and H.B. Lowman (2000) Chem. Biol. 7:R177-R184; Skerra, A. (2001) J. Biotechnol. 74:257-275). The protein architecture of lipocalins can include a beta-barrel having eight antiparallel beta-strands, which supports four loops at its open end. These loops form the natural ligand-binding site of the lipocalins, a site which can be re-engineered in vitro by amino acid substitutions to impart novel binding specificities. The amino acid substitutions can be made using methods known in the art or described herein, and can include conservative substitutions (e.g., substitutions that do not alter binding specificity) or substitutions that modestly, moderately, or significantly alter binding specificity.

In one embodiment, screening for compounds which specifically bind to, stimulate, or inhibit NAAP involves producing appropriate cells which express NAAP, either as a secreted protein or on the cell membrane. Preferred cells can include cells from mammals, yeast, *Drosophila*, or *E. coli*. Cells expressing NAAP or cell membrane fractions which contain NAAP are then contacted with a test compound and binding, stimulation, or inhibition of activity of either NAAP or the compound is analyzed.

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An assay may simply test binding of a test compound to the polypeptide, wherein binding is detected by a fluorophore, radioisotope, enzyme conjugate, or other detectable label. For example, the assay may comprise the steps of combining at least one test compound with NAAP, either in solution or affixed to a solid support, and detecting the binding of NAAP to the compound. Alternatively, the assay may detect or measure binding of a test compound in the presence of a labeled competitor. Additionally, the assay may be carried out using cell-free preparations, chemical libraries, or natural product mixtures, and the test compound(s) may be free in solution or affixed to a solid support.

An assay can be used to assess the ability of a compound to bind to its natural ligand and/or to inhibit the binding of its natural ligand to its natural receptors. Examples of such assays include radio-labeling assays such as those described in U.S. Patent No. 5,914,236 and U.S. Patent No. 6,372,724. In a related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a receptor) to improve or alter its ability to bind to its natural ligands (Matthews, D.J. and J.A. Wells. (1994) Chem. Biol. 1:25-30). In another related embodiment, one or more amino acid substitutions can be introduced into a polypeptide compound (such as a ligand) to improve or alter its ability to bind to its natural receptors (Cunningham, B.C. and J.A. Wells (1991) Proc. Natl. Acad. Sci. USA 88:3407-3411; Lowman, H.B. et al. (1991) J. Biol. Chem. 266:10982-10988).

NAAP, fragments of NAAP, or variants of NAAP may be used to screen for compounds

that modulate the activity of NAAP. Such compounds may include agonists, antagonists, or partial or inverse agonists. In one embodiment, an assay is performed under conditions permissive for NAAP activity, wherein NAAP is combined with at least one test compound, and the activity of NAAP in the presence of a test compound is compared with the activity of NAAP in the absence of the test compound. A change in the activity of NAAP in the presence of the test compound is indicative of a compound that modulates the activity of NAAP. Alternatively, a test compound is combined with an in vitro or cell-free system comprising NAAP under conditions suitable for NAAP activity, and the assay is performed. In either of these assays, a test compound which modulates the activity of NAAP may do so indirectly and need not come in direct contact with the test compound. At least one and up to a plurality of test compounds may be screened.

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In another embodiment, polynucleotides encoding NAAP or their mammalian homologs may be "knocked out" in an animal model system using homologous recombination in embryonic stem (ES) cells. Such techniques are well known in the art and are useful for the generation of animal models of human disease (see, e.g., U.S. Patent No. 5,175,383 and U.S. Patent No. 5,767,337). For example, mouse ES cells, such as the mouse 129/SvJ cell line, are derived from the early mouse embryo and grown in culture. The ES cells are transformed with a vector containing the gene of interest disrupted by a marker gene, e.g., the neomycin phosphotransferase gene (neo; Capecchi, M.R. (1989) Science 244:1288-1292). The vector integrates into the corresponding region of the host genome by homologous recombination. Alternatively, homologous recombination takes place using the Cre-loxP system to knockout a gene of interest in a tissue- or developmental stage-specific manner (Marth, J.D. (1996) Clin. Invest. 97:1999-2002; Wagner, K.U. et al. (1997) Nucleic Acids Res. 25:4323-4330). Transformed ES cells are identified and microinjected into mouse cell blastocysts such as those from the C57BL/6 mouse strain. The blastocysts are surgically transferred to pseudopregnant dams, and the resulting chimeric progeny are genotyped and bred to produce heterozygous or homozygous strains. Transgenic animals thus generated may be tested with potential therapeutic or toxic agents.

Polynucleotides encoding NAAP may also be manipulated *in vitro* in ES cells derived from human blastocysts. Human ES cells have the potential to differentiate into at least eight separate cell lineages including endoderm, mesoderm, and ectodermal cell types. These cell lineages differentiate into, for example, neural cells, hematopoietic lineages, and cardiomyocytes (Thomson, J.A. et al. (1998) Science 282:1145-1147).

Polynucleotides encoding NAAP can also be used to create "knockin" humanized animals (pigs) or transgenic animals (mice or rats) to model human disease. With knockin technology, a region

of a polynucleotide encoding NAAP is injected into animal ES cells, and the injected sequence integrates into the animal cell genome. Transformed cells are injected into blastulae, and the blastulae are implanted as described above. Transgenic progeny or inbred lines are studied and treated with potential pharmaceutical agents to obtain information on treatment of a human disease. Alternatively, a mammal inbred to overexpress NAAP, e.g., by secreting NAAP in its milk, may also serve as a convenient source of that protein (Janne, J. et al. (1998) Biotechnol. Annu. Rev. 4:55-74).

THERAPEUTICS

Chemical and structural similarity, e.g., in the context of sequences and motifs, exists between regions of NAAP and nucleic acid-associated proteins. In addition, examples of tissues expressing NAAP can be found in Table 6 and can also be found in Example XI. Therefore, NAAP appears to play a role in cell proliferative, neurological, developmental, and autoimmune/inflammatory disorders, and infections. In the treatment of disorders associated with increased NAAP expression or activity, it is desirable to decrease the expression or activity of NAAP. In the treatment of disorders associated with decreased NAAP expression or activity, it is desirable to increase the expression or activity of NAAP.

Therefore, in one embodiment, NAAP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of NAAP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, a cancer of the adrenal gland, bladder, bone, bone marrow, brain, breast, cervix, colon, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease, Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial

insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorder of the central nervous system, cerebral palsy, a neuroskeletal disorder, an autonomic nervous system disorder, a cranial nerve disorder, a spinal cord disease, muscular dystrophy and other neuromuscular disorder, a peripheral nervous system disorder, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathy, myasthenia gravis, periodic paralysis, a mental disorder including mood, anxiety, and schizophrenic disorder, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, and Tourette's disorder; a developmental disorder such as renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism, hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis, glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and an infection, such as those caused by a viral agent classified as adenovirus, arenavirus, bunyavirus, calicivirus, coronavirus, filovirus, hepadnavirus, herpesvirus, flavivirus, orthomyxovirus, parvovirus, papovavirus, paramyxovirus, picornavirus, poxvirus, reovirus, retrovirus, rhabdovirus, or togavirus; an infection

caused by a bacterial agent classified as pneumococcus, staphylococcus, streptococcus, bacillus, corynebacterium, clostridium, meningococcus, gonococcus, listeria, moraxella, kingella, haemophilus, legionella, bordetella, gram-negative enterobacterium including shigella, salmonella, or campylobacter, pseudomonas, vibrio, brucella, francisella, yersinia, bartonella, norcardium, actinomyces, mycobacterium, spirochaetale, rickettsia, chlamydia, or mycoplasma; an infection caused by a fungal agent classified as aspergillus, blastomyces, dermatophytes, cryptococcus, coccidioides, malasezzia, histoplasma, or other mycosis-causing fungal agent; and an infection caused by a parasite classified as plasmodium or malaria-causing, parasitic entamoeba, leishmania, trypanosoma, toxoplasma, pneumocystis carinii, intestinal protozoa such as giardia, trichomonas, tissue nematode such as trichinella, intestinal nematode such as ascaris, lymphatic filarial nematode, trematode such as schistosoma, and cestode such as tapeworm.

In another embodiment, a vector capable of expressing NAAP or a fragment or derivative thereof may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of NAAP including, but not limited to, those described above.

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In a further embodiment, a composition comprising a substantially purified NAAP in conjunction with a suitable pharmaceutical carrier may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of NAAP including, but not limited to, those provided above.

In still another embodiment, an agonist which modulates the activity of NAAP may be administered to a subject to treat or prevent a disorder associated with decreased expression or activity of NAAP including, but not limited to, those listed above.

In a further embodiment, an antagonist of NAAP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of NAAP. Examples of such disorders include, but are not limited to, those cell proliferative, neurological, developmental, and autoimmune/inflammatory disorders, and infections described above. In one aspect, an antibody which specifically binds NAAP may be used directly as an antagonist or indirectly as a targeting or delivery mechanism for bringing a pharmaceutical agent to cells or tissues which express NAAP.

In an additional embodiment, a vector expressing the complement of the polynucleotide encoding NAAP may be administered to a subject to treat or prevent a disorder associated with increased expression or activity of NAAP including, but not limited to, those described above.

In other embodiments, any protein, agonist, antagonist, antibody, complementary sequence, or vector embodiments may be administered in combination with other appropriate therapeutic agents.

Selection of the appropriate agents for use in combination therapy may be made by one of ordinary skill in the art, according to conventional pharmaceutical principles. The combination of therapeutic agents may act synergistically to effect the treatment or prevention of the various disorders described above. Using this approach, one may be able to achieve therapeutic efficacy with lower dosages of each agent, thus reducing the potential for adverse side effects.

An antagonist of NAAP may be produced using methods which are generally known in the art. In particular, purified NAAP may be used to produce antibodies or to screen libraries of pharmaceutical agents to identify those which specifically bind NAAP. Antibodies to NAAP may also be generated using methods that are well known in the art. Such antibodies may include, but are not limited to, polyclonal, monoclonal, chimeric, and single chain antibodies, Fab fragments, and fragments produced by a Fab expression library. In an embodiment, neutralizing antibodies (i.e., those which inhibit dimer formation) can be used therapeutically. Single chain antibodies (e.g., from camels or llamas) may be potent enzyme inhibitors and may have application in the design of peptide mimetics, and in the development of immuno-adsorbents and biosensors (Muyldermans, S. (2001) J. Biotechnol. 74:277-302).

For the production of antibodies, various hosts including goats, rabbits, rats, mice, camels, dromedaries, llamas, humans, and others may be immunized by injection with NAAP or with any fragment or oligopeptide thereof which has immunogenic properties. Depending on the host species, various adjuvants may be used to increase immunological response. Such adjuvants include, but are not limited to, Freund's, mineral gels such as aluminum hydroxide, and surface active substances such as lysolecithin, pluronic polyols, polyanions, peptides, oil emulsions, KLH, and dinitrophenol. Among adjuvants used in humans, BCG (bacilli Calmette-Guerin) and *Corynebacterium parvum* are especially preferable.

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It is preferred that the oligopeptides, peptides, or fragments used to induce antibodies to NAAP have an amino acid sequence consisting of at least about 5 amino acids, and generally will consist of at least about 10 amino acids. It is also preferable that these oligopeptides, peptides, or fragments are substantially identical to a portion of the amino acid sequence of the natural protein. Short stretches of NAAP amino acids may be fused with those of another protein, such as KLH, and antibodies to the chimeric molecule may be produced.

Monoclonal antibodies to NAAP may be prepared using any technique which provides for the production of antibody molecules by continuous cell lines in culture. These include, but are not limited to, the hybridoma technique, the human B-cell hybridoma technique, and the EBV-hybridoma

technique (Kohler, G. et al. (1975) Nature 256:495-497; Kozbor, D. et al. (1985) J. Immunol. Methods 81:31-42; Cote, R.J. et al. (1983) Proc. Natl. Acad. Sci. USA 80:2026-2030; Cole, S.P. et al. (1984) Mol. Cell Biol. 62:109-120).

In addition, techniques developed for the production of "chimeric antibodies," such as the splicing of mouse antibody genes to human antibody genes to obtain a molecule with appropriate antigen specificity and biological activity, can be used (Morrison, S.L. et al. (1984) Proc. Natl. Acad. Sci. USA 81:6851-6855; Neuberger, M.S. et al. (1984) Nature 312:604-608; Takeda, S. et al. (1985) Nature 314:452-454). Alternatively, techniques described for the production of single chain antibodies may be adapted, using methods known in the art, to produce NAAP-specific single chain antibodies. Antibodies with related specificity, but of distinct idiotypic composition, may be generated by chain shuffling from random combinatorial immunoglobulin libraries (Burton, D.R. (1991) Proc. Natl. Acad. Sci. USA 88:10134-10137).

Antibodies may also be produced by inducing *in vivo* production in the lymphocyte population or by screening immunoglobulin libraries or panels of highly specific binding reagents as disclosed in the literature (Orlandi, R. et al. (1989) Proc. Natl. Acad. Sci. USA 86:3833-3837; Winter, G. et al. (1991) Nature 349:293-299).

Antibody fragments which contain specific binding sites for NAAP may also be generated. For example, such fragments include, but are not limited to, $F(ab')_2$ fragments produced by pepsin digestion of the antibody molecule and Fab fragments generated by reducing the disulfide bridges of the $F(ab')_2$ fragments. Alternatively, Fab expression libraries may be constructed to allow rapid and easy identification of monoclonal Fab fragments with the desired specificity (Huse, W.D. et al. (1989) Science 246:1275-1281).

Various immunoassays may be used for screening to identify antibodies having the desired specificity. Numerous protocols for competitive binding or immunoradiometric assays using either polyclonal or monoclonal antibodies with established specificities are well known in the art. Such immunoassays typically involve the measurement of complex formation between NAAP and its specific antibody. A two-site, monoclonal-based immunoassay utilizing monoclonal antibodies reactive to two non-interfering NAAP epitopes is generally used, but a competitive binding assay may also be employed (Pound, *supra*).

Various methods such as Scatchard analysis in conjunction with radioimmunoassay techniques may be used to assess the affinity of antibodies for NAAP. Affinity is expressed as an association constant, K_a, which is defined as the molar concentration of NAAP-antibody complex divided by the

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molar concentrations of free antigen and free antibody under equilibrium conditions. The K_a determined for a preparation of polyclonal antibodies, which are heterogeneous in their affinities for multiple NAAP epitopes, represents the average affinity, or avidity, of the antibodies for NAAP. The K_a determined for a preparation of monoclonal antibodies, which are monospecific for a particular NAAP epitope, represents a true measure of affinity. High-affinity antibody preparations with K_a ranging from about 10⁹ to 10¹² L/mole are preferred for use in immunoassays in which the NAAP-antibody complex must withstand rigorous manipulations. Low-affinity antibody preparations with K_a ranging from about 10⁶ to 10⁷ L/mole are preferred for use in immunopurification and similar procedures which ultimately require dissociation of NAAP, preferably in active form, from the antibody (Catty, D. (1988) Antibodies, Volume I: A Practical Approach, IRL Press, Washington DC; Liddell, J.E. and A. Cryer (1991) A Practical Guide to Monoclonal Antibodies, John Wiley & Sons, New York NY).

The titer and avidity of polyclonal antibody preparations may be further evaluated to determine the quality and suitability of such preparations for certain downstream applications. For example, a polyclonal antibody preparation containing at least 1-2 mg specific antibody/ml, preferably 5-10 mg specific antibody/ml, is generally employed in procedures requiring precipitation of NAAP-antibody complexes. Procedures for evaluating antibody specificity, titer, and avidity, and guidelines for antibody quality and usage in various applications, are generally available (Catty, *supra*; Coligan et al., *supra*).

In another embodiment of the invention, polynucleotides encoding NAAP, or any fragment or complement thereof, may be used for therapeutic purposes. In one aspect, modifications of gene expression can be achieved by designing complementary sequences or antisense molecules (DNA, RNA, PNA, or modified oligonucleotides) to the coding or regulatory regions of the gene encoding NAAP. Such technology is well known in the art, and antisense oligonucleotides or larger fragments can be designed from various locations along the coding or control regions of sequences encoding NAAP (Agrawal, S., ed. (1996) Antisense Therapeutics, Humana Press, Totawa NJ).

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In therapeutic use, any gene delivery system suitable for introduction of the antisense sequences into appropriate target cells can be used. Antisense sequences can be delivered intracellularly in the form of an expression plasmid which, upon transcription, produces a sequence complementary to at least a portion of the cellular sequence encoding the target protein (Slater, J.E. et al. (1998) J. Allergy Clin. Immunol. 102:469-475; Scanlon, K.J. et al. (1995) 9:1288-1296). Antisense sequences can also be introduced intracellularly through the use of viral vectors, such as retrovirus and

adeno-associated virus vectors (Miller, A.D. (1990) Blood 76:271; Ausubel et al., *supra*; Uckert, W. and W. Walther (1994) Pharmacol. Ther. 63:323-347). Other gene delivery mechanisms include liposome-derived systems, artificial viral envelopes, and other systems known in the art (Rossi, J.J. (1995) Br. Med. Bull. 51:217-225; Boado, R.J. et al. (1998) J. Pharm. Sci. 87:1308-1315; Morris, M.C. et al. (1997) Nucleic Acids Res. 25:2730-2736).

In another embodiment of the invention, polynucleotides encoding NAAP may be used for somatic or germline gene therapy. Gene therapy may be performed to (i) correct a genetic deficiency (e.g., in the cases of severe combined immunodeficiency (SCID)-X1 disease characterized by Xlinked inheritance (Cavazzana-Calvo, M. et al. (2000) Science 288:669-672), severe combined immunodeficiency syndrome associated with an inherited adenosine deaminase (ADA) deficiency (Blaese, R.M. et al. (1995) Science 270:475-480; Bordignon, C. et al. (1995) Science 270:470-475), cystic fibrosis (Zabner, J. et al. (1993) Cell 75:207-216; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:643-666; Crystal, R.G. et al. (1995) Hum. Gene Therapy 6:667-703), thalassamias, familial hypercholesterolemia, and hemophilia resulting from Factor VIII or Factor IX deficiencies (Crystal, R.G. (1995) Science 270:404-410; Verma, I.M. and N. Somia (1997) Nature 389:239-242)), (ii) express a conditionally lethal gene product (e.g., in the case of cancers which result from unregulated cell proliferation), or (iii) express a protein which affords protection against intracellular parasites (e.g., against human retroviruses, such as human immunodeficiency virus (HIV) (Baltimore, D. (1988) Nature 335:395-396; Poeschla, E. et al. (1996) Proc. Natl. Acad. Sci. USA 93:11395-11399), hepatitis B or C virus (HBV, HCV); fungal parasites, such as Candida albicans and Paracoccidioides brasiliensis; and protozoan parasites such as Plasmodium falciparum and Trypanosoma cruzi). In the case where a genetic deficiency in NAAP expression or regulation causes disease, the expression of NAAP from an appropriate population of transduced cells may alleviate the clinical manifestations caused by the genetic deficiency.

In a further embodiment of the invention, diseases or disorders caused by deficiencies in NAAP are treated by constructing mammalian expression vectors encoding NAAP and introducing these vectors by mechanical means into NAAP-deficient cells. Mechanical transfer technologies for use with cells *in vivo* or *ex vitro* include (i) direct DNA microinjection into individual cells, (ii) ballistic gold particle delivery, (iii) liposome-mediated transfection, (iv) receptor-mediated gene transfer, and (v) the use of DNA transposons (Morgan, R.A. and W.F. Anderson (1993) Annu. Rev. Biochem. 62:191-217; Ivics, Z. (1997) Cell 91:501-510; Boulay, J.-L. and H. Récipon (1998) Curr. Opin. Biotechnol. 9:445-450).

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Expression vectors that may be effective for the expression of NAAP include, but are not limited to, the PCDNA 3.1, EPITAG, PRCCMV2, PREP, PVAX, PCR2-TOPOTA vectors (Invitrogen, Carlsbad CA), PCMV-SCRIPT, PCMV-TAG, PEGSH/PERV (Stratagene, La Jolla CA), and PTET-OFF, PTET-ON, PTRE2, PTRE2-LUC, PTK-HYG (Clontech, Palo Alto CA). NAAP may be expressed using (i) a constitutively active promoter, (e.g., from cytomegalovirus (CMV), Rous sarcoma virus (RSV), SV40 virus, thymidine kinase (TK), or β-actin genes), (ii) an inducible promoter (e.g., the tetracycline-regulated promoter (Gossen, M. and H. Bujard (1992) Proc. Natl. Acad. Sci. USA 89:5547-5551; Gossen, M. et al. (1995) Science 268:1766-1769; Rossi, F.M.V. and H.M. Blau (1998) Curr. Opin. Biotechnol. 9:451-456), commercially available in the T-REX plasmid (Invitrogen)); the ecdysone-inducible promoter (available in the plasmids PVGRXR and PIND; Invitrogen); the FK506/rapamycin inducible promoter; or the RU486/mifepristone inducible promoter (Rossi, F.M.V. and H.M. Blau, supra)), or (iii) a tissue-specific promoter or the native promoter of the endogenous gene encoding NAAP from a normal individual.

Commercially available liposome transformation kits (e.g., the PERFECT LIPID TRANSFECTION KIT, available from Invitrogen) allow one with ordinary skill in the art to deliver polynucleotides to target cells in culture and require minimal effort to optimize experimental parameters. In the alternative, transformation is performed using the calcium phosphate method (Graham, F.L. and A.J. Eb (1973) Virology 52:456-467), or by electroporation (Neumann, E. et al. (1982) EMBO J. 1:841-845). The introduction of DNA to primary cells requires modification of these standardized mammalian transfection protocols.

In another embodiment of the invention, diseases or disorders caused by genetic defects with respect to NAAP expression are treated by constructing a retrovirus vector consisting of (i) the polynucleotide encoding NAAP under the control of an independent promoter or the retrovirus long terminal repeat (LTR) promoter, (ii) appropriate RNA packaging signals, and (iii) a Rev-responsive element (RRE) along with additional retrovirus cis-acting RNA sequences and coding sequences required for efficient vector propagation. Retrovirus vectors (e.g., PFB and PFBNEO) are commercially available (Stratagene) and are based on published data (Riviere, I. et al. (1995) Proc. Natl. Acad. Sci. USA 92:6733-6737), incorporated by reference herein. The vector is propagated in an appropriate vector producing cell line (VPCL) that expresses an envelope gene with a tropism for receptors on the target cells or a promiscuous envelope protein such as VSVg (Armentano, D. et al. (1987) J. Virol. 61:1647-1650; Bender, M.A. et al. (1987) J. Virol. 61:1639-1646; Adam, M.A. and A.D. Miller (1988) J. Virol. 62:3802-3806; Dull, T. et al. (1998) J. Virol. 72:8463-8471; Zufferey, R. et

al. (1998) J. Virol. 72:9873-9880). U.S. Patent No. 5,910,434 to Rigg ("Method for obtaining retrovirus packaging cell lines producing high transducing efficiency retroviral supernatant") discloses a method for obtaining retrovirus packaging cell lines and is hereby incorporated by reference. Propagation of retrovirus vectors, transduction of a population of cells (e.g., CD4+ T-cells), and the return of transduced cells to a patient are procedures well known to persons skilled in the art of gene therapy and have been well documented (Ranga, U. et al. (1997) J. Virol. 71:7020-7029; Bauer, G. et al. (1997) Blood 89:2259-2267; Bonyhadi, M.L. (1997) J. Virol. 71:4707-4716; Ranga, U. et al. (1998) Proc. Natl. Acad. Sci. USA 95:1201-1206; Su, L. (1997) Blood 89:2283-2290).

In an embodiment, an adenovirus-based gene therapy delivery system is used to deliver polynucleotides encoding NAAP to cells which have one or more genetic abnormalities with respect to the expression of NAAP. The construction and packaging of adenovirus-based vectors are well known to those with ordinary skill in the art. Replication defective adenovirus vectors have proven to be versatile for importing genes encoding immunoregulatory proteins into intact islets in the pancreas (Csete, M.E. et al. (1995) Transplantation 27:263-268). Potentially useful adenoviral vectors are described in U.S. Patent No. 5,707,618 to Armentano ("Adenovirus vectors for gene therapy"), hereby incorporated by reference. For adenoviral vectors, see also Antinozzi, P.A. et al. (1999; Annu. Rev. Nutr. 19:511-544) and Verma, I.M. and N. Somia (1997; Nature 18:389:239-242).

In another embodiment, a herpes-based, gene therapy delivery system is used to deliver polynucleotides encoding NAAP to target cells which have one or more genetic abnormalities with respect to the expression of NAAP. The use of herpes simplex virus (HSV)-based vectors may be especially valuable for introducing NAAP to cells of the central nervous system, for which HSV has a tropism. The construction and packaging of herpes-based vectors are well known to those with ordinary skill in the art. A replication-competent herpes simplex virus (HSV) type 1-based vector has been used to deliver a reporter gene to the eyes of primates (Liu, X. et al. (1999) Exp. Eye Res. 169:385-395). The construction of a HSV-1 virus vector has also been disclosed in detail in U.S. Patent No. 5,804,413 to DeLuca ("Herpes simplex virus strains for gene transfer"), which is hereby incorporated by reference. U.S. Patent No. 5,804,413 teaches the use of recombinant HSV d92 which consists of a genome containing at least one exogenous gene to be transferred to a cell under the control of the appropriate promoter for purposes including human gene therapy. Also taught by this patent are the construction and use of recombinant HSV strains deleted for ICP4, ICP27 and ICP22. For HSV vectors, see also Goins, W.F. et al. (1999; J. Virol. 73:519-532) and Xu, H. et al. (1994; Dev. Biol. 163:152-161). The manipulation of cloned herpesvirus sequences, the generation of

recombinant virus following the transfection of multiple plasmids containing different segments of the large herpesvirus genomes, the growth and propagation of herpesvirus, and the infection of cells with herpesvirus are techniques well known to those of ordinary skill in the art.

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In another embodiment, an alphavirus (positive, single-stranded RNA virus) vector is used to deliver polynucleotides encoding NAAP to target cells. The biology of the prototypic alphavirus, Semliki Forest Virus (SFV), has been studied extensively and gene transfer vectors have been based on the SFV genome (Garoff, H. and K.-J. Li (1998) Curr. Opin. Biotechnol. 9:464-469). During alphavirus RNA replication, a subgenomic RNA is generated that normally encodes the viral capsid proteins. This subgenomic RNA replicates to higher levels than the full length genomic RNA, resulting in the overproduction of capsid proteins relative to the viral proteins with enzymatic activity (e.g., protease and polymerase). Similarly, inserting the coding sequence for NAAP into the alphavirus genome in place of the capsid-coding region results in the production of a large number of NAAP-coding RNAs and the synthesis of high levels of NAAP in vector transduced cells. While alphavirus infection is typically associated with cell lysis within a few days, the ability to establish a persistent infection in hamster normal kidney cells (BHK-21) with a variant of Sindbis virus (SIN) indicates that the lytic replication of alphaviruses can be altered to suit the needs of the gene therapy application (Dryga, S.A. et al. (1997) Virology 228:74-83). The wide host range of alphaviruses will allow the introduction of NAAP into a variety of cell types. The specific transduction of a subset of cells in a population may require the sorting of cells prior to transduction. The methods of manipulating infectious cDNA clones of alphaviruses, performing alphavirus cDNA and RNA transfections, and performing alphavirus infections, are well known to those with ordinary skill in the art.

Oligonucleotides derived from the transcription initiation site, e.g., between about positions -10 and +10 from the start site, may also be employed to inhibit gene expression. Similarly, inhibition can be achieved using triple helix base-pairing methodology. Triple helix pairing is useful because it causes inhibition of the ability of the double helix to open sufficiently for the binding of polymerases, transcription factors, or regulatory molecules. Recent therapeutic advances using triplex DNA have been described in the literature (Gee, J.E. et al. (1994) in Huber, B.E. and B.I. Carr, Molecular and Immunologic Approaches, Futura Publishing, Mt. Kisco NY, pp. 163-177). A complementary sequence or antisense molecule may also be designed to block translation of mRNA by preventing the transcript from binding to ribosomes.

Ribozymes, enzymatic RNA molecules, may also be used to catalyze the specific cleavage of

RNA. The mechanism of ribozyme action involves sequence-specific hybridization of the ribozyme molecule to complementary target RNA, followed by endonucleolytic cleavage. For example, engineered hammerhead motif ribozyme molecules may specifically and efficiently catalyze endonucleolytic cleavage of RNA molecules encoding NAAP.

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Specific ribozyme cleavage sites within any potential RNA target are initially identified by scanning the target molecule for ribozyme cleavage sites, including the following sequences: GUA, GUU, and GUC. Once identified, short RNA sequences of between 15 and 20 ribonucleotides, corresponding to the region of the target gene containing the cleavage site, may be evaluated for secondary structural features which may render the oligonucleotide inoperable. The suitability of candidate targets may also be evaluated by testing accessibility to hybridization with complementary oligonucleotides using ribonuclease protection assays.

Complementary ribonucleic acid molecules and ribozymes may be prepared by any method known in the art for the synthesis of nucleic acid molecules. These include techniques for chemically synthesizing oligonucleotides such as solid phase phosphoramidite chemical synthesis. Alternatively, RNA molecules may be generated by *in vitro* and *in vivo* transcription of DNA molecules encoding NAAP. Such DNA sequences may be incorporated into a wide variety of vectors with suitable RNA polymerase promoters such as T7 or SP6. Alternatively, these cDNA constructs that synthesize complementary RNA, constitutively or inducibly, can be introduced into cell lines, cells, or tissues.

RNA molecules may be modified to increase intracellular stability and half-life. Possible modifications include, but are not limited to, the addition of flanking sequences at the 5' and/or 3' ends of the molecule, or the use of phosphorothioate or 2' O-methyl rather than phosphodiesterase linkages within the backbone of the molecule. This concept is inherent in the production of PNAs and can be extended in all of these molecules by the inclusion of nontraditional bases such as inosine, queosine, and wybutosine, as well as acetyl-, methyl-, thio-, and similarly modified forms of adenine, cytidine, guanine, thymine, and uridine which are not as easily recognized by endogenous endonucleases.

In other embodiments of the invention, the expression of one or more selected polynucleotides of the present invention can be altered, inhibited, decreased, or silenced using RNA interference (RNAi) or post-transcriptional gene silencing (PTGS) methods known in the art. RNAi is a post-transcriptional mode of gene silencing in which double-stranded RNA (dsRNA) introduced into a targeted cell specifically suppresses the expression of the homologous gene (i.e., the gene bearing the sequence complementary to the dsRNA). This effectively knocks out or substantially reduces the expression of the targeted gene. PTGS can also be accomplished by use of DNA or DNA fragments

as well. RNAi methods are described by Fire, A. et al. (1998; Nature 391:806-811) and Gura, T. (2000; Nature 404:804-808). PTGS can also be initiated by introduction of a complementary segment of DNA into the selected tissue using gene delivery and/or viral vector delivery methods described herein or known in the art.

RNAi can be induced in mammalian cells by the use of small interfering RNA also known as siRNA. SiRNA are shorter segments of dsRNA (typically about 21 to 23 nucleotides in length) that result *in vivo* from cleavage of introduced dsRNA by the action of an endogenous ribonuclease. SiRNA appear to be the mediators of the RNAi effect in mammals. The most effective siRNAs appear to be 21 nucleotide dsRNAs with 2 nucleotide 3' overhangs. The use of siRNA for inducing RNAi in mammalian cells is described by Elbashir, S.M. et al. (2001; Nature 411:494-498).

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SiRNA can either be generated indirectly by introduction of dsRNA into the targeted cell, or directly by mammalian transfection methods and agents described herein or known in the art (such as liposome-mediated transfection, viral vector methods, or other polynucleotide delivery/introductory methods). Suitable SiRNAs can be selected by examining a transcript of the target polynucleotide (e.g., mRNA) for nucleotide sequences downstream from the AUG start codon and recording the occurrence of each nucleotide and the 3' adjacent 19 to 23 nucleotides as potential siRNA target sites, with sequences having a 21 nucleotide length being preferred. Regions to be avoided for target siRNA sites include the 5' and 3' untranslated regions (UTRs) and regions near the start codon (within 75 bases), as these may be richer in regulatory protein binding sites. UTR-binding proteins and/or translation initiation complexes may interfere with binding of the siRNP endonuclease complex. The selected target sites for siRNA can then be compared to the appropriate genome database (e.g., human, etc.) using BLAST or other sequence comparison algorithms known in the art. Target sequences with significant homology to other coding sequences can be eliminated from consideration. The selected SiRNAs can be produced by chemical synthesis methods known in the art or by in vitro transcription using commercially available methods and kits such as the SILENCER siRNA construction kit (Ambion, Austin TX).

In alternative embodiments, long-term gene silencing and/or RNAi effects can be induced in selected tissue using expression vectors that continuously express siRNA. This can be accomplished using expression vectors that are engineered to express hairpin RNAs (shRNAs) using methods known in the art (see, e.g., Brummelkamp, T.R. et al. (2002) Science 296:550-553; and Paddison, P.J. et al. (2002) Genes Dev. 16:948-958). In these and related embodiments, shRNAs can be delivered to target cells using expression vectors known in the art. An example of a suitable expression vector for

delivery of siRNA is the PSILENCER1.0-U6 (circular) plasmid (Ambion). Once delivered to the target tissue, shRNAs are processed *in vivo* into siRNA-like molecules capable of carrying out genespecific silencing.

In various embodiments, the expression levels of genes targeted by RNAi or PTGS methods can be determined by assays for mRNA and/or protein analysis. Expression levels of the mRNA of a targeted gene, can be determined by northern analysis methods using, for example, the NORTHERNMAX-GLY kit (Ambion); by microarray methods; by PCR methods; by real time PCR methods; and by other RNA/polynucleotide assays known in the art or described herein. Expression levels of the protein encoded by the targeted gene can be determined by Western analysis using standard techniques known in the art.

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An additional embodiment of the invention encompasses a method for screening for a compound which is effective in altering expression of a polynucleotide encoding NAAP. Compounds which may be effective in altering expression of a specific polynucleotide may include, but are not limited to, oligonucleotides, antisense oligonucleotides, triple helix-forming oligonucleotides, transcription factors and other polypeptide transcriptional regulators, and non-macromolecular chemical entities which are capable of interacting with specific polynucleotide sequences. Effective compounds may alter polynucleotide expression by acting as either inhibitors or promoters of polynucleotide expression. Thus, in the treatment of disorders associated with increased NAAP expression or activity, a compound which specifically inhibits expression of the polynucleotide encoding NAAP may be therapeutically useful, and in the treatment of disorders associated with decreased NAAP expression or activity, a compound which specifically promotes expression of the polynucleotide encoding NAAP may be therapeutically useful.

In various embodiments, one or more test compounds may be screened for effectiveness in altering expression of a specific polynucleotide. A test compound may be obtained by any method commonly known in the art, including chemical modification of a compound known to be effective in altering polynucleotide expression; selection from an existing, commercially-available or proprietary library of naturally-occurring or non-natural chemical compounds; rational design of a compound based on chemical and/or structural properties of the target polynucleotide; and selection from a library of chemical compounds created combinatorially or randomly. A sample comprising a polynucleotide encoding NAAP is exposed to at least one test compound thus obtained. The sample may comprise, for example, an intact or permeabilized cell, or an *in vitro* cell-free or reconstituted biochemical system. Alterations in the expression of a polynucleotide encoding NAAP are assayed by

any method commonly known in the art. Typically, the expression of a specific nucleotide is detected by hybridization with a probe having a nucleotide sequence complementary to the sequence of the polynucleotide encoding NAAP. The amount of hybridization may be quantified, thus forming the basis for a comparison of the expression of the polynucleotide both with and without exposure to one or more test compounds. Detection of a change in the expression of a polynucleotide exposed to a test compound indicates that the test compound is effective in altering the expression of the polynucleotide. A screen for a compound effective in altering expression of a specific polynucleotide can be carried out, for example, using a Schizosaccharomyces pombe gene expression system (Atkins, D. et al. (1999) U.S. Patent No. 5,932,435; Arndt, G.M. et al. (2000) Nucleic Acids Res. 28:E15) or a human cell line such as HeLa cell (Clarke, M.L. et al. (2000) Biochem. Biophys. Res. Commun. 268:8-13). A particular embodiment of the present invention involves screening a combinatorial library of oligonucleotides (such as deoxyribonucleotides, ribonucleotides, peptide nucleic acids, and modified oligonucleotides) for antisense activity against a specific polynucleotide sequence (Bruice, T.W. et al. (1997) U.S. Patent No. 5,686,242; Bruice, T.W. et al. (2000) U.S. Patent No. 6,022,691).

Many methods for introducing vectors into cells or tissues are available and equally suitable for use in vivo, in vitro, and ex vivo. For ex vivo therapy, vectors may be introduced into stem cells taken from the patient and clonally propagated for autologous transplant back into that same patient. Delivery by transfection, by liposome injections, or by polycationic amino polymers may be achieved using methods which are well known in the art (Goldman, C.K. et al. (1997) Nat. Biotechnol. 15:462-466).

Any of the therapeutic methods described above may be applied to any subject in need of such therapy, including, for example, mammals such as humans, dogs, cats, cows, horses, rabbits, and monkeys.

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An additional embodiment of the invention relates to the administration of a composition which generally comprises an active ingredient formulated with a pharmaceutically acceptable excipient.

Excipients may include, for example, sugars, starches, celluloses, gums, and proteins. Various formulations are commonly known and are thoroughly discussed in the latest edition of Remington's Pharmaceutical Sciences (Maack Publishing, Easton PA). Such compositions may consist of NAAP, antibodies to NAAP, and mimetics, agonists, antagonists, or inhibitors of NAAP.

In various embodiments, the compositions described herein, such as pharmaceutical compositions, may be administered by any number of routes including, but not limited to, oral,

intravenous, intramuscular, intra-arterial, intramedullary, intrathecal, intraventricular, pulmonary, transdermal, subcutaneous, intraperitoneal, intranasal, enteral, topical, sublingual, or rectal means.

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Compositions for pulmonary administration may be prepared in liquid or dry powder form. These compositions are generally aerosolized immediately prior to inhalation by the patient. In the case of small molecules (e.g. traditional low molecular weight organic drugs), aerosol delivery of fast-acting formulations is well-known in the art. In the case of macromolecules (e.g. larger peptides and proteins), recent developments in the field of pulmonary delivery via the alveolar region of the lung have enabled the practical delivery of drugs such as insulin to blood circulation (see, e.g., Patton, J.S. et al., U.S. Patent No. 5,997,848). Pulmonary delivery allows administration without needle injection, and obviates the need for potentially toxic penetration enhancers.

Compositions suitable for use in the invention include compositions wherein the active ingredients are contained in an effective amount to achieve the intended purpose. The determination of an effective dose is well within the capability of those skilled in the art.

Specialized forms of compositions may be prepared for direct intracellular delivery of macromolecules comprising NAAP or fragments thereof. For example, liposome preparations containing a cell-impermeable macromolecule may promote cell fusion and intracellular delivery of the macromolecule. Alternatively, NAAP or a fragment thereof may be joined to a short cationic N-terminal portion from the HIV Tat-1 protein. Fusion proteins thus generated have been found to transduce into the cells of all tissues, including the brain, in a mouse model system (Schwarze, S.R. et al. (1999) Science 285:1569-1572).

For any compound, the therapeutically effective dose can be estimated initially either in cell culture assays, e.g., of neoplastic cells, or in animal models such as mice, rats, rabbits, dogs, monkeys, or pigs. An animal model may also be used to determine the appropriate concentration range and route of administration. Such information can then be used to determine useful doses and routes for administration in humans.

A therapeutically effective dose refers to that amount of active ingredient, for example NAAP or fragments thereof, antibodies of NAAP, and agonists, antagonists or inhibitors of NAAP, which ameliorates the symptoms or condition. Therapeutic efficacy and toxicity may be determined by standard pharmaceutical procedures in cell cultures or with experimental animals, such as by calculating the ED₅₀ (the dose therapeutically effective in 50% of the population) or LD₅₀ (the dose lethal to 50% of the population) statistics. The dose ratio of toxic to therapeutic effects is the therapeutic index, which can be expressed as the LD₅₀/ED₅₀ ratio. Compositions which exhibit large

therapeutic indices are preferred. The data obtained from cell culture assays and animal studies are used to formulate a range of dosage for human use. The dosage contained in such compositions is preferably within a range of circulating concentrations that includes the ED₅₀ with little or no toxicity. The dosage varies within this range depending upon the dosage form employed, the sensitivity of the patient, and the route of administration.

The exact dosage will be determined by the practitioner, in light of factors related to the subject requiring treatment. Dosage and administration are adjusted to provide sufficient levels of the active moiety or to maintain the desired effect. Factors which may be taken into account include the severity of the disease state, the general health of the subject, the age, weight, and gender of the subject, time and frequency of administration, drug combination(s), reaction sensitivities, and response to therapy. Long-acting compositions may be administered every 3 to 4 days, every week, or biweekly depending on the half-life and clearance rate of the particular formulation.

Normal dosage amounts may vary from about 0.1 μ g to 100,000 μ g, up to a total dose of about 1 gram, depending upon the route of administration. Guidance as to particular dosages and methods of delivery is provided in the literature and generally available to practitioners in the art. Those skilled in the art will employ different formulations for nucleotides than for proteins or their inhibitors. Similarly, delivery of polynucleotides or polypeptides will be specific to particular cells, conditions, locations, etc.

DIAGNOSTICS

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In another embodiment, antibodies which specifically bind NAAP may be used for the diagnosis of disorders characterized by expression of NAAP, or in assays to monitor patients being treated with NAAP or agonists, antagonists, or inhibitors of NAAP. Antibodies useful for diagnostic purposes may be prepared in the same manner as described above for therapeutics. Diagnostic assays for NAAP include methods which utilize the antibody and a label to detect NAAP in human body fluids or in extracts of cells or tissues. The antibodies may be used with or without modification, and may be labeled by covalent or non-covalent attachment of a reporter molecule. A wide variety of reporter molecules, several of which are described above, are known in the art and may be used.

A variety of protocols for measuring NAAP, including ELISAs, RIAs, and FACS, are known in the art and provide a basis for diagnosing altered or abnormal levels of NAAP expression. Normal or standard values for NAAP expression are established by combining body fluids or cell extracts taken from normal mammalian subjects, for example, human subjects, with antibodies to NAAP under conditions suitable for complex formation. The amount of standard complex formation may be

quantitated by various methods, such as photometric means. Quantities of NAAP expressed in subject, control, and disease samples from biopsied tissues are compared with the standard values. Deviation between standard and subject values establishes the parameters for diagnosing disease.

In another embodiment of the invention, polynucleotides encoding NAAP may be used for diagnostic purposes. The polynucleotides which may be used include oligonucleotides, complementary RNA and DNA molecules, and PNAs. The polynucleotides may be used to detect and quantify gene expression in biopsied tissues in which expression of NAAP may be correlated with disease. The diagnostic assay may be used to determine absence, presence, and excess expression of NAAP, and to monitor regulation of NAAP levels during therapeutic intervention.

In one aspect, hybridization with PCR probes which are capable of detecting polynucleotides, including genomic sequences, encoding NAAP or closely related molecules may be used to identify nucleic acid sequences which encode NAAP. The specificity of the probe, whether it is made from a highly specific region, e.g., the 5' regulatory region, or from a less specific region, e.g., a conserved motif, and the stringency of the hybridization or amplification will determine whether the probe identifies only naturally occurring sequences encoding NAAP, allelic variants, or related sequences.

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Probes may also be used for the detection of related sequences, and may have at least 50% sequence identity to any of the NAAP encoding sequences. The hybridization probes of the subject invention may be DNA or RNA and may be derived from the sequence of SEQ ID NO:58-114 or from genomic sequences including promoters, enhancers, and introns of the NAAP gene.

Means for producing specific hybridization probes for polynucleotides encoding NAAP include the cloning of polynucleotides encoding NAAP or NAAP derivatives into vectors for the production of mRNA probes. Such vectors are known in the art, are commercially available, and may be used to synthesize RNA probes *in vitro* by means of the addition of the appropriate RNA polymerases and the appropriate labeled nucleotides. Hybridization probes may be labeled by a variety of reporter groups, for example, by radionuclides such as ³²P or ³⁵S, or by enzymatic labels, such as alkaline phosphatase coupled to the probe via avidin/biotin coupling systems, and the like.

Polynucleotides encoding NAAP may be used for the diagnosis of disorders associated with expression of NAAP. Examples of such disorders include, but are not limited to, a cell proliferative disorder such as actinic keratosis, arteriosclerosis, atherosclerosis, bursitis, cirrhosis, hepatitis, mixed connective tissue disease (MCTD), myelofibrosis, paroxysmal nocturnal hemoglobinuria, polycythemia vera, psoriasis, primary thrombocythemia, and cancers including adenocarcinoma, leukemia, lymphoma, melanoma, myeloma, sarcoma, teratocarcinoma, and, in particular, a cancer of the adrenal

gland, bladder, bone, bone marrow, brain, breast, cervix, colon, gall bladder, ganglia, gastrointestinal tract, heart, kidney, liver, lung, muscle, ovary, pancreas, parathyroid, penis, prostate, salivary glands, skin, spleen, testis, thymus, thyroid, and uterus; a neurological disorder such as epilepsy, ischemic cerebrovascular disease, stroke, cerebral neoplasms, Alzheimer's disease, Pick's disease,

Huntington's disease, dementia, Parkinson's disease and other extrapyramidal disorders, amyotrophic lateral sclerosis and other motor neuron disorders, progressive neural muscular atrophy, retinitis pigmentosa, hereditary ataxias, multiple sclerosis and other demyelinating diseases, bacterial and viral meningitis, brain abscess, subdural empyema, epidural abscess, suppurative intracranial thrombophlebitis, myelitis and radiculitis, viral central nervous system disease, prion diseases including kuru, Creutzfeldt-Jakob disease, and Gerstmann-Straussler-Scheinker syndrome, fatal familial insomnia, nutritional and metabolic diseases of the nervous system, neurofibromatosis, tuberous sclerosis, cerebelloretinal hemangioblastomatosis, encephalotrigeminal syndrome, mental retardation and other developmental disorder of the central nervous system, cerebral palsy, a neuroskeletal disorder, an autonomic nervous system disorder, a cranial nerve disorder, a spinal cord disease, muscular dystrophy and other neuromuscular disorder, a peripheral nervous system disorder, dermatomyositis and polymyositis, inherited, metabolic, endocrine, and toxic myopathy, myasthenia gravis, periodic paralysis, a mental disorder including mood, anxiety, and schizophrenic disorder, seasonal affective disorder (SAD), akathesia, amnesia, catatonia, diabetic neuropathy, tardive dyskinesia, dystonias, paranoid psychoses, postherpetic neuralgia, and Tourette's disorder; a developmental disorder such as renal tubular acidosis, anemia, Cushing's syndrome, achondroplastic dwarfism, Duchenne and Becker muscular dystrophy, epilepsy, gonadal dysgenesis, WAGR syndrome (Wilms' tumor, aniridia, genitourinary abnormalities, and mental retardation), Smith-Magenis syndrome, myelodysplastic syndrome, hereditary mucoepithelial dysplasia, hereditary keratodermas, hereditary

hydrocephalus, seizure disorders such as Syndenham's chorea and cerebral palsy, spina bifida, anencephaly, craniorachischisis, congenital glaucoma, cataract, and sensorineural hearing loss; an autoimmune/inflammatory disorder such as acquired immunodeficiency syndrome (AIDS), Addison's disease, adult respiratory distress syndrome, allergies, ankylosing spondylitis, amyloidosis, anemia, asthma, atherosclerosis, autoimmune hemolytic anemia, autoimmune thyroiditis, autoimmune polyendocrinopathy-candidiasis-ectodermal dystrophy (APECED), bronchitis, cholecystitis, contact dermatitis, Crohn's disease, atopic dermatitis, dermatomyositis, diabetes mellitus, emphysema, episodic lymphopenia with lymphocytotoxins, erythroblastosis fetalis, erythema nodosum, atrophic gastritis,

neuropathies such as Charcot-Marie-Tooth disease and neurofibromatosis, hypothyroidism,

glomerulonephritis, Goodpasture's syndrome, gout, Graves' disease, Hashimoto's thyroiditis, hypereosinophilia, irritable bowel syndrome, multiple sclerosis, myasthenia gravis, myocardial or pericardial inflammation, osteoarthritis, osteoporosis, pancreatitis, polymyositis, psoriasis, Reiter's syndrome, rheumatoid arthritis, scleroderma, Sjögren's syndrome, systemic anaphylaxis, systemic lupus erythematosus, systemic sclerosis, thrombocytopenic purpura, ulcerative colitis, uveitis, Werner syndrome, complications of cancer, hemodialysis, and extracorporeal circulation, viral, bacterial, fungal, parasitic, protozoal, and helminthic infections, and trauma; and an infection, such as those caused by a viral agent classified as adenovirus, arenavirus, bunyavirus, calicivirus, coronavirus, filovirus, hepadnavirus, herpesvirus, flavivirus, orthomyxovirus, parvovirus, papovavirus, 10 paramyxovirus, picornavirus, poxvirus, reovirus, retrovirus, rhabdovirus, or togavirus; an infection caused by a bacterial agent classified as pneumococcus, staphylococcus, streptococcus, bacillus, corynebacterium, clostridium, meningococcus, gonococcus, listeria, moraxella, kingella, haemophilus, legionella, bordetella, gram-negative enterobacterium including shigella, salmonella, or campylobacter, pseudomonas, vibrio, brucella, francisella, yersinia, bartonella, norcardium, actinomyces, mycobacterium, spirochaetale, rickettsia, chlamydia, or mycoplasma; an infection caused by a fungal agent classified as aspergillus, blastomyces, dermatophytes, cryptococcus, coccidioides, malasezzia, histoplasma, or other mycosis-causing fungal agent; and an infection caused by a parasite classified as plasmodium or malaria-causing, parasitic entamoeba, leishmania, trypanosoma, toxoplasma, pneumocystis carinii, intestinal protozoa such as giardia, trichomonas, tissue nematode such as trichinella, intestinal nematode such as ascaris, lymphatic filarial nematode, trematode such as schistosoma, and cestode such as tapeworm. Polynucleotides encoding NAAP may be used in Southern or northern analysis, dot blot, or other membrane-based technologies; in PCR technologies; in dipstick, pin, and multiformat ELISA-like assays; and in microarrays utilizing fluids or tissues from patients to detect altered NAAP expression. Such qualitative or quantitative methods are well known 25 in the art.

In a particular embodiment, polynucleotides encoding NAAP may be used in assays that detect the presence of associated disorders, particularly those mentioned above. Polynucleotides complementary to sequences encoding NAAP may be labeled by standard methods and added to a fluid or tissue sample from a patient under conditions suitable for the formation of hybridization complexes. After a suitable incubation period, the sample is washed and the signal is quantified and compared with a standard value. If the amount of signal in the patient sample is significantly altered in comparison to a control sample then the presence of altered levels of polynucleotides encoding NAAP

92

in the sample indicates the presence of the associated disorder. Such assays may also be used to evaluate the efficacy of a particular therapeutic treatment regimen in animal studies, in clinical trials, or to monitor the treatment of an individual patient.

In order to provide a basis for the diagnosis of a disorder associated with expression of NAAP, a normal or standard profile for expression is established. This may be accomplished by combining body fluids or cell extracts taken from normal subjects, either animal or human, with a sequence, or a fragment thereof, encoding NAAP, under conditions suitable for hybridization or amplification. Standard hybridization may be quantified by comparing the values obtained from normal subjects with values from an experiment in which a known amount of a substantially purified polynucleotide is used. Standard values obtained in this manner may be compared with values obtained from samples from patients who are symptomatic for a disorder. Deviation from standard values is used to establish the presence of a disorder.

Once the presence of a disorder is established and a treatment protocol is initiated, hybridization assays may be repeated on a regular basis to determine if the level of expression in the patient begins to approximate that which is observed in the normal subject. The results obtained from successive assays may be used to show the efficacy of treatment over a period ranging from several days to months.

With respect to cancer, the presence of an abnormal amount of transcript (either under- or overexpressed) in biopsied tissue from an individual may indicate a predisposition for the development of the disease; or may provide a means for detecting the disease prior to the appearance of actual clinical symptoms. A more definitive diagnosis of this type may allow health professionals to employ preventative measures or aggressive treatment earlier, thereby preventing the development or further progression of the cancer.

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Additional diagnostic uses for oligonucleotides designed from the sequences encoding NAAP may involve the use of PCR. These oligomers may be chemically synthesized, generated enzymatically, or produced *in vitro*. Oligomers will preferably contain a fragment of a polynucleotide encoding NAAP, or a fragment of a polynucleotide complementary to the polynucleotide encoding NAAP, and will be employed under optimized conditions for identification of a specific gene or condition. Oligomers may also be employed under less stringent conditions for detection or quantification of closely related DNA or RNA sequences.

In a particular aspect, oligonucleotide primers derived from polynucleotides encoding NAAP may be used to detect single nucleotide polymorphisms (SNPs). SNPs are substitutions, insertions and

deletions that are a frequent cause of inherited or acquired genetic disease in humans. Methods of SNP detection include, but are not limited to, single-stranded conformation polymorphism (SSCP) and fluorescent SSCP (fSSCP) methods. In SSCP, oligonucleotide primers derived from polynucleotides encoding NAAP are used to amplify DNA using the polymerase chain reaction (PCR). The DNA may be derived, for example, from diseased or normal tissue, biopsy samples, bodily fluids, and the like. SNPs in the DNA cause differences in the secondary and tertiary structures of PCR products in single-stranded form, and these differences are detectable using gel electrophoresis in non-denaturing gels. In fSCCP, the oligonucleotide primers are fluorescently labeled, which allows detection of the amplimers in high-throughput equipment such as DNA sequencing machines. Additionally, sequence database analysis methods, termed in silico SNP (isSNP), are capable of identifying polymorphisms by comparing the sequence of individual overlapping DNA fragments which assemble into a common consensus sequence. These computer-based methods filter out sequence variations due to laboratory preparation of DNA and sequencing errors using statistical models and automated analyses of DNA sequence chromatograms. In the alternative, SNPs may be detected and characterized by mass spectrometry using, for example, the high throughput MASSARRAY system (Sequenom, Inc., San Diego CA).

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SNPs may be used to study the genetic basis of human disease. For example, at least 16 common SNPs have been associated with non-insulin-dependent diabetes mellitus. SNPs are also useful for examining differences in disease outcomes in monogenic disorders, such as cystic fibrosis, sickle cell anemia, or chronic granulomatous disease. For example, variants in the mannose-binding lectin, MBL2, have been shown to be correlated with deleterious pulmonary outcomes in cystic fibrosis. SNPs also have utility in pharmacogenomics, the identification of genetic variants that influence a patient's response to a drug, such as life-threatening toxicity. For example, a variation in N-acetyl transferase is associated with a high incidence of peripheral neuropathy in response to the anti-tuberculosis drug isoniazid, while a variation in the core promoter of the ALOX5 gene results in diminished clinical response to treatment with an anti-asthma drug that targets the 5-lipoxygenase pathway. Analysis of the distribution of SNPs in different populations is useful for investigating genetic drift, mutation, recombination, and selection, as well as for tracing the origins of populations and their migrations (Taylor, J.G. et al. (2001) Trends Mol. Med. 7:507-512; Kwok, P.-Y. and Z. Gu (1999) Mol. Med. Today 5:538-543; Nowotny, P. et al. (2001) Curr. Opin. Neurobiol. 11:637-641).

Methods which may also be used to quantify the expression of NAAP include radiolabeling or biotinylating nucleotides, coamplification of a control nucleic acid, and interpolating results from

standard curves (Melby, P.C. et al. (1993) J. Immunol. Methods 159:235-244; Duplaa, C. et al. (1993) Anal. Biochem. 212:229-236). The speed of quantitation of multiple samples may be accelerated by running the assay in a high-throughput format where the oligomer or polynucleotide of interest is presented in various dilutions and a spectrophotometric or colorimetric response gives rapid quantitation.

In further embodiments, oligonucleotides or longer fragments derived from any of the polynucleotides described herein may be used as elements on a microarray. The microarray can be used in transcript imaging techniques which monitor the relative expression levels of large numbers of genes simultaneously as described below. The microarray may also be used to identify genetic variants, mutations, and polymorphisms. This information may be used to determine gene function, to understand the genetic basis of a disorder, to diagnose a disorder, to monitor progression/regression of disease as a function of gene expression, and to develop and monitor the activities of therapeutic agents in the treatment of disease. In particular, this information may be used to develop a pharmacogenomic profile of a patient in order to select the most appropriate and effective treatment regimen for that patient. For example, therapeutic agents which are highly effective and display the fewest side effects may be selected for a patient based on his/her pharmacogenomic profile.

In another embodiment, NAAP, fragments of NAAP, or antibodies specific for NAAP may be used as elements on a microarray. The microarray may be used to monitor or measure protein-protein interactions, drug-target interactions, and gene expression profiles, as described above.

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A particular embodiment relates to the use of the polynucleotides of the present invention to generate a transcript image of a tissue or cell type. A transcript image represents the global pattern of gene expression by a particular tissue or cell type. Global gene expression patterns are analyzed by quantifying the number of expressed genes and their relative abundance under given conditions and at a given time (Seilhamer et al., "Comparative Gene Transcript Analysis," U.S. Patent No. 5,840,484; hereby expressly incorporated by reference herein). Thus a transcript image may be generated by hybridizing the polynucleotides of the present invention or their complements to the totality of transcripts or reverse transcripts of a particular tissue or cell type. In one embodiment, the hybridization takes place in high-throughput format, wherein the polynucleotides of the present invention or their complements comprise a subset of a plurality of elements on a microarray. The resultant transcript image would provide a profile of gene activity.

Transcript images may be generated using transcripts isolated from tissues, cell lines, biopsies, or other biological samples. The transcript image may thus reflect gene expression in vivo, as in the

case of a tissue or biopsy sample, or in vitro, as in the case of a cell line.

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Transcript images which profile the expression of the polynucleotides of the present invention may also be used in conjunction with in vitro model systems and preclinical evaluation of pharmaceuticals, as well as toxicological testing of industrial and naturally-occurring environmental compounds. All compounds induce characteristic gene expression patterns, frequently termed molecular fingerprints or toxicant signatures, which are indicative of mechanisms of action and toxicity (Nuwaysir, E.F. et al. (1999) Mol. Carcinog. 24:153-159; Steiner, S. and N.L. Anderson (2000) Toxicol. Lett. 112-113:467-471). If a test compound has a signature similar to that of a compound with known toxicity, it is likely to share those toxic properties. These fingerprints or signatures are most useful and refined when they contain expression information from a large number of genes and gene families. Ideally, a genome-wide measurement of expression provides the highest quality signature. Even genes whose expression is not altered by any tested compounds are important as well, as the levels of expression of these genes are used to normalize the rest of the expression data. The normalization procedure is useful for comparison of expression data after treatment with different compounds. While the assignment of gene function to elements of a toxicant signature aids in interpretation of toxicity mechanisms, knowledge of gene function is not necessary for the statistical matching of signatures which leads to prediction of toxicity (see, for example, Press Release 00-02 from the National Institute of Environmental Health Sciences, released February 29, 2000, available at http://www.niehs.nih.gov/oc/news/toxchip.htm). Therefore, it is important and desirable in toxicological screening using toxicant signatures to include all expressed gene sequences.

In an embodiment, the toxicity of a test compound can be assessed by treating a biological sample containing nucleic acids with the test compound. Nucleic acids that are expressed in the treated biological sample are hybridized with one or more probes specific to the polynucleotides of the present invention, so that transcript levels corresponding to the polynucleotides of the present invention may be quantified. The transcript levels in the treated biological sample are compared with levels in an untreated biological sample. Differences in the transcript levels between the two samples are indicative of a toxic response caused by the test compound in the treated sample.

Another embodiment relates to the use of the polypeptides disclosed herein to analyze the proteome of a tissue or cell type. The term proteome refers to the global pattern of protein expression in a particular tissue or cell type. Each protein component of a proteome can be subjected individually to further analysis. Proteome expression patterns, or profiles, are analyzed by quantifying the number of expressed proteins and their relative abundance under given conditions and at a given time. A

profile of a cell's proteome may thus be generated by separating and analyzing the polypeptides of a particular tissue or cell type. In one embodiment, the separation is achieved using two-dimensional gel electrophoresis, in which proteins from a sample are separated by isoelectric focusing in the first dimension, and then according to molecular weight by sodium dodecyl sulfate slab gel electrophoresis in the second dimension (Steiner and Anderson, supra). The proteins are visualized in the gel as discrete and uniquely positioned spots, typically by staining the gel with an agent such as Coomassie Blue or silver or fluorescent stains. The optical density of each protein spot is generally proportional to the level of the protein in the sample. The optical densities of equivalently positioned protein spots from different samples, for example, from biological samples either treated or untreated with a test compound or therapeutic agent, are compared to identify any changes in protein spot density related to the treatment. The proteins in the spots are partially sequenced using, for example, standard methods employing chemical or enzymatic cleavage followed by mass spectrometry. The identity of the protein in a spot may be determined by comparing its partial sequence, preferably of at least 5 contiguous amino acid residues, to the polypeptide sequences of interest. In some cases, further sequence data may be obtained for definitive protein identification.

A proteomic profile may also be generated using antibodies specific for NAAP to quantify the levels of NAAP expression. In one embodiment, the antibodies are used as elements on a microarray, and protein expression levels are quantified by exposing the microarray to the sample and detecting the levels of protein bound to each array element (Lueking, A. et al. (1999) Anal. Biochem. 270:103-111; Mendoze, L.G. et al. (1999) Biotechniques 27:778-788). Detection may be performed by a variety of methods known in the art, for example, by reacting the proteins in the sample with a thiol- or amino-reactive fluorescent compound and detecting the amount of fluorescence bound at each array element.

Toxicant signatures at the proteome level are also useful for toxicological screening, and should be analyzed in parallel with toxicant signatures at the transcript level. There is a poor correlation between transcript and protein abundances for some proteins in some tissues (Anderson, N.L. and J. Seilhamer (1997) Electrophoresis 18:533-537), so proteome toxicant signatures may be useful in the analysis of compounds which do not significantly affect the transcript image, but which alter the proteomic profile. In addition, the analysis of transcripts in body fluids is difficult, due to rapid degradation of mRNA, so proteomic profiling may be more reliable and informative in such cases.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins that are expressed in the treated

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biological sample are separated so that the amount of each protein can be quantified. The amount of each protein is compared to the amount of the corresponding protein in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample. Individual proteins are identified by sequencing the amino acid residues of the individual proteins and comparing these partial sequences to the polypeptides of the present invention.

In another embodiment, the toxicity of a test compound is assessed by treating a biological sample containing proteins with the test compound. Proteins from the biological sample are incubated with antibodies specific to the polypeptides of the present invention. The amount of protein recognized by the antibodies is quantified. The amount of protein in the treated biological sample is compared with the amount in an untreated biological sample. A difference in the amount of protein between the two samples is indicative of a toxic response to the test compound in the treated sample.

Microarrays may be prepared, used, and analyzed using methods known in the art (Brennan, T.M. et al. (1995) U.S. Patent No. 5,474,796; Schena, M. et al. (1996) Proc. Natl. Acad. Sci. USA 93:10614-10619; Baldeschweiler et al. (1995) PCT application WO95/251116; Shalon, D. et al. (1995) PCT application WO95/35505; Heller, R.A. et al. (1997) Proc. Natl. Acad. Sci. USA 94:2150-2155; Heller, M.J. et al. (1997) U.S. Patent No. 5,605,662). Various types of microarrays are well known and thoroughly described in Schena, M., ed. (1999; <u>DNA Microarrays: A Practical Approach</u>, Oxford University Press, London).

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In another embodiment of the invention, nucleic acid sequences encoding NAAP may be used to generate hybridization probes useful in mapping the naturally occurring genomic sequence. Either coding or noncoding sequences may be used, and in some instances, noncoding sequences may be preferable over coding sequences. For example, conservation of a coding sequence among members of a multi-gene family may potentially cause undesired cross hybridization during chromosomal mapping. The sequences may be mapped to a particular chromosome, to a specific region of a chromosome, or to artificial chromosome constructions, e.g., human artificial chromosomes (HACs), yeast artificial chromosomes (YACs), bacterial artificial chromosomes (BACs), bacterial P1 constructions, or single chromosome cDNA libraries (Harrington, J.J. et al. (1997) Nat. Genet. 15:345-355; Price, C.M. (1993) Blood Rev. 7:127-134; Trask, B.J. (1991) Trends Genet. 7:149-154). Once mapped, the nucleic acid sequences may be used to develop genetic linkage maps, for example, which correlate the inheritance of a disease state with the inheritance of a particular chromosome region or restriction fragment length polymorphism (RFLP) (Lander, E.S. and D. Botstein (1986) Proc. Natl.

Acad. Sci. USA 83:7353-7357).

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Fluorescent in situ hybridization (FISH) may be correlated with other physical and genetic map data (Heinz-Ulrich, et al. (1995) in Meyers, supra, pp. 965-968). Examples of genetic map data can be found in various scientific journals or at the Online Mendelian Inheritance in Man (OMIM) World Wide Web site. Correlation between the location of the gene encoding NAAP on a physical map and a specific disorder, or a predisposition to a specific disorder, may help define the region of DNA associated with that disorder and thus may further positional cloning efforts.

In situ hybridization of chromosomal preparations and physical mapping techniques, such as linkage analysis using established chromosomal markers, may be used for extending genetic maps.

Often the placement of a gene on the chromosome of another mammalian species, such as mouse, may reveal associated markers even if the exact chromosomal locus is not known. This information is valuable to investigators searching for disease genes using positional cloning or other gene discovery techniques. Once the gene or genes responsible for a disease or syndrome have been crudely localized by genetic linkage to a particular genomic region, e.g., ataxia-telangiectasia to 11q22-23, any sequences mapping to that area may represent associated or regulatory genes for further investigation (Gatti, R.A. et al. (1988) Nature 336:577-580). The nucleotide sequence of the instant invention may also be used to detect differences in the chromosomal location due to translocation, inversion, etc., among normal, carrier, or affected individuals.

In another embodiment of the invention, NAAP, its catalytic or immunogenic fragments, or oligopeptides thereof can be used for screening libraries of compounds in any of a variety of drug screening techniques. The fragment employed in such screening may be free in solution, affixed to a solid support, borne on a cell surface, or located intracellularly. The formation of binding complexes between NAAP and the agent being tested may be measured.

Another technique for drug screening provides for high throughput screening of compounds having suitable binding affinity to the protein of interest (Geysen, et al. (1984) PCT application WO84/03564). In this method, large numbers of different small test compounds are synthesized on a solid substrate. The test compounds are reacted with NAAP, or fragments thereof, and washed. Bound NAAP is then detected by methods well known in the art. Purified NAAP can also be coated directly onto plates for use in the aforementioned drug screening techniques. Alternatively, non-neutralizing antibodies can be used to capture the peptide and immobilize it on a solid support.

In another embodiment, one may use competitive drug screening assays in which neutralizing antibodies capable of binding NAAP specifically compete with a test compound for binding NAAP.

In this manner, antibodies can be used to detect the presence of any peptide which shares one or more antigenic determinants with NAAP.

In additional embodiments, the nucleotide sequences which encode NAAP may be used in any molecular biology techniques that have yet to be developed, provided the new techniques rely on properties of nucleotide sequences that are currently known, including, but not limited to, such properties as the triplet genetic code and specific base pair interactions.

Without further elaboration, it is believed that one skilled in the art can, using the preceding description, utilize the present invention to its fullest extent. The following embodiments are, therefore, to be construed as merely illustrative, and not limitative of the remainder of the disclosure in any way whatsoever.

The disclosures of all patents, applications, and publications mentioned above and below, including U.S. Ser. No. 60/343,004, U.S. Ser. No. 60/347,633, U.S. Ser. No. 60/359,498, and U.S. Ser. No. 60/351,749, are hereby expressly incorporated by reference.

15 EXAMPLES

I. Construction of cDNA Libraries

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Incyte cDNAs were derived from cDNA libraries described in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA). Some tissues were homogenized and lysed in guanidinium isothiocyanate, while others were homogenized and lysed in phenol or in a suitable mixture of denaturants, such as TRIZOL (Invitrogen), a monophasic solution of phenol and guanidine isothiocyanate. The resulting lysates were centrifuged over CsCl cushions or extracted with chloroform. RNA was precipitated from the lysates with either isopropanol or sodium acetate and ethanol, or by other routine methods.

Phenol extraction and precipitation of RNA were repeated as necessary to increase RNA purity. In some cases, RNA was treated with DNase. For most libraries, poly(A)+ RNA was isolated using oligo d(T)-coupled paramagnetic particles (Promega), OLIGOTEX latex particles (QIAGEN, Chatsworth CA), or an OLIGOTEX mRNA purification kit (QIAGEN). Alternatively, RNA was isolated directly from tissue lysates using other RNA isolation kits, e.g., the POLY(A)PURE mRNA purification kit (Ambion, Austin TX).

In some cases, Stratagene was provided with RNA and constructed the corresponding cDNA libraries. Otherwise, cDNA was synthesized and cDNA libraries were constructed with the UNIZAP vector system (Stratagene) or SUPERSCRIPT plasmid system (Invitrogen), using the

recommended procedures or similar methods known in the art (Ausubel et al., supra, ch. 5). Reverse transcription was initiated using oligo d(T) or random primers. Synthetic oligonucleotide adapters were ligated to double stranded cDNA, and the cDNA was digested with the appropriate restriction enzyme or enzymes. For most libraries, the cDNA was size-selected (300-1000 bp) using SEPHACRYL S1000, SEPHAROSE CL2B, or SEPHAROSE CL4B column chromatography (Amersham Biosciences) or preparative agarose gel electrophoresis. cDNAs were ligated into compatible restriction enzyme sites of the polylinker of a suitable plasmid, e.g., PBLUESCRIPT plasmid (Stratagene), PSPORT1 plasmid (Invitrogen, Carlsbad CA), PCDNA2.1 plasmid (Invitrogen), PBK-CMV plasmid (Stratagene), PCR2-TOPOTA plasmid (Invitrogen), PCMV-ICIS plasmid (Stratagene), pIGEN (Incyte Genomics, Palo Alto CA), pRARE (Incyte Genomics), or pINCY (Incyte Genomics), or derivatives thereof. Recombinant plasmids were transformed into competent E. coli cells including XL1-Blue, XL1-BlueMRF, or SOLR from Stratagene or DH5α, DH10B, or ElectroMAX DH10B from Invitrogen.

II. Isolation of cDNA Clones

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Plasmids obtained as described in Example I were recovered from host cells by in vivo excision using the UNIZAP vector system (Stratagene) or by cell lysis. Plasmids were purified using at least one of the following: a Magic or WIZARD Minipreps DNA purification system (Promega); an AGTC Miniprep purification kit (Edge Biosystems, Gaithersburg MD); and QIAWELL 8 Plasmid, QIAWELL 8 Plus Plasmid, QIAWELL 8 Ultra Plasmid purification systems or the R.E.A.L. PREP 96 plasmid purification kit from QIAGEN. Following precipitation, plasmids were resuspended in 0.1 ml of distilled water and stored, with or without lyophilization, at 4°C.

Alternatively, plasmid DNA was amplified from host cell lysates using direct link PCR in a high-throughput format (Rao, V.B. (1994) Anal. Biochem. 216:1-14). Host cell lysis and thermal cycling steps were carried out in a single reaction mixture. Samples were processed and stored in 384-well plates, and the concentration of amplified plasmid DNA was quantified fluorometrically using PICOGREEN dye (Molecular Probes, Eugene OR) and a FLUOROSKAN II fluorescence scanner (Labsystems Oy, Helsinki, Finland).

III. Sequencing and Analysis

Incyte cDNA recovered in plasmids as described in Example II were sequenced as follows.

Sequencing reactions were processed using standard methods or high-throughput instrumentation such as the ABI CATALYST 800 (Applied Biosystems) thermal cycler or the PTC-200 thermal cycler (MJ Research) in conjunction with the HYDRA microdispenser (Robbins Scientific) or the

MICROLAB 2200 (Hamilton) liquid transfer system. cDNA sequencing reactions were prepared using reagents provided by Amersham Biosciences or supplied in ABI sequencing kits such as the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems). Electrophoretic separation of cDNA sequencing reactions and detection of labeled polynucleotides were carried out using the MEGABACE 1000 DNA sequencing system (Amersham Biosciences); the ABI PRISM 373 or 377 sequencing system (Applied Biosystems) in conjunction with standard ABI protocols and base calling software; or other sequence analysis systems known in the art. Reading frames within the cDNA sequences were identified using standard methods (Ausubel et al., supra, ch. 7). Some of the cDNA sequences were selected for extension using the techniques disclosed in Example VIII.

The polynucleotide sequences derived from Incyte cDNAs were validated by removing vector, linker, and poly(A) sequences and by masking ambiguous bases, using algorithms and programs based on BLAST, dynamic programming, and dinucleotide nearest neighbor analysis. The Incyte cDNA sequences or translations thereof were then queried against a selection of public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases, and BLOCKS, PRINTS, DOMO, PRODOM; PROTEOME databases with sequences from Homo sapiens, Rattus norvegicus, Mus musculus, Caenorhabditis elegans, Saccharomyces cerevisiae, Schizosaccharomyces pombe, and Candida albicans (Incyte Genomics, Palo Alto CA); hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM (Haft, D.H. et al. (2001) Nucleic Acids Res. 29:41-43); and HMM-based protein domain databases such as SMART (Schultz, J. et al. (1998) Proc. Natl. Acad. Sci. USA 95:5857-5864; Letunic, I. et al. (2002) Nucleic Acids Res. 30:242-244). (HMM is a probabilistic approach which analyzes consensus primary structures of gene families; see, for example, Eddy, S.R. (1996) Curr. Opin. Struct. Biol. 6:361-365.) The queries were performed using programs based on BLAST, FASTA, BLIMPS, and HMMER. The Incyte cDNA sequences were assembled to produce full length polynucleotide sequences. Alternatively, GenBank cDNAs, GenBank ESTs, stitched sequences, stretched sequences, or Genscan-predicted coding sequences (see Examples IV and V) were used to extend Incyte cDNA assemblages to full length. Assembly was performed using programs based on Phred, Phrap, and Consed, and cDNA assemblages were screened for open reading frames using programs based on GeneMark, BLAST, and FASTA. The full length polynucleotide sequences were translated to derive the corresponding full length polypeptide sequences. Alternatively, a polypeptide may begin

at any of the methionine residues of the full length translated polypeptide. Full length polypeptide

sequences were subsequently analyzed by querying against databases such as the GenBank protein databases (genpept), SwissProt, the PROTEOME databases, BLOCKS, PRINTS, DOMO, PRODOM, Prosite, hidden Markov model (HMM)-based protein family databases such as PFAM, INCY, and TIGRFAM; and HMM-based protein domain databases such as SMART. Full length polynucleotide sequences are also analyzed using MACDNASIS PRO software (MiraiBio, Alameda CA) and LASERGENE software (DNASTAR). Polynucleotide and polypeptide sequence alignments are generated using default parameters specified by the CLUSTAL algorithm as incorporated into the MEGALIGN multisequence alignment program (DNASTAR), which also calculates the percent identity between aligned sequences.

Table 7 summarizes the tools, programs, and algorithms used for the analysis and assembly of Incyte cDNA and full length sequences and provides applicable descriptions, references, and threshold parameters. The first column of Table 7 shows the tools, programs, and algorithms used, the second column provides brief descriptions thereof, the third column presents appropriate references, all of which are incorporated by reference herein in their entirety, and the fourth column presents, where applicable, the scores, probability values, and other parameters used to evaluate the strength of a match between two sequences (the higher the score or the lower the probability value, the greater the identity between two sequences).

The programs described above for the assembly and analysis of full length polynucleotide and polypeptide sequences were also used to identify polynucleotide sequence fragments from SEQ ID NO:58-114. Fragments from about 20 to about 4000 nucleotides which are useful in hybridization and amplification technologies are described in Table 4, column 2.

IV. Identification and Editing of Coding Sequences from Genomic DNA

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Putative nucleic acid-associated proteins were initially identified by running the Genscan gene identification program against public genomic sequence databases (e.g., gbpri and gbhtg). Genscan is a general-purpose gene identification program which analyzes genomic DNA sequences from a variety of organisms (Burge, C. and S. Karlin (1997) J. Mol. Biol. 268:78-94; Burge, C. and S. Karlin (1998) Curr. Opin. Struct. Biol. 8:346-354). The program concatenates predicted exons to form an assembled cDNA sequence extending from a methionine to a stop codon. The output of Genscan is a FASTA database of polynucleotide and polypeptide sequences. The maximum range of sequence for Genscan to analyze at once was set to 30 kb. To determine which of these Genscan predicted cDNA sequences encode nucleic acid-associated proteins, the encoded polypeptides were analyzed by querying against PFAM models for nucleic acid-associated proteins. Potential nucleic acid-associated

proteins were also identified by homology to Incyte cDNA sequences that had been annotated as nucleic acid-associated proteins. These selected Genscan-predicted sequences were then compared by BLAST analysis to the genpept and gbpri public databases. Where necessary, the Genscan-predicted sequences were then edited by comparison to the top BLAST hit from genpept to correct errors in the sequence predicted by Genscan, such as extra or omitted exons. BLAST analysis was also used to find any Incyte cDNA or public cDNA coverage of the Genscan-predicted sequences, thus providing evidence for transcription. When Incyte cDNA coverage was available, this information was used to correct or confirm the Genscan predicted sequence. Full length polynucleotide sequences were obtained by assembling Genscan-predicted coding sequences with Incyte cDNA sequences and/or public cDNA sequences using the assembly process described in Example III. Alternatively, full length polynucleotide sequences were derived entirely from edited or unedited Genscan-predicted coding sequences.

V. Assembly of Genomic Sequence Data with cDNA Sequence Data "Stitched" Sequences

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Partial cDNA sequences were extended with exons predicted by the Genscan gene identification program described in Example IV. Partial cDNAs assembled as described in Example III were mapped to genomic DNA and parsed into clusters containing related cDNAs and Genscan exon predictions from one or more genomic sequences. Each cluster was analyzed using an algorithm based on graph theory and dynamic programming to integrate cDNA and genomic information, generating possible splice variants that were subsequently confirmed, edited, or extended to create a full length sequence. Sequence intervals in which the entire length of the interval was present on more than one sequence in the cluster were identified, and intervals thus identified were considered to be equivalent by transitivity. For example, if an interval was present on a cDNA and two genomic sequences, then all three intervals were considered to be equivalent. This process allows unrelated but consecutive genomic sequences to be brought together, bridged by cDNA sequence. Intervals thus identified were then "stitched" together by the stitching algorithm in the order that they appear along their parent sequences to generate the longest possible sequence, as well as sequence variants. Linkages between intervals which proceed along one type of parent sequence (cDNA to cDNA or genomic sequence to genomic sequence) were given preference over linkages which change parent type (cDNA to genomic sequence). The resultant stitched sequences were translated and compared by BLAST analysis to the genpept and gbpri public databases. Incorrect exons predicted by Genscan were corrected by comparison to the top BLAST hit from genpept. Sequences were further extended

with additional cDNA sequences, or by inspection of genomic DNA, when necessary.

"Stretched" Sequences

Partial DNA sequences were extended to full length with an algorithm based on BLAST analysis. First, partial cDNAs assembled as described in Example III were queried against public databases such as the GenBank primate, rodent, mammalian, vertebrate, and eukaryote databases using the BLAST program. The nearest GenBank protein homolog was then compared by BLAST analysis to either Incyte cDNA sequences or GenScan exon predicted sequences described in Example IV. A chimeric protein was generated by using the resultant high-scoring segment pairs (HSPs) to map the translated sequences onto the GenBank protein homolog. Insertions or deletions may occur in the chimeric protein with respect to the original GenBank protein homolog. The GenBank protein homolog, the chimeric protein, or both were used as probes to search for homologous genomic sequences from the public human genome databases. Partial DNA sequences were therefore "stretched" or extended by the addition of homologous genomic sequences. The resultant stretched sequences were examined to determine whether it contained a complete gene.

VI. Chromosomal Mapping of NAAP Encoding Polynucleotides

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The sequences which were used to assemble SEQ ID NO:58-114 were compared with sequences from the Incyte LIFESEQ database and public domain databases using BLAST and other implementations of the Smith-Waterman algorithm. Sequences from these databases that matched SEQ ID NO:58-114 were assembled into clusters of contiguous and overlapping sequences using assembly algorithms such as Phrap (Table 7). Radiation hybrid and genetic mapping data available from public resources such as the Stanford Human Genome Center (SHGC), Whitehead Institute for Genome Research (WIGR), and Généthon were used to determine if any of the clustered sequences had been previously mapped. Inclusion of a mapped sequence in a cluster resulted in the assignment of all sequences of that cluster, including its particular SEQ ID NO:, to that map location.

Map locations are represented by ranges, or intervals, of human chromosomes. The map position of an interval, in centiMorgans, is measured relative to the terminus of the chromosome's parm. (The centiMorgan (cM) is a unit of measurement based on recombination frequencies between chromosomal markers. On average, 1 cM is roughly equivalent to 1 megabase (Mb) of DNA in humans, although this can vary widely due to hot and cold spots of recombination.) The cM distances are based on genetic markers mapped by Généthon which provide boundaries for radiation hybrid markers whose sequences were included in each of the clusters. Human genome maps and other resources available to the public, such as the NCBI "GeneMap'99" World Wide Web site

(http://www.ncbi.nlm.nih.gov/genemap/), can be employed to determine if previously identified disease genes map within or in proximity to the intervals indicated above.

VII. Analysis of Polynucleotide Expression

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Northern analysis is a laboratory technique used to detect the presence of a transcript of a gene and involves the hybridization of a labeled nucleotide sequence to a membrane on which RNAs from a particular cell type or tissue have been bound (Sambrook and Russell, *supra*, ch. 7; Ausubel et al., *supra*, ch. 4).

Analogous computer techniques applying BLAST were used to search for identical or related molecules in databases such as GenBank or LIFESEQ (Incyte Genomics). This analysis is much faster than multiple membrane-based hybridizations. In addition, the sensitivity of the computer search can be modified to determine whether any particular match is categorized as exact or similar. The basis of the search is the product score, which is defined as:

BLAST Score x Percent Identity

5 x minimum {length(Seq. 1), length(Seq. 2)}

The product score takes into account both the degree of similarity between two sequences and the length of the sequence match. The product score is a normalized value between 0 and 100, and is calculated as follows: the BLAST score is multiplied by the percent nucleotide identity and the product is divided by (5 times the length of the shorter of the two sequences). The BLAST score is calculated by assigning a score of +5 for every base that matches in a high-scoring segment pair (HSP), and -4 for every mismatch. Two sequences may share more than one HSP (separated by gaps). If there is more than one HSP, then the pair with the highest BLAST score is used to calculate the product score. The product score represents a balance between fractional overlap and quality in a BLAST alignment. For example, a product score of 100 is produced only for 100% identity over the entire length of the shorter of the two sequences being compared. A product score of 70 is produced either by 100% identity and 70% overlap at one end, or by 88% identity and 100% overlap at the other. A product score of 50 is produced either by 100% identity and 50% overlap at one end, or 79% identity and 100% overlap.

Alternatively, polynucleotides encoding NAAP are analyzed with respect to the tissue sources from which they were derived. For example, some full length sequences are assembled, at least in part, with overlapping Incyte cDNA sequences (see Example III). Each cDNA sequence is derived

from a cDNA library constructed from a human tissue. Each human tissue is classified into one of the following organ/tissue categories: cardiovascular system; connective tissue; digestive system; embryonic structures; endocrine system; exocrine glands; genitalia, female; genitalia, male; germ cells; hemic and immune system; liver; musculoskeletal system; nervous system; pancreas; respiratory system; sense organs; skin; stomatognathic system; unclassified/mixed; or urinary tract. The number of libraries in each category is counted and divided by the total number of libraries across all categories. Similarly, each human tissue is classified into one of the following disease/condition categories: cancer, cell line, developmental, inflammation, neurological, trauma, cardiovascular, pooled, and other, and the number of libraries in each category is counted and divided by the total number of libraries across all categories. The resulting percentages reflect the tissue- and disease-specific expression of cDNA encoding NAAP. cDNA sequences and cDNA library/tissue information are found in the LIFESEQ GOLD database (Incyte Genomics, Palo Alto CA).

VIII. Extension of NAAP Encoding Polynucleotides

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Full length polynucleotides are produced by extension of an appropriate fragment of the full length molecule using oligonucleotide primers designed from this fragment. One primer was synthesized to initiate 5' extension of the known fragment, and the other primer was synthesized to initiate 3' extension of the known fragment. The initial primers were designed using OLIGO 4.06 software (National Biosciences), or another appropriate program, to be about 22 to 30 nucleotides in length, to have a GC content of about 50% or more, and to anneal to the target sequence at temperatures of about 68°C to about 72°C. Any stretch of nucleotides which would result in hairpin structures and primer-primer dimerizations was avoided.

Selected human cDNA libraries were used to extend the sequence. If more than one extension was necessary or desired, additional or nested sets of primers were designed.

High fidelity amplification was obtained by PCR using methods well known in the art. PCR was performed in 96-well plates using the PTC-200 thermal cycler (MJ Research, Inc.). The reaction mix contained DNA template, 200 nmol of each primer, reaction buffer containing Mg²⁺, (NH₄)₂SO₄, and 2-mercaptoethanol, Taq DNA polymerase (Amersham Biosciences), ELONGASE enzyme (Invitrogen), and Pfu DNA polymerase (Stratagene), with the following parameters for primer pair PCI A and PCI B: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times; Step 6: 68°C, 5 min; Step 7: storage at 4°C. In the alternative, the parameters for primer pair T7 and SK+ were as follows: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 57°C, 1 min; Step 4: 68°C, 2 min; Step 5: Steps 2, 3, and 4 repeated 20 times;

Step 6: 68°C, 5 min; Step 7: storage at 4°C.

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The concentration of DNA in each well was determined by dispensing 100 \(mu\)l PICOGREEN quantitation reagent (0.25% (v/v) PICOGREEN; Molecular Probes, Eugene OR) dissolved in 1X TE and 0.5 μ l of undiluted PCR product into each well of an opaque fluorimeter plate (Corning Costar, Acton MA), allowing the DNA to bind to the reagent. The plate was scanned in a Fluoroskan II (Labsystems Oy, Helsinki, Finland) to measure the fluorescence of the sample and to quantify the concentration of DNA. A 5 μ l to 10 μ l aliquot of the reaction mixture was analyzed by electrophoresis on a 1 % agarose gel to determine which reactions were successful in extending the sequence.

The extended nucleotides were desalted and concentrated, transferred to 384-well plates, digested with CviII cholera virus endonuclease (Molecular Biology Research, Madison WI), and sonicated or sheared prior to religation into pUC 18 vector (Amersham Biosciences). For shotgun sequencing, the digested nucleotides were separated on low concentration (0.6 to 0.8%) agarose gels, fragments were excised, and agar digested with Agar ACE (Promega). Extended clones were religated using T4 ligase (New England Biolabs, Beverly MA) into pUC 18 vector (Amersham Biosciences), treated with Pfu DNA polymerase (Stratagene) to fill-in restriction site overhangs, and transfected into competent E. coli cells. Transformed cells were selected on antibiotic-containing media, and individual colonies were picked and cultured overnight at 37°C in 384-well plates in LB/2x carb liquid media.

The cells were lysed, and DNA was amplified by PCR using Taq DNA polymerase (Amersham Biosciences) and Pfu DNA polymerase (Stratagene) with the following parameters: Step 1: 94°C, 3 min; Step 2: 94°C, 15 sec; Step 3: 60°C, 1 min; Step 4: 72°C, 2 min; Step 5: steps 2, 3, and 4 repeated 29 times; Step 6: 72°C, 5 min; Step 7: storage at 4°C. DNA was quantified by PICOGREEN reagent (Molecular Probes) as described above. Samples with low DNA recoveries 25 were reamplified using the same conditions as described above. Samples were diluted with 20% dimethysulfoxide (1:2, v/v), and sequenced using DYENAMIC energy transfer sequencing primers and the DYENAMIC DIRECT kit (Amersham Biosciences) or the ABI PRISM BIGDYE Terminator cycle sequencing ready reaction kit (Applied Biosystems).

In like manner, full length polynucleotides are verified using the above procedure or are used to obtain 5' regulatory sequences using the above procedure along with oligonucleotides designed for such extension, and an appropriate genomic library.

IX. Identification of Single Nucleotide Polymorphisms in NAAP Encoding

Polynucleotides

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Common DNA sequence variants known as single nucleotide polymorphisms (SNPs) were identified in SEQ ID NO:58-114 using the LIFESEQ database (Incyte Genomics). Sequences from the same gene were clustered together and assembled as described in Example III, allowing the identification of all sequence variants in the gene. An algorithm consisting of a series of filters was used to distinguish SNPs from other sequence variants. Preliminary filters removed the majority of basecall errors by requiring a minimum Phred quality score of 15, and removed sequence alignment errors and errors resulting from improper trimming of vector sequences, chimeras, and splice variants. An automated procedure of advanced chromosome analysis analysed the original chromatogram files in the vicinity of the putative SNP. Clone error filters used statistically generated algorithms to identify errors introduced during laboratory processing, such as those caused by reverse transcriptase, polymerase, or somatic mutation. Clustering error filters used statistically generated algorithms to identify errors resulting from clustering of close homologs or pseudogenes, or due to contamination by non-human sequences. A final set of filters removed duplicates and SNPs found in immunoglobulins or T-cell receptors.

Certain SNPs were selected for further characterization by mass spectrometry using the high throughput MASSARRAY system (Sequenom, Inc.) to analyze allele frequencies at the SNP sites in four different human populations. The Caucasian population comprised 92 individuals (46 male, 46 female), including 83 from Utah, four French, three Venezualan, and two Amish individuals. The African population comprised 194 individuals (97 male, 97 female), all African Americans. The Hispanic population comprised 324 individuals (162 male, 162 female), all Mexican Hispanic. The Asian population comprised 126 individuals (64 male, 62 female) with a reported parental breakdown of 43% Chinese, 31% Japanese, 13% Korean, 5% Vietnamese, and 8% other Asian. Allele frequencies were first analyzed in the Caucasian population; in some cases those SNPs which showed no allelic variance in this population were not further tested in the other three populations.

X. Labeling and Use of Individual Hybridization Probes

Hybridization probes derived from SEQ ID NO:58-114 are employed to screen cDNAs, genomic DNAs, or mRNAs. Although the labeling of oligonucleotides, consisting of about 20 base pairs, is specifically described, essentially the same procedure is used with larger nucleotide fragments. Oligonucleotides are designed using state-of-the-art software such as OLIGO 4.06 software (National Biosciences) and labeled by combining 50 pmol of each oligomer, 250 μ Ci of $[\gamma^{-32}P]$ adenosine triphosphate (Amersham Biosciences), and T4 polynucleotide kinase (DuPont NEN,

Boston MA). The labeled oligonucleotides are substantially purified using a SEPHADEX G-25 superfine size exclusion dextran bead column (Amersham Biosciences). An aliquot containing 10⁷ counts per minute of the labeled probe is used in a typical membrane-based hybridization analysis of human genomic DNA digested with one of the following endonucleases: Ase I, Bgl II, Eco RI, Pst I, Xba I, or Pvu II (DuPont NEN).

The DNA from each digest is fractionated on a 0.7% agarose gel and transferred to nylon membranes (Nytran Plus, Schleicher & Schuell, Durham NH). Hybridization is carried out for 16 hours at 40°C. To remove nonspecific signals, blots are sequentially washed at room temperature under conditions of up to, for example, 0.1 x saline sodium citrate and 0.5% sodium dodecyl sulfate. Hybridization patterns are visualized using autoradiography or an alternative imaging means and compared.

XI. Microarrays

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The linkage or synthesis of array elements upon a microarray can be achieved utilizing photolithography, piezoelectric printing (ink-jet printing; see, e.g., Baldeschweiler et al., supra), mechanical microspotting technologies, and derivatives thereof. The substrate in each of the aforementioned technologies should be uniform and solid with a non-porous surface (Schena, M., ed. (1999) DNA Microarrays: A Practical Approach, Oxford University Press, London). Suggested substrates include silicon, silica, glass slides, glass chips, and silicon wafers. Alternatively, a procedure analogous to a dot or slot blot may also be used to arrange and link elements to the surface of a substrate using thermal, UV, chemical, or mechanical bonding procedures. A typical array may be produced using available methods and machines well known to those of ordinary skill in the art and may contain any appropriate number of elements (Schena, M. et al. (1995) Science 270:467-470; Shalon, D. et al. (1996) Genome Res. 6:639-645; Marshall, A. and J. Hodgson (1998) Nat. Biotechnol. 16:27-31).

Full length cDNAs, Expressed Sequence Tags (ESTs), or fragments or oligomers thereof may comprise the elements of the microarray. Fragments or oligomers suitable for hybridization can be selected using software well known in the art such as LASERGENE software (DNASTAR). The array elements are hybridized with polynucleotides in a biological sample. The polynucleotides in the biological sample are conjugated to a fluorescent label or other molecular tag for ease of detection. After hybridization, nonhybridized nucleotides from the biological sample are removed, and a fluorescence scanner is used to detect hybridization at each array element. Alternatively, laser desorbtion and mass spectrometry may be used for detection of hybridization. The degree of

complementarity and the relative abundance of each polynucleotide which hybridizes to an element on the microarray may be assessed. In one embodiment, microarray preparation and usage is described in detail below.

Tissue or Cell Sample Preparation

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Total RNA is isolated from tissue samples using the guanidinium thiocyanate method and poly(A)* RNA is purified using the oligo-(dT) cellulose method. Each poly(A)* RNA sample is reverse transcribed using MMLV reverse-transcriptase, 0.05 pg/μl oligo-(dT) primer (21mer), 1X first strand buffer, 0.03 units/μl RNase inhibitor, 500 μM dATP, 500 μM dGTP, 500 μM dTTP, 40 μM dCTP, 40 μM dCTP-Cy3 (BDS) or dCTP-Cy5 (Amersham Biosciences). The reverse transcription reaction is performed in a 25 ml volume containing 200 ng poly(A)* RNA with GEMBRIGHT kits (Incyte Genomics). Specific control poly(A)* RNAs are synthesized by *in vitro* transcription from non-coding yeast genomic DNA. After incubation at 37°C for 2 hr, each reaction sample (one with Cy3 and another with Cy5 labeling) is treated with 2.5 ml of 0.5M sodium hydroxide and incubated for 20 minutes at 85°C to the stop the reaction and degrade the RNA. Samples are purified using two successive CHROMA SPIN 30 gel filtration spin columns (Clontech, Palo Alto CA) and after combining, both reaction samples are ethanol precipitated using 1 ml of glycogen (1 mg/ml), 60 ml sodium acetate, and 300 ml of 100% ethanol. The sample is then dried to completion using a SpeedVAC (Savant Instruments Inc., Holbrook NY) and resuspended in 14 μl 5X SSC/0.2% SDS.

Microarray Preparation

Sequences of the present invention are used to generate array elements. Each array element is amplified from bacterial cells containing vectors with cloned cDNA inserts. PCR amplification uses primers complementary to the vector sequences flanking the cDNA insert. Array elements are amplified in thirty cycles of PCR from an initial quantity of 1-2 ng to a final quantity greater than 5 μ g. Amplified array elements are then purified using SEPHACRYL-400 (Amersham Biosciences).

Purified array elements are immobilized on polymer-coated glass slides. Glass microscope slides (Corning) are cleaned by ultrasound in 0.1% SDS and acetone, with extensive distilled water washes between and after treatments. Glass slides are etched in 4% hydrofluoric acid (VWR Scientific Products Corporation (VWR), West Chester PA), washed extensively in distilled water, and coated with 0.05% aminopropyl silane (Sigma) in 95% ethanol. Coated slides are cured in a 110°C oven.

Array elements are applied to the coated glass substrate using a procedure described in U.S. Patent No. 5,807,522, incorporated herein by reference. 1 μ l of the array element DNA, at an average

concentration of 100 ng/µl, is loaded into the open capillary printing element by a high-speed robotic apparatus. The apparatus then deposits about 5 nl of array element sample per slide.

Microarrays are UV-crosslinked using a STRATALINKER UV-crosslinker (Stratagene). Microarrays are washed at room temperature once in 0.2% SDS and three times in distilled water. Non-specific binding sites are blocked by incubation of microarrays in 0.2% casein in phosphate buffered saline (PBS) (Tropix, Inc., Bedford MA) for 30 minutes at 60°C followed by washes in 0.2% SDS and distilled water as before.

Hybridization

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Hybridization reactions contain 9 μ l of sample mixture consisting of 0.2 μ g each of Cy3 and Cy5 labeled cDNA synthesis products in 5X SSC, 0.2% SDS hybridization buffer. The sample mixture is heated to 65°C for 5 minutes and is aliquoted onto the microarray surface and covered with an 1.8 cm² coverslip. The arrays are transferred to a waterproof chamber having a cavity just slightly larger than a microscope slide. The chamber is kept at 100% humidity internally by the addition of 140 μl of 5X SSC in a corner of the chamber. The chamber containing the arrays is incubated for about 15 6.5 hours at 60°C. The arrays are washed for 10 min at 45°C in a first wash buffer (1X SSC, 0.1% SDS), three times for 10 minutes each at 45°C in a second wash buffer (0.1X SSC), and dried.

Detection

Reporter-labeled hybridization complexes are detected with a microscope equipped with an Innova 70 mixed gas 10 W laser (Coherent, Inc., Santa Clara CA) capable of generating spectral lines at 488 nm for excitation of Cy3 and at 632 nm for excitation of Cy5. The excitation laser light is focused on the array using a 20X microscope objective (Nikon, Inc., Melville NY). The slide containing the array is placed on a computer-controlled X-Y stage on the microscope and rasterscanned past the objective. The 1.8 cm x 1.8 cm array used in the present example is scanned with a resolution of 20 micrometers.

In two separate scans, a mixed gas multiline laser excites the two fluorophores sequentially. Emitted light is split, based on wavelength, into two photomultiplier tube detectors (PMT R1477, Harnamatsu Photonics Systems, Bridgewater NJ) corresponding to the two fluorophores. Appropriate filters positioned between the array and the photomultiplier tubes are used to filter the signals. The emission maxima of the fluorophores used are 565 nm for Cy3 and 650 nm for Cy5. Each array is typically scanned twice, one scan per fluorophore using the appropriate filters at the laser source, although the apparatus is capable of recording the spectra from both fluorophores simultaneously.

The sensitivity of the scans is typically calibrated using the signal intensity generated by a

cDNA control species added to the sample mixture at a known concentration. A specific location on the array contains a complementary DNA sequence, allowing the intensity of the signal at that location to be correlated with a weight ratio of hybridizing species of 1:100,000. When two samples from different sources (e.g., representing test and control cells), each labeled with a different fluorophore, are hybridized to a single array for the purpose of identifying genes that are differentially expressed, the calibration is done by labeling samples of the calibrating cDNA with the two fluorophores and adding identical amounts of each to the hybridization mixture.

The output of the photomultiplier tube is digitized using a 12-bit RTI-835H analog-to-digital (A/D) conversion board (Analog Devices, Inc., Norwood MA) installed in an IBM-compatible PC computer. The digitized data are displayed as an image where the signal intensity is mapped using a linear 20-color transformation to a pseudocolor scale ranging from blue (low signal) to red (high signal). The data is also analyzed quantitatively. Where two different fluorophores are excited and measured simultaneously, the data are first corrected for optical crosstalk (due to overlapping emission spectra) between the fluorophores using each fluorophore's emission spectrum.

A grid is superimposed over the fluorescence signal image such that the signal from each spot is centered in each element of the grid. The fluorescence signal within each element is then integrated to obtain a numerical value corresponding to the average intensity of the signal. The software used for signal analysis is the GEMTOOLS gene expression analysis program (Incyte Genomics). Array elements that exhibit at least about a two-fold change in expression, a signal-to-background ratio of at least about 2.5, and an element spot size of at least about 40%, are considered to be differentially expressed.

Expression

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For example, both SEQ ID NO:62 and SEQ ID NO:71 showed differential expression in cancer cell lines or tumorous tissue versus non-cancerous cell lines or tissues, as determined by microarray analysis. The expression of SEQ ID NO:62 was decreased by at least two-fold in ovarian tumor tissue from a 79 year-old female donor as compared to matched normal ovarian tissue from the same donor. Matched normal and tumorigenic ovarian tissue samples were provided by the Huntsman Cancer Institute, (Salt Lake City, UT). Therefore, SEQ ID NO:62 is useful in diagnostic assays for ovarian cancer. The expression of SEQ ID NO:71 was increased by at least two-fold in lung squamous cell carcinoma tissue as compared to matched normal lung tissue from the same donor. Matched normal and tumorigenic lung tissue samples were provided by the Roy Castle International Centre for Lung Cancer Research (Liverpool, UK). Therefore, SEQ ID NO:71 is useful in diagnostic

assays for lung cancer.

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In addition, the expression of SEQ ID NO:71 was increased at least two-fold in Tangier disease-derived fibroblasts compared to normal fibroblasts. Both types of cells were also cultured in the presence of cholesterol and compared with the same cell type cultured in the absence of cholesterol. Human fibroblasts were obtained from skin explants from both normal subjects and two patients with homozygous Tangier disease. Cell lines were immortalized by transfection with human papillomavirus 16 genes E6 and E7 and a neomycin resistance selectable marker. TD derived cells are deficient in an assay of apoA-I mediated tritiated cholesterol efflux. Therefore, SEQ ID NO:71 is also useful in diagnostic assays for Tangier disease.

In an alternative example, SEQ ID NO:79 and SEQ ID NO:81 were differentially expressed in human breast tumor cells. Histological and molecular evaluation of breast tumors reveals that the development of breast cancer evolves through a multi-step process whereby pre-malignant mammary epithelial cells undergo a relatively defined sequence of events leading to tumor formation. An early event in tumor development is ductal hyperplasia. Cells undergoing rapid neoplastic growth gradually progress to invasive carcinoma and become metastatic to the lung, bone, and potentially other organs. Several variables that may influence the process of tumor progression and malignant transformation include genetic factors, environmental factors, growth factors, and hormones. Based on the complexity of this process, it is critical to study a population of human mammary epithelial cells undergoing the process of malignant transformation, and to associate specific stages of progression with phenotypic and molecular characteristics. Primary breast epithelial cells (HMECs) were compared to various breast carcinoma lines at various stages of tumor progression.

Gene expression profiles of nonmalignant mammary epithelial cells were compared to gene expression profiles of various breast carcinoma lines at different stages of tumor progression. The cells were grown in defined serum-free H14 medium to 70-80% confluence prior to RNA harvest. Cell lines compared included: a) HMEC, a primary breast epithelial cell line isolated from a normal donor, b)MCF-10A, a breast mammary gland cell line isolated from a 36-year-old woman with fibrocystic breast disease, c)MCF7, a nonmalignant breast adenocarcinoma cell line isolated from the pleural effusion of a 69- year-old female, d)T-47D, a breast carcinoma cell line isolated from a pleural effusion obtained from a 54-year-old female with an infiltrating ductal carcinoma of the breast, e)Sk-BR-3, a breast adenocarcinoma cell line isolated from a malignant pleural effusion of a 43-year-old female, f)BT-20, a breast carcinoma cell line derived *in vitro* from cells emigrating out of thin slices of the tumor mass isolated from a 74-year-old female, g)MDA-mb-231, a breast tumor cell line isolated

from the pleural effusion of a 51-year-old female, and h)MDA-mb-435S, a spindle-shaped strain that evolved from the parent line (435) isolated by R. Cailleau from pleural effusion of a 31-year-old female with metastatic, ductal adenocarcinoma of the breast.

In various microarray experiments, both SEQ ID NO:79 and SEQ ID NO:81 were underexpressed by at least two-fold in the following tumor cell lines: MCF7 (nonmalignant breast adenocarcinoma), T47D (breast carcinoma), BT-20 (breast carcinoma), MDA-mb-231 (breast tumor cell line which also expresses the Wnt3 oncogene, EGF, and TGF-alpha), and MDA-mb-435S (spindle-shaped strain of breast tumor cell evolved from a metastatic, ductal adenocarcinoma cell of the breast).

These experiments indicate that SEQ ID NO:79 and SEQ ID NO:81 exhibited significant differential expression patterns using microarray techniques, and further establish their utility as diagnostic markers, disease staging or therapeutic agents which may be useful in a variety of conditions and diseases involving nucleic acid-associated proteins, including breast cancer.

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In an alternative example, SEQ ID NO:90, SEQ ID NO:91, and SEQ ID NO:93 showed

differential expression in tumorous or diseased tissue versus non-tumorous or healthy tissues, as
determined by microarray analysis. Array elements that exhibited about at least a two-fold change in
expression and a signal intensity over 250 units, a signal-to-background ratio of a least 2.5, and an
element spot size of at least 40% were identified as differentially expressed using the GEMTOOLS
program (Incyte Genomics).

For example, SEQ ID NO:90 showed increased expression in lung carcinoma tissue versus normal lung tissue as determined by microarray analysis. In one experiment, grossly uninvolved lung tissue from a 66 year-old male was compared to lung squamous cell carcinoma tissue from the same donor (Roy Castle International Centre for Lung Cancer Research, Liverpool, UK). In a second experiment, grossly uninvolved lung tissue from a 66 year-old female was compared to lung adenocarcinoma tissue from the same donor (Roy Castle International Centre for Lung Cancer Research, Liverpool, UK). Therefore, SEQ ID NO:90 is useful in monitoring treatment of, and diagnostic assays for, lung cancer and other cell proliferative disorders.

In another example, SEQ ID NO:90 showed differential expression in amygdala enterorhinal cortex tissue versus pooled brain tissue as determined by microarray analysis. Specific brain regions from 4 individual male donors (47, 48, 59, and 60 years old) were compared to a pooled brain control. The pooled brain control was reconstituted from the purified mRNA isolated from the major regions of the brain from two male brains (the 47-year-old male and the 48-year-old male). Therefore, SEQ ID

PCT/US02/41115 WO 03/054219

NO:90 is useful in monitoring treatment of, and diagnostic assays for, Alzheimer's disease and other neurological disorders.

In another example, SEQ ID NO:91 showed decreased expression in breast carcinoma cell lines versus a primary breast epithelial cell line isolated from a normal donor, as determined by 5 microarray analysis. The gene expression profile of a nonmalignant mammary epithelial cell line was compared to the gene expression profiles of breast carcinoma lines at different stages of tumor progression. Cell lines compared included: a) BT-20, a breast carcinoma cell line derived in vitro from the cells emigrating out of thin slices of tumor mass isolated from a 74-year-old female, b) BT-474, a breast ductal carcinoma cell line that was isolated from a solid, invasive ductal carcinoma of the breast obtained from a 60-year-old woman, c) BT-483, a breast ductal carcinoma cell line that was isolated from a papillary invasive ductal tumor obtained from a 23-year-old normal, menstruating, parous female with a family history of breast cancer, d) Hs 578T, a breast ductal carcinoma cell line isolated from a 74-year-old female with breast carcinoma, e) MCF7, a nonmalignant breast adenocarcinoma cell line isolated from the pleural effusion of a 69-year-old female, f) MCF-10A, a 15 breast mammary gland (luminal ductal characteristics) cell line isolated. from a 36-year-old woman with fibrocystic breast disease, g) MDA-MB-468, a breast adenocarcinoma cell line isolated from the pleural effusion of a 51-year-old female with metastatic adenocarcinoma of the breast, and h) HMEC, a primary breast epithelial cell line isolated from a normal donor. SEQ ID NO:91 showed decreased expression in lines a) through e) above. Therefore, SEQ ID NO:91 is useful in monitoring treatment of, and diagnostic assays for, breast cancer and other cell proliferative disorders.

In another example, SEQ ID NO:93 showed decreased expression in breast carcinoma cells treated with estradiol versus untreated breast carcinoma cells as determined by microarray analysis. BT-20 is a breast carcinoma cell line derived in vitro from the cells emigrating out thin slices of the tumor mass isolated from a 74-year-old female. BT-20 cells were treated with α-estradiol for 4, 8, 14, 24, 36, and 48 hours. These treated cells were compared to untreated BT-20 cells kept in culture for the same amount of time. Therefore, SEQ ID NO:93 is useful in monitoring treatment of, and diagnostic assays for, breast cancer and other cell proliferative disorders.

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In another example, SEQ ID NO:93 showed increased expression in coronary artery epithelial cells versus pooled vascular endothelial cells as determined by microarray analysis. Human vascular and microvascular endothelial cells were isolated from coronary artery, umbilical artery and vein, uterine microvasculature, iliac artery, dermal microvasculature, pulmonary artery, aorta. All cell types in this experiment are used as experimental models for investigating the role of endothelium in human

vascular biology. Therefore, SEQ ID NO:93 is useful in monitoring treatment of, and diagnostic assays for, cardiovascular and other autoimmune/inflammation disorders.

In an alternative example, SEQ ID NO:101, SEQ ID NO:103, SEQ ID NO:106, SEQ ID NO:112, and SEQ ID NO:113 showed differential expression in tumorous tissues versus non-tumorous tissues or in treated versus untreated cell lines, as determined by microarray analysis. Array elements that exhibited about at least a two-fold change in expression and a signal intensity over 250 units, a signal-to-background ratio of a least 2.0, and an element spot size of at least 40% were identified as differentially expressed using the GEMTOOLS program (Incyte Genomics).

For example, the expression of SEQ ID NO:101 was decreased at least two-fold in an ovarian adenocarcinoma when matched with normal tissue from the same donor. The tumorous ovary tissue was obtained from ovarian adenocarcinoma from a 79-year-old female. Normal ovary tissue was obtained from ovary from the same donor. Therefore, SEQ ID NO:101 is useful in diagnostic assays for ovarian adenocarcinoma. Matched normal and tumorigenic ovary tissue samples were provided by the Huntsman Cancer Institute, (Salt Lake City, UT).

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The gene expression profile of a nonmalignant mammary epithelial cell line (MCF-10A) or a nonmalignant mammary epithelial cell line (HMEC) was compared to the gene expression profiles of breast carcinoma lines at different stages of tumor progression. Cell lines compared included: a) HMEC, a primary breast epithelial cell line isolated from a normal donor, b) MCF-10A, a breast mammary gland cell line isolated from a 36-year-old woman with fibrocystic breast disease; c)MCF7, a nonmalignant breast adenocarcinoma cell line isolated from the pleural effusion of a 69- year-old female; d)T-47D, a breast carcinoma cell line isolated from a pleural effusion obtained from a 54year-old female with an infiltrating ductal carcinoma of the breast; e)Sk-BR-3, a breast adenocarcinoma cell line isolated from a malignant pleural effusion of a 43-year-old female; f)BT-20, a breast carcinoma cell line derived in vitro from tumor mass isolated from a 74-year-old female; g)MDA-mb-231, a breast tumor cell line isolated from the pleural effusion of a 51-year old female; and h) MDA-mb-435S, a spindle shaped strain that evolved from the parent line (435) isolated from the pleural effusion of a 31-year-old female with metastatic, ductal adenocarcinoma of the breast. All cell lines were grown in the supplier's recommended medium to 70-80% confluence prior to comparison. The expression of SEQ ID NO:101 was decreased at least two-fold in three (MCF7, T-47D, and MDA-mb-231) of the seven cell lines when compared with HMEC or MCF-10A cell lines. These experiments indicate that SEQ ID NO:101 was significantly under-expressed in the breast tumor cell lines tested, establishing the utility of SEQ ID NO:101 as a diagnostic marker for disease

staging or as a potential therapeutic target for breast cancer.

In another example, SEQ ID NO:103, exhibited differential expression by microarray analysis. Early confluent C3A cells were treated with mifepristone, progesterone, beclomethasone, medroxyprogesterone, budesonide, prednisone, dexamethasone, betamethasone, or danazol at concentrations of 1 μM, 10 μM, and 100 μM for 1, 3, and 6 hours. In all cases mRNA from untreated early confluent C3A cells were prepared in parallel as described below. The expression of SEQ ID NO:103 was decreased at least two-fold in HCA3 cells when treated with Progesterone, Beclomethasone, Medroxyprogesterone, Bedusonide, Prednisone, Dexamethasone, and Betamethasone at doses between 1μM and 100 μM for between 1 and 6 hours, and with Catechol plus water at 100μM for 1, 3, and 6 hours. Therefore, SEQ ID NO:103 is useful as a diagnostic marker for disease staging or as a potential therapeutic target for liver disorders such as adenomatosis, cholestasis, cirrhosis, hemangioma, Henoch-Schonlein purpura, hepatitis, hepatocellular and metastatic carcinomas, idiopathic thrombocytopenic purpura, porphyria, sarcoidosis, and Wilson disease.

In another example, the expression of SEQ ID NO:106 was increased at least four-fold in

lung squamous cell carcinoma when matched with normal tissue from the same donor. The tumorous
tissue was obtained from lung squamous cell carcinoma from a 68-year-old female. Normal lung
tissue was obtained from the same donor. Therefore, SEQ ID NO:106 is useful in diagnostic assays
for lung squamous cell carcinoma, in disease staging and as a potential therapeutic target. Matched
normal and tumorigenic lung tissue samples are provided by the Roy Castle International Centre for

Lung Cancer Research (Liverpool UK). Further, the expression of SEQ ID NO:106 was decreased
at least three-fold in colon tumor when matched with normal tissue from the same donor. Tumorous
colon tissue was obtained from a 67-year-old donor with moderately differentiated colon
adenocarcinoma. Normal tissue was obtained from the colon of the same donor. Therefore, SEQ ID
NO:106 is useful as a diagnostic marker, for disease staging, or as a potential therapeutic target for
colon adenocarcinoma. Matched normal and tumorigenic colon tissue samples were provided by the
Huntsman Cancer Institute (Salt Lake City, UT).

In another example, the expression of SEQ ID NO:112 was decreased at least two-fold in two colon adenocarcinomas when matched with normal tissue from the same donor. The tumorous tissue was obtained from a 64-year-old female with moderately differentiated colon adenocarcinoma and from an 83-year old female with colon adenocarcinoma. Normal tissues were obtained from the colons of the same donors. Therefore, SEQ ID NO:112 is useful as a diagnostic marker, for disease staging, or as a potential therapeutic target for colon adenocarcinoma. Matched normal and

tumorigenic colon tissue samples are provided by the Huntsman Cancer Institute (Salt Lake City, UT).

In yet another example, SEQ ID NO:113 showed differential expression in inflammatory responses as determined by microarray analysis. The expression of SEQ ID NO:113 was increased by at least two-fold in three cell lines, Daudi (a B lymphoblast cell line [Burkitt's lymphoma]), Jurkat (an acute T cell leukemia cell line that grows in the absence of external stimuli), and THP-1 (a promonocyte cell line isolated from the peripheral blood of a 1-year-old male with acute monocytic leukemia). These three cell lines are used extensively for the study of signaling in human T cells and B cells. These cell lines were treated with various concentrations of PMA (a broad activator of protein kinase C-dependent (PKC) pathways), with or without Ionomycin (a calcium ionophore that permits the entry of calcium in the cell), with or without Brefeldin A (an antiviral antibiotic factor produced by Penicillium brefeldianum), with or without antibodies to CD3 or CD8, or with or without LPS (lipopolysaccharide). The expression of SEQ ID NO:113 was increased by at least two-fold in three tumorous cell lines from breast (MDA-mb-231, T-47D; and MCF7) when treated with 1 or 10 ng/ml of TNFa. See descriptions supra. The expression of SEQ ID NO:113 was also increased by at least 2.9-fold in DU 145, a prostate carcinoma cell line isolated from a metastatic site in the brain of a 69-year-old male with widespread metastatic prostate carcinoma, when compared to Prec, a primary prostate epithelial cell line isolated from a normal donor; and increased by at least two-fold in the following endothelial cell lines: ECV304 (a cell line derived from the endothelium of the human umbilical vein) treated with PMA and Ionomycin or TNFα; HIAEC (primary cells derived from the endothelium of an iliac artery); HMVEC (primary cells derived from microvascular of human skin endothelial cells); HUAEC (primary cells derived from the endothelium of an umbilical artery); and UtMVECMyo (primary cells derived from the uterine myometrium microvasculature) all treated with 10 ng/ml TNF-a when compared to untreated cells of the same cell line; and HUVEC (primary cells derived from the endothelium of the human umbilical vein) treated with TNFa and cycloheximide when compared to untreated HUVEC cells treated with cycloheximide.

Further, the expression of SEQ ID NO:113 was decreased by at least two-fold in HepG2 (a human hepatoma cell line isolated from a 15-year-old male with a liver tumor) when treated with TNFα. SEQ ID NO:113 was decreased at least two-fold in five out of six breast tumor cell lines (BT20, HMEC, MDA-mb-231, MDA-mb-435S, T-47D; see descriptions supra) when compared with MCF10A cell line. The expression of SEQ ID NO:113 was decreased at least 2.5-fold in a colon cancer when matched with normal tissue from the same donor. The tumouous colon tissue was obtained from a 73-year-old female with colon cancer. Normal colon tissue was obtained from the

colon of the same donor. Also, the expression of SEQ ID NO:113 was decreased at least two-fold in a donor with lung adenocarcinoma and decreased at least 2.5-fold in four donors with lung squamous cell carcinoma. The adenocarcinoma tissue was obtained from a 66-year-old female with lung adenocarcinoma. The squamous cell carcinoma tissue was obtained from a 73-year-old male, a 68-year-old female, a 66-year-old male, and a 73-year-old male. Normal lung tissues were obtained from the lungs of the same donors. All matched normal and tumorous lung tissue samples were provided by the Roy Castle International Centre for Lung Cancer Research (Liverpool UK). Therefore, SEQ ID NO:113 is useful in monitoring treatment of, and diagnostic assays for, autoimmune/inflammation disorders, breast cancer, colon cancer, and lung cancer.

0 XII. Complementary Polynucleotides

Sequences complementary to the NAAP-encoding sequences, or any parts thereof, are used to detect, decrease, or inhibit expression of naturally occurring NAAP. Although use of oligonucleotides comprising from about 15 to 30 base pairs is described, essentially the same procedure is used with smaller or with larger sequence fragments. Appropriate oligonucleotides are designed using OLIGO 4.06 software (National Biosciences) and the coding sequence of NAAP. To inhibit transcription, a complementary oligonucleotide is designed from the most unique 5' sequence and used to prevent promoter binding to the coding sequence. To inhibit translation, a complementary oligonucleotide is designed to prevent ribosomal binding to the NAAP-encoding transcript.

XIII. Expression of NAAP

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Expression and purification of NAAP is achieved using bacterial or virus-based expression systems. For expression of NAAP in bacteria, cDNA is subcloned into an appropriate vector containing an antibiotic resistance gene and an inducible promoter that directs high levels of cDNA transcription. Examples of such promoters include, but are not limited to, the *trp-lac* (*tac*) hybrid promoter and the T5 or T7 bacteriophage promoter in conjunction with the *lac* operator regulatory element. Recombinant vectors are transformed into suitable bacterial hosts, e.g., BL21(DE3). Antibiotic resistant bacteria express NAAP upon induction with isopropyl beta-D-thiogalactopyranoside (IPTG). Expression of NAAP in eukaryotic cells is achieved by infecting insect or mammalian cell lines with recombinant *Autographica californica* nuclear polyhedrosis virus (AcMNPV), commonly known as baculovirus. The nonessential polyhedrin gene of baculovirus is replaced with cDNA encoding NAAP by either homologous recombination or bacterial-mediated transposition involving transfer plasmid intermediates. Viral infectivity is maintained and the strong polyhedrin promoter drives high levels of cDNA transcription. Recombinant baculovirus is used to

PCT/US02/41115 WO 03/054219

infect Spodoptera frugiperda (Sf9) insect cells in most cases, or human hepatocytes, in some cases. Infection of the latter requires additional genetic modifications to baculovirus (Engelhard, E.K. et al. (1994) Proc. Natl. Acad. Sci. USA 91:3224-3227; Sandig, V. et al. (1996) Hum. Gene Ther. 7:1937-1945).

In most expression systems, NAAP is synthesized as a fusion protein with, e.g., glutathione Stransferase (GST) or a peptide epitope tag, such as FLAG or 6-His, permitting rapid, single-step, affinity-based purification of recombinant fusion protein from crude cell lysates. GST, a 26-kilodalton enzyme from Schistosoma japonicum, enables the purification of fusion proteins on immobilized glutathione under conditions that maintain protein activity and antigenicity (Amersham Biosciences). Following purification, the GST moiety can be proteolytically cleaved from NAAP at specifically engineered sites. FLAG, an 8-amino acid peptide, enables immunoaffinity purification using commercially available monoclonal and polyclonal anti-FLAG antibodies (Eastman Kodak). 6-His, a stretch of six consecutive histidine residues, enables purification on metal-chelate resins (QIAGEN). Methods for protein expression and purification are discussed in Ausubel et al. (supra, ch. 10 and 16). 15 Purified NAAP obtained by these methods can be used directly in the assays shown in Examples XVII, XVIII, and XIX, where applicable.

XIV. Functional Assays

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NAAP function is assessed by expressing the sequences encoding NAAP at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include PCMV SPORT plasmid (Invitrogen, Carlsbad CA) and PCR3.1 plasmid (Invitrogen), both of which contain the cytomegalovirus promoter. 5-10 μ g of recombinant vector are transiently transfected into a human cell line, for example, an endothelial or hematopoietic cell line, using either liposome formulations or electroporation. 1-2 μ g of an additional plasmid containing sequences encoding a marker protein are co-transfected. Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; Clontech), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated, laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties. FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium

iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M.G. (1994; Flow Cytometry, Oxford, New York NY).

The influence of NAAP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding NAAP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding NAAP and other genes of interest can be analyzed by northern analysis or microarray techniques.

XV. Production of NAAP Specific Antibodies

NAAP substantially purified using polyacrylamide gel electrophoresis (PAGE; see, e.g., Harrington, M.G. (1990) Methods Enzymol. 182:488-495), or other purification techniques, is used to immunize animals (e.g., rabbits, mice, etc.) and to produce antibodies using standard protocols.

Alternatively, the NAAP amino acid sequence is analyzed using LASERGENE software (DNASTAR) to determine regions of high immunogenicity, and a corresponding oligopeptide is synthesized and used to raise antibodies by means known to those of skill in the art. Methods for selection of appropriate epitopes, such as those near the C-terminus or in hydrophilic regions are well described in the art (Ausubel et al., supra, ch. 11).

Typically, oligopeptides of about 15 residues in length are synthesized using an ABI 431A peptide synthesizer (Applied Biosystems) using FMOC chemistry and coupled to KLH (Sigma-Aldrich, St. Louis MO) by reaction with N-maleimidobenzoyl-N-hydroxysuccinimide ester (MBS) to increase immunogenicity (Ausubel et al., *supra*). Rabbits are immunized with the oligopeptide-KLH complex in complete Freund's adjuvant. Resulting antisera are tested for antipeptide and anti-NAAP activity by, for example, binding the peptide or NAAP to a substrate, blocking with 1% BSA, reacting with rabbit antisera, washing, and reacting with radio-iodinated goat anti-rabbit IgG.

XVI. Purification of Naturally Occurring NAAP Using Specific Antibodies

Naturally occurring or recombinant NAAP is substantially purified by immunoaffinity

chromatography using antibodies specific for NAAP. An immunoaffinity column is constructed by covalently coupling anti-NAAP antibody to an activated chromatographic resin, such as CNBr-activated SEPHAROSE (Amersham Biosciences). After the coupling, the resin is blocked and washed according to the manufacturer's instructions.

Media containing NAAP are passed over the immunoaffinity column, and the column is washed under conditions that allow the preferential absorbance of NAAP (e.g., high ionic strength buffers in the presence of detergent). The column is eluted under conditions that disrupt antibody/NAAP binding (e.g., a buffer of pH 2 to pH 3, or a high concentration of a chaotrope, such as urea or thiocyanate ion), and NAAP is collected.

XVII. Identification of Molecules Which Interact with NAAP

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NAAP, or biologically active fragments thereof, are labeled with ¹²⁵I Bolton-Hunter reagent (Bolton, A.E. and W.M. Hunter (1973) Biochem. J. 133:529-539). Candidate molecules previously arrayed in the wells of a multi-well plate are incubated with the labeled NAAP, washed, and any wells with labeled NAAP complex are assayed. Data obtained using different concentrations of NAAP are used to calculate values for the number, affinity, and association of NAAP with the candidate molecules.

Alternatively, molecules interacting with NAAP are analyzed using the yeast two-hybrid system as described in Fields, S. and O. Song (1989; Nature 340:245-246), or using commercially available kits based on the two-hybrid system, such as the MATCHMAKER system (Clontech).

NAAP may also be used in the PATHCALLING process (CuraGen Corp., New Haven CT) which employs the yeast two-hybrid system in a high-throughput manner to determine all interactions between the proteins encoded by two large libraries of genes (Nandabalan, K. et al. (2000) U.S. Patent No. 6,057,101).

XVIII. Demonstration of NAAP Activity

NAAP activity is measured by its ability to stimulate transcription of a reporter gene (Liu, H.Y. et al. (1997) EMBO J. 16:5289-5298). The assay entails the use of a well characterized reporter gene construct, LexA_{op}-LacZ, that consists of LexA DNA transcriptional control elements (LexA_{op}) fused to sequences encoding the E. coli LacZ enzyme. The methods for constructing and expressing fusion genes, introducing them into cells, and measuring LacZ enzyme activity, are well known to those skilled in the art. Sequences encoding NAAP are cloned into a plasmid that directs the synthesis of a fusion protein, LexA-NAAP, consisting of NAAP and a DNA binding domain derived from the LexA transcription factor. The resulting plasmid, encoding a LexA-NAAP fusion

protein, is introduced into yeast cells along with a plasmid containing the LexA_{op}-LacZ reporter gene.

The amount of LacZ enzyme activity associated with LexA-NAAP transfected cells, relative to control cells, is proportional to the amount of transcription stimulated by the NAAP.

Alternatively, NAAP activity is measured by its ability to bind zinc. A 5-10 µM sample solution in 2.5 mM ammonium acetate solution at pH 7.4 is combined with 0.05 M zinc sulfate solution (Aldrich, Milwaukee WI) in the presence of 100 µM dithiothreitol with 10% methanol added. The sample and zinc sulfate solutions are allowed to incubate for 20 minutes. The reaction solution is passed through a VYDAC column (Grace Vydac, Hesperia, CA) with approximately 300 Angstrom bore size and 5 µM particle size to isolate zinc-sample complex from the solution, and into a mass spectrometer (PE Sciex, Ontario, Canada). Zinc bound to sample is quantified using the functional atomic mass of 63.5 Da observed by Whittal, R. M. et al. ((2000) Biochemistry 39:8406-8417).

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In the alternative, a method to determine nucleic acid binding activity of NAAP involves a polyacrylamide gel mobility-shift assay. In preparation for this assay, NAAP is expressed by transforming a mammalian cell line such as COS7, HeLa or CHO with a eukaryotic expression vector containing NAAP cDNA. The cells are incubated for 48-72 hours after transformation under conditions appropriate for the cell line to allow expression and accumulation of NAAP. Extracts containing solubilized proteins can be prepared from cells expressing NAAP by methods well known in the art. Portions of the extract containing NAAP are added to [32P]-labeled RNA or DNA. Radioactive nucleic acid can be synthesized *in vitro* by techniques well known in the art. The mixtures are incubated at 25°C in the presence of RNase- and DNase-inhibitors under buffered conditions for 5-10 minutes. After incubation, the samples are analyzed by polyacrylamide gel electrophoresis followed by autoradiography. The presence of a band on the autoradiogram indicates the formation of a complex between NAAP and the radioactive transcript. A band of similar mobility will not be present in samples prepared using control extracts prepared from untransformed cells.

In the alternative, a method to determine methylase activity of NAAP measures transfer of radiolabeled methyl groups between a donor substrate and an acceptor substrate. Reaction mixtures (50 μ l final volume) contain 15 mM HEPES, pH 7.9, 1.5 mM MgCl₂, 10 mM dithiothreitol, 3% polyvinylalcohol, 1.5 μ Ci [methyl-³H]AdoMet (0.375 μ M AdoMet) (DuPont-NEN), 0.6 μ g NAAP, and acceptor substrate (e.g., 0.4 μ g [³⁵S]RNA, or 6-mercaptopurine (6-MP) to 1 mM final concentration). Reaction mixtures are incubated at 30°C for 30 minutes, then 65°C for 5 minutes.

Analysis of [methyl- 3 H]RNA is as follows: (1) 50 μ l of 2 x loading buffer (20 mM Tris-HCl, pH 7.6, 1 M LiCl, 1 mM EDTA, 1% sodium dodecyl sulphate (SDS)) and 50 μ l oligo d(T)-cellulose

(10 mg/ml in 1 x loading buffer) are added to the reaction mixture, and incubated at ambient temperature with shaking for 30 minutes. (2) Reaction mixtures are transferred to a 96-well filtration plate attached to a vacuum apparatus. (3) Each sample is washed sequentially with three 2.4 ml aliquots of 1 x oligo d(T) loading buffer containing 0.5% SDS, 0.1% SDS, or no SDS. (4) RNA is eluted with 300 μ l of water into a 96-well collection plate, transferred to scintillation vials containing liquid scintillant, and radioactivity determined.

Analysis of [methyl- 3 H]6-MP is as follows: (1) 500 μ l 0.5 M borate buffer, pH 10.0, and then 2.5 ml of 20% (v/v) isoamyl alcohol in toluene are added to the reaction mixtures. (2) The samples are mixed by vigorous vortexing for ten seconds. (3) After centrifugation at 700g for 10 minutes, 1.5 ml of the organic phase is transferred to scintillation vials containing 0.5 ml absolute ethanol and liquid scintillant, and radioactivity determined. (4) Results are corrected for the extraction of 6-MP into the organic phase (approximately 41%).

In the alternative, type I topoisomerase activity of NAAP can be assayed based on the relaxation of a supercoiled DNA substrate. NAAP is incubated with its substrate in a buffer lacking Mg²⁺ and ATP, the reaction is terminated, and the products are loaded on an agarose gel. Altered topoisomers can be distinguished from supercoiled substrate electrophoretically. This assay is specific for type I topoisomerase activity because Mg²⁺ and ATP are necessary cofactors for type II topoisomerases.

Type II topoisomerase activity of NAAP can be assayed based on the decatenation of a kinetoplast DNA (KDNA) substrate. NAAP is incubated with KDNA, the reaction is terminated, and the products are loaded on an agarose gel. Monomeric circular KDNA can be distinguished from catenated KDNA electrophoretically. Kits for measuring type I and type II topoisomerase activities are available commercially from Topogen (Columbus OH).

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ATP-dependent RNA helicase unwinding activity of NAAP can be measured by the method described by Zhang and Grosse (1994; Biochemistry 33:3906-3912). The substrate for RNA unwinding consists of ³²P-labeled RNA composed of two RNA strands of 194 and 130 nucleotides in length containing a duplex region of 17 base-pairs. The RNA substrate is incubated together with ATP, Mg²⁺, and varying amounts of NAAP in a Tris-HCl buffer, pH 7.5, at 37°C for 30 minutes. The single-stranded RNA product is then separated from the double-stranded RNA substrate by electrophoresis through a 10% SDS-polyacrylamide gel, and quantitated by autoradiography. The amount of single-stranded RNA recovered is proportional to the amount of NAAP in the preparation.

In the alternative, NAAP function is assessed by expressing the sequences encoding NAAP

at physiologically elevated levels in mammalian cell culture systems. cDNA is subcloned into a mammalian expression vector containing a strong promoter that drives high levels of cDNA expression. Vectors of choice include pCMV SPORT (Life Technologies) and pCR3.1 (Invitrogen Corporation, Carlsbad CA), both of which contain the cytomegalovirus promoter. 5-10 μg of recombinant vector are transiently transfected into a human cell line, preferably of endothelial or hematopoietic origin, using either liposome formulations or electroporation. 1-2 μg of an additional plasmid containing sequences encoding a marker protein are co-transfected.

Expression of a marker protein provides a means to distinguish transfected cells from nontransfected cells and is a reliable predictor of cDNA expression from the recombinant vector. Marker proteins of choice include, e.g., Green Fluorescent Protein (GFP; CLONTECH), CD64, or a CD64-GFP fusion protein. Flow cytometry (FCM), an automated laser optics-based technique, is used to identify transfected cells expressing GFP or CD64-GFP and to evaluate the apoptotic state of the cells and other cellular properties.

FCM detects and quantifies the uptake of fluorescent molecules that diagnose events preceding or coincident with cell death. These events include changes in nuclear DNA content as measured by staining of DNA with propidium iodide; changes in cell size and granularity as measured by forward light scatter and 90 degree side light scatter; down-regulation of DNA synthesis as measured by decrease in bromodeoxyuridine uptake; alterations in expression of cell surface and intracellular proteins as measured by reactivity with specific antibodies; and alterations in plasma membrane composition as measured by the binding of fluorescein-conjugated Annexin V protein to the cell surface. Methods in flow cytometry are discussed in Ormerod, M. G. (1994) Flow Cytometry, Oxford, New York NY.

The influence of NAAP on gene expression can be assessed using highly purified populations of cells transfected with sequences encoding NAAP and either CD64 or CD64-GFP. CD64 and CD64-GFP are expressed on the surface of transfected cells and bind to conserved regions of human immunoglobulin G (IgG). Transfected cells are efficiently separated from nontransfected cells using magnetic beads coated with either human IgG or antibody against CD64 (DYNAL, Inc., Lake Success NY). mRNA can be purified from the cells using methods well known by those of skill in the art. Expression of mRNA encoding NAAP and other genes of interest can be analyzed by northern analysis or microarray techniques.

Pseudouridine synthase activity of NAAP is assayed using a tritium (³H) release assay modified from Nurse et al. ((1995) RNA 1:102-112), which measures the release of ³H from the C₅

position of the pyrimidine component of uridylate (U) when 3 H-radiolabeled U in RNA is isomerized to pseudouridine (ψ). A typical 500 μ l assay mixture contains 50 mM HEPES buffer (pH 7.5), 100 mM ammonium acetate, 5 mM dithiothreitol, 1 mM EDTA, 30 units RNase inhibitor, and 0.1-4.2 μ M [5- 3 H]tRNA (approximately 1 μ Ci/nmol tRNA). The reaction is initiated by the addition of <5 μ l of a concentrated solution of NAAP (or sample containing NAAP) and incubated for 5 min at 37 $^{\circ}$ C. Portions of the reaction mixture are removed at various times (up to 30 min) following the addition of NAAP and quenched by dilution into 1 ml 0.1 M HCl containing Norit-SA3 (12% w/v). The quenched reaction mixtures are centrifuged for 5 min at maximum speed in a microcentrifuge, and the supernatants are filtered through a plug of glass wool. The pellet is washed twice by resuspension in 1 ml 0.1 M HCl, followed by centrifugation. The supernatants from the washes are separately passed through the glass wool plug and combined with the original filtrate. A portion of the combined filtrate is mixed with scintillation fluid (up to 10 ml) and counted using a scintillation counter. The amount of 3 H released from the RNA and present in the soluble filtrate is proportional to the amount of peudouridine synthase activity in the sample (Ramamurthy, V. (1999) J. Biol. Chem. 274:22225-22230).

In the alternative, pseudouridine synthase activity of NAAP is assayed at 30 °C to 37 °C in a mixture containing 100 mM Tris-HCl (pH 8.0), 100 mM ammonium acetate, 5 mM MgCl₂, 2 mM dithiothreitol, 0.1 mM EDTA, and 1-2 fmol of [³²P]-radiolabeled runoff transcripts (generated *in vitro* by an appropriate RNA polymerase, i.e., T7 or SP6) as substrates. NAAP is added to initiate the reaction or omitted from the reaction in control samples. Following incubation, the RNA is extracted with phenol-chloroform, precipitated in ethanol, and hydrolyzed completely to 3-nucleotide monophosphates using RNase T₂. The hydrolysates are analyzed by two-dimensional thin layer chromatography, and the amount of ³²P radiolabel present in the ψMP and UMP spots are evaluated after exposing the thin layer chromatography plates to film or a PhosphorImager screen. Taking into account the relative number of uridylate residues in the substrate RNA, the relative amount ψMP and UMP are determined and used to calculate the relative amount of ψ per tRNA molecule (expressed in mol ψ /mol of tRNA or mol ψ /mol of tRNA/minute), which corresponds to the amount of pseudouridine synthase activity in the NAAP sample (Lecointe, F. et al. (1998) J. Biol. Chem. 273:1316-1323).

N²,N²-dimethylguanosine transferase ((m²₂G)methyltransferase) activity of NAAP is measured in a 160 μl reaction mixture containing 100 mM Tris-HCl (pH 7.5), 0.1 mM EDTA, 10 mM MgCl₂, 20 mM NH₄Cl, 1mM dithiothreitol, 6.2 μM S-adenosyl-L-[methyl-³H]methionine (30-70)

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Ci/mM), 8 µg m²₂G-deficient tRNA or wild type tRNA from yeast, and approximately 100 µg of purified NAAP or a sample comprising NAAP. The reactions are incubated at 30 °C for 90 min and chilled on ice. A portion of each reaction is diluted to 1 ml in water containing 100 µg BSA. 1 ml of 2 M HCl is added to each sample and the acid insoluble products are allowed to precipitate on ice for 20 min before being collected by filtration through glass fiber filters. The collected material is washed several times with HCl and quantitated using a liquid scintillation counter. The amount of ³H incorporated into the m²₂G-deficient, acid-insoluble tRNAs is proportional to the amount of N²,N²-dimethylguanosine transferase activity in the NAAP sample. Reactions comprising no substrate tRNAs, or wild-type tRNAs that have already been modified, serve as control reactions which should not yield acid-insoluble ³H-labeled products.

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Polyadenylation activity of NAAP is measured using an in vitro polyadenylation reaction. The reaction mixture is assembled on ice and comprises 10 µl of 5 mM dithiothreitol, 0.025% (v/v) NONIDET P-40, 50 mM creatine phosphate, 6.5% (w/v) polyvinyl alcohol, 0.5 unit/µl RNAGUARD (Pharmacia), 0.025 µg/µl creatine kinase, 1.25 mM cordycepin 5'-triphosphate, and 3.75 mM MgCl₂, in a total volume of 25 μ l. 60 fmol of CstF, 50 fmol of CPSF, 240 fmol of PAP, 4 μ l of crude or partially purified CF II and various amounts of amounts CF I are then added to the reaction mix. The volume is adjusted to 23.5 μ l with a buffer containing 50 mM TrisHCl, pH 7.9, 10% (v/v) glycerol, and 0.1 mM Na-EDTA. The final ammonium sulfate concentration should be below 20 mM. The reaction is initiated (on ice) by the addition of 15 fmol of 32 P-labeled pre-mRNA template, along with 2.5 μ g of unlabeled tRNA, in 1.5 µl of water. Reactions are then incubated at 30 °C for 75-90 min and stopped by the addition of 75 μ l (approximately two-volumes) of proteinase K mix (0.2 M Tris-HCl, pH 7.9, 300 mM NaCl, 25 mM Na-EDTA, 2% (w/v) SDS), 1 µl of 10 mg/ml proteinase K, 0.25 µl of 20 mg/ml glycogen, and 23.75 μ l of water). Following incubation, the RNA is precipitated with ethanol and analyzed on a 6% (w/v) polyacrylamide, 8.3 M urea sequencing gel. The dried gel is developed by autoradiography or using a phosphoimager. Cleavage activity is determined by comparing the amount of cleavage product to the amount of pre-mRNA template. The omission of any of the polypeptide components of the reaction and substitution of NAAP is useful for identifying the specific biological function of NAAP in pre-mRNA polyadenylation (Rüegsegger, U. et al. (1996) J. Biol. Chem. 271:6107-6113; and references within).

tRNA synthetase activity is measured as the aminoacylation of a substrate tRNA in the presence of [14C]-labeled amino acid. NAAP is incubated with [14C]-labeled amino acid and the appropriate cognate tRNA (for example, [14C]alanine and tRNA4b) in a buffered solution. 14C-

labeled product is separated from free [14C]amino acid by chromatography, and the incorporated 14C is quantified by scintillation counter. The amount of 14C-labeled product detected is proportional to the activity of NAAP in this assay.

In the alternative, NAAP activity is measured by incubating a sample containing NAAP in a solution containing 1 mM ATP, 5 mM Hepes-KOH (pH 7.0), 2.5 mM KCl, 1.5 mM magnesium chloride, and 0.5 mM DTT along with misacylated [14C]-Glu-tRNAGln (e.g., 1 μM) and a similar concentration of unlabeled L-glutamine. Following the quenching of the reaction with 3 M sodium acetate (pH 5.0), the mixture is extracted with an equal volume of water-saturated phenol, and the aqueous and organic phases are separated by centrifugation at 15,000 x g at room temperature for 1 min. The aqueous phase is removed and precipitated with 3 volumes of ethanol at -70°C for 15 min. The precipitated aminoacyl-tRNAs are recovered by centrifugation at 15,000 × g at 4°C for 15 min. The pellet is resuspended in of 25 mM KOH, deacylated at 65°C for 10 min., neutralized with 0.1 M HCl (to final pH 6-7), and dried under vacuum. The dried pellet is resuspended in water and spotted onto a cellulose TLC plate. The plate is developed in either isopropanol/formic acid/water or 15 ammonia/water/chloroform/ methanol. The image is subjected to densitometric analysis and the relative amounts of Glu and Gln are calculated based on the Rf values and relative intensities of the spots. NAAP activity is calculated based on the amount of Gln resulting from the transformation of Glu while acylated as Glu-tRNA^{Gln} (adapted from Curnow, A.W. et al. (1997) Proc. Natl. Acad. Sci. USA 94:11819-26).

20 XIX. Identification of NAAP Agonists and Antagonists

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Agonists or antagonists of NAAP activation or inhibition may be tested using the assays described in section XVIII. Agonists cause an increase in NAAP activity and antagonists cause a decrease in NAAP activity.

Various modifications and variations of the described compositions, methods, and systems of the invention will be apparent to those skilled in the art without departing from the scope and spirit of the invention. It will be appreciated that the invention provides novel and useful proteins, and their encoding polynucleotides, which can be used in the drug discovery process, as well as methods for using these compositions for the detection, diagnosis, and treatment of diseases and conditions.

Although the invention has been described in connection with certain embodiments, it should be understood that the invention as claimed should not be unduly limited to such specific embodiments.

Nor should the description of such embodiments be considered exhaustive or limit the invention to the

precise forms disclosed. Furthermore, elements from one embodiment can be readily recombined with elements from one or more other embodiments. Such combinations can form a number of embodiments within the scope of the invention. It is intended that the scope of the invention be defined by the following claims and their equivalents.

Table 1

Incyte Project ID	Polypeptide	Incyte	Polynucleotide	Incyte	
	SEQ ID NO:	Polypeptide ID	SEQ ID NO:	Polynucleotide	Incyte Full Length
				D .	Clones
7506140	1	7506140CD1	85	7506140CB1	
1889415	2	1889415CD1	65	1889415CB1	
7506047	3	7506047CD1	09	7506047CB1	
7505849	4	7505849CD1	19	7505849CB1	90179245CA2,
					90179313CA2
7505972	5	7505972CD1	62	7505972CB1	6717651CA2
7505991	9	7505991CD1	63	7505991CB1	
7506003	7	7506003CD1	64	7506003CB1	
6483977	8	6483977CD1	92	6483977CB1	8692157CA2
6301777	6	6301777CD1	99	6301777CB1	6301777CA2
7505976	10	7505976CD1	<i>L</i> 9	7505976CB1	
7506016	11	7506016CD1	89	7506016CB1	
7506086	12	7506086CD1	69	7506086CB1	
4176657	13	4176657CD1	70	4176657CB1	
7506056	14	7506056CD1	17	7506056CB1	
7506185	15	7506185CD1	72	7506185CB1	
8096611	91	8096611CD1	73	8096611CB1	
8174603	17	8174603CD1	74	8174603CB1	
3101042	18	3101042CD1	52	3101042CB1	
4972035	61	4972035CD1	9/	4972035CB1	
7506265	20	7506265CD1	77	7506265CB1	
7506304	21	7506304CD1	82	7506304CB1	
7506198	22	7506198CD1	61	180619027	90097138CA2
1381261	23	1381261CD1	80	1381261CB1	
6803876	24	6803876CD1	18	6803876CB1	5386286CA2
7506281	25	7506281CD1	82	7506281CB1	90070544CA2
7506175	26	7506175CD1	83	7506175CB1	
7506303	27	7506303CD1	84	7506303CB1	4996649CA2
7353336	28	7353336CD1	85	7353336CB1	

Incyte Project ID	Polypeptide	Incvte	Polynucleotide	Incyte	
,	SEO ID NO:	Polypeptide ID	SEO ID NO:	Polynucleotide	Incyte Full Length
				ID ,	Clones
3001652	29	3001652CD1	98	3001652CB1	
1689128	30	1689128CD1	28	1689128CB1	
2362969	31	2362969CD1	88	2362969CB1	
4753527	32	4753527CD1	68	4753527CB1	
6928688	33	6928688CD1	06	6928688CB1	
7506388	34	7506388CD1	91	7506388CB1	
7376372	35	7376372CD1	92	7376372CB1	
2754344	36	2754344CD1	93	2754344CB1	
8268822	37	8268822CD1	76	8268822CB1	
1814553	38	1814553CD1	56	1814553CB1	4521921CA2
71217830	39	71217830CD1	96	71217830CB1	3056930CA2
7506252	40	7506252CD1	<i>L</i> 6	7506252CB1	
2270608	41	2270608CD1	86	2270608CB1	3805161CA2
7502428	42	7502428CD1	-66	7502428CB1	
368741	43	368741CD1	100	368741CB1	7667604CA2
7506379	44	7506379CD1	101	7506379CB1	
7506253	45	7506253CD1	102	7506253CB1	8736487CA2
7506353	46	7506353CD1	103	7506353CB1	
7506372	47	7506372CD1	104	7506372CB1	
7506335	48	7506335CD1	105	7506335CB1	
5546982	49	5546982CD1	106	5546982CB1	
7507432	50	7507432CD1	107	7507432CBI	
5639578	51	5639578CD1	108	5639578CB1	
7509080	52	7509080CD1	109	7509080CB1	
7505899	53	7505899CD1	110	7505899CB1	
7505904	54	7505904CD1	111	7505904CB1	
7509224	55	7509224CD1	112	7509224CB1	
7505922	56	7505922CD1	113	7505922CB1	
7507695	57	7507695CD1	114	7507695CB1	

Table 2

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
_	7506140CD1	g2696611	0.0	[Rattus norvegicus] RNA splicing-related protein Imai, Y., et al. (1998) Cloning of a gene, YT521, for a novel RNA splicing-related protein induced by hypoxia/reoxygenation. Brain Res. Mol. Brain Res. 53, 33-40
		328064Rn.2155	0.0	[Rattus norvegicus][RNA-binding protein; Small molecule-binding protein] YT521, a nuclear protein with glutamic acid, proline and arginine rich regions, predicted to be a component of the spiceosomal complex, may play a role in the determination of pre-mRNA splice site selection Imai, supra.
		661120 FLJ2194 7.0E-28 0	7.0E-28	[Homo sapiens] Protein of unknown function, has a region of high similarity to a region of rat Rn.2155, which interacts with splicing factors and is induced by oxygen free radicals
2	1889415CD1	g6573115	0.0	[Mus musculus] p300 transcriptional cofactor JMY Shikama, N., et al. (1999) A novel cofactor for p300 that regulates the p53 response. Mol. Cell 4, 365-376
		618568[Jmy	0.0	[Mus musculus][Transcription factor] Junction mediating and regulatory protein, a transcription cofactor that forms a coactivator complex with p300 (CBP) and augments p53 (Trp53)-dependent transcription and apoptosis Partanen, A., et al. (1999) Developmentally regulated expression of the transcriptional cofactors/histone acetyltransferases CBP and p300 during mouse embryogenesis. Int. J. Dev. Biol. 43, 487-94
3	7506047CD1	g1511630	0.0	[Homo sapiens] homeodomain protein Zinovieva, R.D., et al. (1996) Structure and chromosomal localization of the human homeobox gene Prox 1. Genomics 35, 517-522

Table (

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score	Probability Score	Annotation
3 cont		337236 PROX1	0.0	[Homo sapiens][Transcription factor; DNA-binding protein] Prospero-related homeobox 1, a homeodomain-containing transcription factor that may be involved in development of the lens Torii Ma, et al. (1999) Transcription factors Mash-1 and Prox-1 delineate early steps in differentiation of neural stem cells in the developing central nervous system. Dev Suppl 126, 443-56
		325588 Prox1	0.0	[Mus musculus][Transcription factor; DNA-binding protein] Prospero-related homeobox 1, a member of the homeodomain-containing family of transcription factors, involved in the development of the lens, the lymphatic and central nervous systems, also plays a role in apoptosis, cell migration, and proliferation Wigle, J. T., and Oliver, G. (1999) Prox1 function is required for the development of the murine lymphatic system. Cell 98,
4	7505849CD1	g13559367	9.7E-79	[Homo sapiens] mitochondrial ribosomal protein L11 (L11mt) Suzuki, T., et al. (2001) Structural Compensation for the Deficit of rRNA with Proteins in the Mammalian Mitochondrial Ribosome. SYSTEMATIC ANALYSIS OF PROTEIN COMPONENTS OF THE LARGE RIBOSOMAL SUBUNIT FROM MAMMALIAN MITOCHONDRIA J. Biol. Chem. 276, 21724-21736
		475577 MRPL11 8.5E-80	8.5E-80	[Homo sapiens][RNA-binding protein; Ribosomal subunit][Cytoplasmic; Mitochondrial] Protein of the large 60S ribosomal subunit, has moderate similarity to S. cerevisiae Mrpl19p, which is a mitochondrial ribosomal protein of the large subunit
		276059 B0303.15 2.4E-29	2.4E-29	[Caenorhabditis elegans][RNA-binding protein; Ribosomal subunit][Cytoplasmic] Member of the ribosomal protein L12 protein family
5	7505972CD1	g12654167	1.6E-41	[Homo sapiens] cold inducible RNA-binding protein

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
5 cont		334674 CIRBP	1.4E-42	[Homo sapiens][RNA-binding protein][Nuclear] Cold inducible RNA-binding protein, an RNA-binding protein that suppresses cell proliferation in response to cold shock Nishiyama, H., et al. (1997) Cloning and characterization of human CIRP (coldinducible RNA-binding protein) cDNA and chromosomal assignment of the gene. Gene 204, 115-20
		584099 Cirbp	2.85.42	[Mus musculus][RNA-binding protein][Nuclear] Cold inducible RNA-binding protein, a glycine-rich RNA-binding protein that suppresses cell proliferation in response to cold shock, may play a role in biological rhythms Nishiyama, H., et al. (1998) Diurnal change of the cold-inducible RNA-binding protein (Cirp) expression in mouse brain. Biochem Biophys Res Commun 245, 534-8
9	7505991CD1	g12061189 1.0E-166 475561 LOC5100 9.1E-168 8	1.0E-166 9.1E-168	[Homo sapiens] ASC-1 complex subunit P50 [Homo sapiens] [RNA-binding protein] Protein containing a KH domain, which binds RNA
7	7506003CD1	g2460208	0:0	[Homo sapiens] RNA polymerase III largest subunit Sepehri, S. and Hernandez, N. (1997) The largest subunit of human RNA polymerase III is closely related to the largest subunit of yeast and trypanosome RNA polymerase III. Genome Res. 7, 1006-1019
		428328 RPC155 0.0	0.0	[Homo sapiens][Transferase; RNA polymerase subunit] Catalytic subunit of DNA directed RNA polymerase III Sepehri, supra. [Saccharomyces cerevisiae][Transferase; RNA polymerase subunit][Nuclear]
8	6483977CD1	g12654349	3.9E-116	RNA polymerase III, largest subunit [Homo sapiens] splicing factor, arginine/serine-rich 7 (35kD)

Table 2

	g gu	80	- ×	that
Annotation	[Homo sapiens][Spliceosomal subunit; RNA-binding protein][Nuclear] Splicing factor arginine serine rich 7, a splicing factor of the SR protein family containing an RNA binding domain (RBD) and a serine/arginine (SR) domain that is implicated in alternative pre-mRNA splicing Cavaloc, Y., et al. (1994) Characterization and cloning of the human splicing factor 9G8: a novel 35 kDa factor of the serine/arginine protein family. Embo Journal 13, 2639-49	[Mus musculus][Spliceosomal subunit; RNA-binding protein][Nuclear] Splicing factor arginine/serine-rich 3, a splicing factor of the SR family that contains an RRM (RNA recognition motif) domain, influences pre-mRNA splice site selection, regulates alternative splicing of its own mRNA, and is essential for development	[Homo sapiens] hairy and enhancer of split (Drosophila) homolog 2 [Rattus norvegicus][Inhibitor or repressor; DNA-binding protein; Transcription factor] Hairy and enhancer of split 2, contains a basic helix-loop-helix domain, predicted to repress transcription from promoters that contain E-box and N-box sequences Satow, T., et al. (2001) The basic helix-loop-helix gene hesr2 promotes gliogenesis in mouse retina. J Neurosci 21, 1265-73	[Homo sapiens] SCAN-related protein RAZ1 Sander, T.L., et al. (2000) Identification of a novel SCAN box-related protein that interacts with MZF1B. The leucine-rich SCAN box mediates hetero- and homoprotein associations. J. Biol. Chem. 275, 12857-12867
Probability Score	3.4E-117	4.3E-39	5.IE-36 1.6E-16	1.3E-60
GenBank ID NO: Probability or PROTEOME Score ID NO:	343788 SFRS7	588041 Sfrs3	588758 Hes2	.g7673373
ptide ID			6301777CD1	7505976CD1
Polypeptide SEQ Incyte ID NO: Polype	S Cont		6	10

Polypeptide SEQ Incyte		GenBank ID NO: Probability	Probability	Annotation
ON O	Polypeptide ID	or PROTEOME ID NO:	Score	
10 cont		749336 SCAND1 1.1E-61	1.1E-61	[Homo sapiens][Small molecule-binding protein] SCAN-related protein that associates with the MZF1B alternate splice form of the zinc finger transcription factor MZF1, contains a SCAN domain, which mediates hetero- and homeoprotein associations, and an arginine-rich region Castillo, G., et al. (1999) An adipogenic cofactor bound by the differentiation domain of PPARgamma. Embo Journal 18, 3676-87
		611238 Scand1	4.6E-16	[Mus musculus][Ligand][Nuclear] Peroxisome proliferative activated receptor gamma coactivator 2, transcriptional coactivator that binds to and increases the transcriptional activity of PPARgamma (Pparg), promotes adipogenesis
11	7506016CD1	g13278720	5.8E-273	[Homo sapiens] Similar to interferon regulatory factor 5
		.7437 16 IRF5	5.0E-274	[Homo sapiens][DNA-binding protein; Transcription factor] Interferon regulatory factor 5, a transcription factor that induces expression of members of the interferon A family, especially IFNA8, and IFNB1 in response to NDV viral infection Pitha, P. M., et al. (1998) Role of the interferon regulatory factors (IRFs) in virusmediated signaling and regulation of cell growth. Biochimie 80, 651-658
		429942 Irf5	4.2E-238	[Mus musculus][Transcription factor] Interferon regulatory factor 5, has strong similarity to human RF5, which is a transcription factor that induces expression of specific members of the interferon A family and human IFNB1 in response to NDV viral infection
12	7506086CD1	g541678	0.0	[Homo sapiens] hbZ17 Luna, L., et al. (1994) Molecular cloning of a putative novel human bZIP transcription factor on chromosome 17q22. Genomics 22, 553-562

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
12 cont		336592 NFE2L.1	0.0	[Homo sapiens][Activator; DNA-binding protein;Transcription factor][Nuclear] Nuclear factor erythroid derived 2-like 1, transcriptional activator, involved in heme biosynthesis, coordination of nuclear and mitochondrial gene expression, may have a role in mitochondrial genome maintenance and mitochondrial biogenesis Huo, L., and Scarpulla, R. C. (2001) Mitochondrial DNA instability and perimplantation lethality associated with targeted disruption of nuclear respiratory factor 1 in mice Mol Cell Biol 21, 644-54
		582223 Nfe211	7.3E-258	[Mus musculus][Activator; DNA-binding protein; Transcription factor] Nuclear factor erythroid derived 2-like 1, transcriptional activator, involved in erythropoiesis, oxidative stress response, mitochondrial genome maintenance, gastrulation, and mesoderm determination
13	4176657CD1	g6581093	2.1E-92	[Mus musculus] transposase-like protein Smit, A.F.A. (1999) Interspersed repeats and other mementos of transposable elements in mammalian genomes. Curr. Onin. Genet. Dev. 9, 657-63
14	7506056CD1	g2832260	1.0E-271	[Homo sapiens] DNA polymerase epsilon small subunit Jokela, M., et al. (1998) The small subunits of human and mouse DNA polymerase epsilon are homologous to the second largest subunit of the yeast Saccharomyces cerevisiae DNA polymerase epsilon. Nucleic Acids Res. 26, 730-734
		337072 POLE2	1.1E-270	[Homo sapiens][Regulatory subunit; Transferase; DNA polymerase or subunit] Polymerase epsilon 2, an accessory subunit of DNA polymerase epsilon, binds the catalytic subunit and stabilizes the polymerase complex Jokela, supra
		675635 Pole2	2.0E-32	[Mus musculus][Regulatory subunit; Transferase; DNA polymerase or subunit] Polymerase epsilon 2, small subunit of DNA polymerase epsilon
15	7506185CD1	lg1517816	0:0	[Homo sapiens] helicase

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score	Probability Score	Annotation
		ED NO:		
15 cont		347402 DDX12	0.0	[Homo sapiens][Hydrolase; Helicase; DNA-binding protein; ATPase][Nuclear; Nuclear nucleolus] DEAD box protein 12, a member of the DEAD/H box family
				of RNA or DNA helicases, expressed only in proliferating cells
				Amann, J., et al, (1997) Characterization of putative human homologues of the
				yeast chromosome transmission fidelity gene, CHL1. J Biol Chem 272, 3823-32
		248326 M03C11. 2.7E-116	2.7E-116	[Caenorhabditis elegans][Helicase][Nuclear] Member of the kinetochore protein
		2		(tentative) protein family
91	8096611CD1	g3328235	3.9E-92	[Xenopus laevis] 14S cohesin RAD21 subunit; member of RAD21/SCC1/MCD1
				family
				Losada, A., et al. (1998) Identification of Xenopus SMC protein complexes
				required for sister chromatid cohesion. Genes Dev. 12, 1986-1997
		438159 Rad21	1.2E-90	[Mus musculus] Rad21 (S. Pombe) homolog, putative component of a cohesin
			_	complex involved in sister chromatid cohesion, cleavage during the metaphase-
				anaphase transition may allow chromatid separation
				Darwiche, N., et al. (1999) Characterization of the components of the putative
				mammalian sister chromatid cohesion complex. Gene 233, 39-47
		343734 RAD21	4.0E-90	[Homo sapiens] Rad21 (S. Pombe) homolog, component of a cohesin complex
				involved in sister chromatid cohesion, cleavage during the metaphase-anaphase
				transition may allow chromatid separation, expression is downregulated by
				hypoxia in tumor cells
17	8174603CD1	g23273980	0.0	testis nuclear RNA-binding protein-like [Homo sapiens]
		g673456	2.2E-54	[Mus musculus] testis nuclear RNA binding protein
				Schumacher, J.M. et al. (1995) Biol. Reprod. 52:1274-1283.
		330218 Adarb1	2.6E-34	[Rattus norvegicus][Hydrolase; RNA-binding protein][Nuclear] Double-stranded
				RNA adenosine deaminase, mRNA editing of the glutamate receptor by site-
			•	specific deamination of adenosines; overexpression of human (ADARB1) may
				play a role in Down Syndrome phenotype development.
				Melcher, T. et al. (1996) J. Biol. Chem. 271:31795-31798.

Polypeptide SEQ Incyte	Incyte	GenBank ID NO: Probability	Probability	Annotation
:0 - - -	Polypeptide ID	or PROTEOME ID NO:	Score	
17		334046 ADARB1 8.2E-32	8.2E-32	[Homo sapiens][Hydrolase; RNA-binding protein][Nuclear] Double-stranded
cont				RNA adenosine deaminase, mRNA editing of the glutamate receptor subunit B by
				site-specific deamination of adenosines; overexpression may play a role in Down
				Syndrome phenotype development.
				Mittaz, L. et al. (1997) Genomics 41:210-217.
18	3101042CD1	g7459861	7.7E-122	[Homo sapiens] Zinc finger protein ZNF45
		-P93	2.2E-125	[Homo sapiens] Member of the XRCC1-linked KRAB zinc-finger protein family,
				has similarity tomurine Zfp93.
				Shannon, M. et al. (1996) Genomics 33:112-120.
		609312 Zfp235	9.6E-115	[Mus musculus][Inhibitor or repressor; Transcription factor; DNA-binding
				protein] Protein containing sixteen C2H2 type zinc finger domains, which bind
				nucleic acids, and a KRAB (kruppel-associated box) domain which may mediate
				transcriptional repression.
19	4972035CD1	g14278861	0.0	[Homo sapiens] PHD zinc finger transcription factor
				Yochum, G. S. and Ayer, D. E. (2001) Mol. Cell. Biol. 21: 4110-4118.
		738566 PF1	0.0	[Homo sapiens] mSin3A-interacting protein, has two plant homeodomain (PHD)
				zinc fingers, may link the TLE corepressor and the mSin3A-histone deacetylase
				complex to repress transcription.
				Yochum, et al. (2001) supra.
20	7506265CD1	g6006558	1.0E-61	ribosomal protein \$18 [Homo sapiens]
		g3287678	1.5E-49	[Arabidopsis thaliana] Match to ribosomal S18 gene mRNA gb/Z28701, DNA
				gb Z23165 from A. thaliana. ESTs gb T21121, gb Z17755, gb R64776 and
				gb/R30430 come from this gene.
		252690 Y57G11 3.8E-51	3.8E-51	[Caenorhabditis elegans][RNA-binding protein][Cytoplasmic] Member of the
		C.16		ribosomal protein S18 protein family.

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:		Annotation
20 cont		430218 Rps18	1.3E-46	[Mus musculus][RNA-binding protein; Ribosomal subunit][Cytoplasmic] Ribosomal protein S18, a putative ribosomal protein; human RPS18 is overexpressed in various tumors. MacMurray, A. J., and Shin, H. S. (1992) Mamm. Genome 2: 87-95.
21	7506304CD1	g12652649	1.8E-28	[Homo sapiens] ribosomal protein L28
		82	2.1E-54	[Rattus norvegicus][Structural protein;RNA-binding protein; Ribosomal subunit][Cytoplasmic] Ribosomal protein L28, component of the large 60S ribosomal subunit; human RPL28 gene shows abnormal expression in human colorectal carcinoma cells. Wool, I. G. et al. (1990) Biochim. Biophys. Acta 1050: 69-73.
		82	4.0E-29	[Mus musculus][Structural protein] Ribosomal protein L28, component of the large 60S ribosomal subunit; human RPL28 gene shows abnormal expression in human colorectal carcinoma cells. Burke, P. S. et al. (1994) Gene 142:315-316.
22	7506198CD1	g11139704	0.0	[Homo sapiens] Pumilio 2
		697504 PUM2	0.0	[Homo sapiens][RNA-binding protein] Protein containing eight Pumilio-family (Puf) domains, which bind RNA, has a region of low similarity to a region of S. cerevisiae Mpt5p, which is required for high temperature growth, for recovery from alpha-factor arrest, and for normal lifespan.
		371884 SPAC222 1.6E-96 .02c	1.6E-96	[Schizosaccharomyces pombe] Pumilio family protein.
24	6803876CD1	g6526355	5.0E-63	[Homo sapiens] hMBF1alpha Kabe, Y. et al. (1999) J. Biol. Chem. 274:34196-34202.
		339778 EDF1	4.3E-64	[Homo sapiens] Endothelial differentiation-related factor I, a putative transcriptional coactivator that binds to calmodulin (CALMI), in a calcium-dependent manner, and to the TATA-binding protein (TBP), may be involved in
:				endoneila ceil growth and differentiation. Mariotti, M. et al. (2000) J. Biol. Chem. 275:24047-24051.

ID NO:
618910 Edf1 1.1E-63 [Mus musculus] Protein with very strong similarity to human EDF1, a putative transcriptional coactivator that binds to calmodulin (CALM1) and to the TATAbinding protein (TBP) and that may be involved in endothelial cell growth and differentiation.
g1289371 7.5E-179 [Homo sapiens] Ikaros/LyF-1 homolog Nietfeld, W. and Meyerhans, A. (1996) Immunol. Lett. 49:139-141.
343040 ZNFN1A 6.5E-180 [Homo sapiens][DNA-binding protein] Zinc finger protein subfamily 1A 1 (Ikaros), a zinc finger transcription factor that regulates the lymphopoietic system development and homeostasis; alterations in corresponding gene expression
contribute to hematological malignancies. Georgonoulos, K. et al. (1994) Cell 79:143-56.
581109 Znfn1a1 1.2E-156 [Mus musculus][Hydrolase; Activator; DNA-binding protein; Transcription factor [Include factor] Zinc finger protein subfamily 1A 1 (Ikaros), a zinc-finger transcription factor regulating the lymphopoietic system development and homeostasis, targets chromatin remodeling factors; human ZNFN1A1 expression is altered in human hematological malignancies. Taubenberger, J. K. et al. (1996) Cell. Immunol. 171:41-47.
g14211691 1.7E-238 [Homo sapiens] elongation factor 1A binding protein Mansilla, F. et al. (2001) Thesis: Structural Biology, IMSB Aarhus University, Gustav Wieds vej 10 C, Aarhus 8000 C, Denmark.
731425 BPOZ 7.0E-159 [Homo sapiens] BPOZ protein, contains an ankyrin repeat, bipartite nuclear localization signal, and two BTB/POZ domains, inhibits cell proliferation, may be involved in protein complex assembly and developmental processes; decreased expression is seen in ovarian tumors. Unoki, M., and Nakamura, Y. (2001) Oncogene 20:4457-4465.
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1.0E-220 2.2E-44

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score	Probability Score	Annotation
		į		
28		344416 HMG17 1.9E-45	1.9E-45	[Homo sapiens][DNA-binding protein][Nuclear] High mobility group 17, binds
cont				DNA with low specificity, involved in regulation of transcription and cell
				differentiation, overexpressed in chronic myelogenic leukemia and may be
				mutated in various neoplasms.
				Kondos, H. et al. (1995) Biochem. Mol. Biol. Int. 36:803-809.
		587719 Hmgn2	4.6E-44	[Mus musculus][DNA-binding protein][Nuclear] High mobility group 17, binds
				DNA with variable sequence specificity, involved in regulation of transcription
				and cell differentiation; human HMG17 is overexpressed in chronic myelogenic
				leukemia.
				Bustin, M. et al. (1995) DNA Cell. Biol. 14:997-1005.
29	3001652CD1	g12483904	1.SE-117	[Rattus norvegicus] zinc finger protein HIT-39
		310825 Hs. 18375 0.0	0.0	[Homo sapiens] [Inhibitor or repressor; Transcription factor] Protein containing
		5		KRAB (kruppel-associated box) domains which may mediate protein-protein
				intereactions, contains C2H2 type zinc finger domains, which bind nucleic acids
		308339 ZNF184	5.2E-82	[Homo sapiens] Zinc finger protein 184, a member of the Kruppel zinc finger
				protein family, contains tandomly repeated C2H2-type zinc finger motifs at the C-
				terminus, highly expressed in testis
				Goldwurm, S. et al. (1997) Genomics 40:486-489
				Identification of a novel Krueppel-related zinc finger gene (ZNF184) mapping to
				6p21.3.
30	1689128CD1	g1216477	7.9E-21	[Mus musculus] zinc finger protein 60
				Perez, M. et al. (1996) FEBS Lett. 387:117-121
				Zfp60, a mouse zinc finger gene expressed transiently during in vitro muscle
				differentiation.

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
30 cont		581073 Zfp37	1.6E-23	[Mus musculus] [DNA-binding protein; Transcription factor] Zinc finger protein-37, member of the KRAB (kruppel-associated box) zinc-finger protein family, is expressed in brain and testis and may regulate spermiogenesis de Luis, O., Lopez-Fernandez, L. A., and del Mazo, (1999) J. Exp Cell Res 249:320-326 Tex27, a gene containing a zinc-finger domain, is up-regulated during the haploid stages of spermatogenesis.
		567485 OAZ	3.3E-21	[Homo sapiens] [DNA-binding protein] Protein with very strong similarity to rat Rn.9981, which is a zinc finger protein that regulates olfactory neuronal differentiation by interacting with Olf-1/EBF transcription factors, contains twenty eight C2H2 type zinc finger domains
31	2362969CD1	g5360107	1.3E-117	[Homo sapiens] NY-REN-36 antigen Scanlan, M.J. et al. (1999) Int. J. Cancer 83:456-464 Antigens recognized by autologous antibody in patients with renal-cell carcinoma
		423317 KIAA029 3.9E-107 5	3.9E-107	[Homo sapiens] [DNA-binding protein] Protein containing a C2H2 type zinc finger domain, which bind nucleic acids, has a region of weak similarity to a region of murine Nfh (heavy subunit of neurofilament), which is an intermediate filament
		617824 NEFH	8.8E-11	[Homo sapiens] [Structural protein] [Cytoplasmic; Cytoskeletal] Heavy polypeptide of neurofilament, a structural protein of the cytoskeleton that likely regulates axonal caliber and synaptic transmission; variants may be associated with some cases of amyotrophic lateral sclerosis Robberecht, W. (2000) J Neurol 247:2-6 Genetics of amyotrophic lateral sclerosis.

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
32	4753527CD1	g4106464	6.7E-164	[Mus musculus] AE-1 binding protein AEBP2 He, G.P. et al. (1999) J. Biol. Chem. 274:14678-14684 Cloning and characterization of a novel zinc finger transcriptional repressor. A direct role of the zinc finger motif in repression.
		429622 Aebp2	5.7E-165	[Mus musculus] [Inhibitor or repressor; DNA-binding protein; Transcription factor] [Nuclear] AE-binding protein 2, a transcriptional repressor that contains three Cys2-His2 Kruppel type zinc finger motifs and binds the promoter of fatty acid-binding protein Ap2 (Fabp4), may play a role in the control of adipocyte differentiation He, G. P. et al. (1999) J Biol Chem 274:14678-14684 Cloning and characterization of a novel zinc finger transcriptional repressor. A direct role of the zinc finger motif in repression.
		716429 ZIC4	9.2E-14	[Homo sapiens] Protein containing three C2H2 type zinc finger domains, has high similarity to a region of mouse Zic1, which is a zinc finger transcriptional activator that may be involved in cerebellar differentiation and maintenance of cerebellar granule cells
33	6928688CD1	g10954044	2.3E-104	[Homo sapiens] KRAB zinc finger protein ZFQR Ran, Q. et al. (2001) Exp. Cell Res. 263:156-162 Characterization of a novel zinc finger gene with increased expression in nondividing normal human cells.
		731891 MGC440 2.0E-265 10	2.0E-265	[Homo sapiens] Protein containing 8 C2H2 type zinc finger domains, which bind nucleic acids, and a kruppel-associated box (KRAB) domain, which are involved in protein-protein interactions, has moderate similarity to human RBAK, which binds retinoblastoma protein RB1

Table 2

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
33 cont		622017ZBRK1	3.3E-105	[Homo sapiens] [Inhibitor or repressor; DNA-binding protein; Transcription factor] [Nuclear] Zinc finger and BRCA1-interacting protein with a KRAB domain 1, binds BRCA1 and represses GADD45A transcription through intron 3, inhibits quiescent cells stimulated with growth factors from entering cellcycle and maintains the nondividing state of cells Zheng, L. et al. (2000) Mol Cell 6:757-768 Sequence-Specific Transcriptional Corepressor Function for BRCA1 through a Novel Zinc Finger Protein, ZBRK1.
34	7506388CD1	g532762	4.5E-165	[Homo sapiens] AF-17 Prasad, R. et al. (1994) Proc. Natl. Acad. Sci. U.S.A. 91:8107-8111 Leucine zipper dimerization motif encoded by the AF-17 gene fused to ALL-1 (MLL) in acute leukemia.
		339960 MLLT10 0.0	0.0	[Homo sapiens] [DNA-binding protein; Transcription factor] [Nuclear] Myeloid-lymphoid or mixed-lineage leukemia translocated to 10, putative transcription factor, contains a leukemia associated protein (LAP) domain and a leucine zipper; fusion of the corresponding gene to MLL or PICALM is found in acute leukemias Salmon-Nguyen, F. et al. (2000) Cancer Genet Cytogenet 122:137-140 CALM-AF10 fusion gene in leukemias: simple and inversion-associated translocation (10;11).
		585303 MIIt10	0.0	[Mus musculus] [Inhibitor or repressor; DNA-binding protein; Transcription factor] [Nuclear; Cytoplasmic] Myeloid-lymphoid or mixed-lineage leukemia translocated to 10, putative transcription factor, contains a leukemia associated protein (LAP) domain and a leucine zipper; fusion of human MLLT10 to MLL or PICALM is found in acute leukemias

Table 2

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:		Annotation
35	7376372CD1	g1654112	0.0	[Homo sapiens] ras-responsive element binding protein Thiagalingam, A. et al. (1996) Mol. Cell. Biol. 16:5335-5345 RREB-1, a novel zinc finger protein, is involved in the differentiation response to Ras in human medullary thyroid carcinomas.
		337828 RREB1	0.0	[Homo sapiens] [Activator; DNA-binding protein; Transcription factor] [Nuclear] Ras responsive element binding protein 1, zinc-finger protein implicated in transcriptional activation response to Ras- or Raf-induced cell differentiation
		625895 ZNF197 6.9E-44		[Homo sapiens] [Transcription factor] Zinc finger protein 197, member of the zinc finger transcription factor family, contains twenty C2H2-type zinc finger motifs, high level expression is associated with thyroid papillary carcinomas Gonsky, R., et al. (1997) Nucleic Acids Res 25:3823-3831 Identification of rapid turnover transcripts overexpressed in thyroid tumors and thyroid cancer cell lines: use of a targeted differential RNA display method to select for mRNA subsets.
36	2754344CD1	g387424	4.2E-244	[Mus musculus] mdm-1 Snyder, L.C. et al. (1988) J. Biol. Chem. 263:17150-17158 A gene amplified in a transformed mouse cell line undergoes complex transcriptional processing and encodes a nuclear protein.
		606464 MDM1	9.1E-87	[Homo sapiens] [Nuclear] Protein with high similarity to murine Mdm1, which is a nuclear protein, and the gene for which is amplified in transformed cells
		585283 Mdm1	4.3E.80	[Mus musculus] [Nuclear] Nuclear protein; gene is amplified up to 30-fold in transformed murine cells and generates a variety of alternatively spliced messages Snyder, L. C. et al. (1988) J Biol Chem 263:17150-17158 A gene amplified in a transformed mouse cell line undergoes complex transcriptional processing and encodes a nuclear protein.

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
37	8268822CD1	g5360109	1.4E-90	[Homo sapiens] NY-REN-37 antigen Scanlan, M.J. et al. (1999) Int. J. Cancer 83:456-464 Antigens recognized by autologous antibody in patients with renal-cell carcinoma.
		691420 FLJ1180 1.3E-128 6	1.3E-128	[Homo sapiens] Antigen recognized by autologous antibody in a patient with renal cell carcinoma
38	1814553CD1	g514388	8.6E-13	[Musca domestica] transposase Atkinson, P.W. et al. (1993) Proc. Natl. Acad. Sci. U.S.A. 90:9693-9697 The hobo transposable element of Drosophila can be cross-mobilized in houseflies and excises like the Ac element of maize.
39	71217830CD1	g9837581	1.8E-26	(Drosophila melanogaster) WIBG Ohlstein, B. et al. (2000) Genetics 155:1809-1819 The drosophila cystoblast differentiation factor, benign gonial cell neoplasm, is related to DExH-box proteins and interacts genetically with bag-of-marbles.
40	7506252CD1	g6440969	0.0	[Homo sapiens] RECQL4 helicase Kitao, S. et al. (1999) Genomics 61 (3):268-276 Rothmund-thomson syndrome responsible gene, RECQL4: genomic structure and products.
		341164RECQL4	0.0	[Homo sapiens] [Hydrolase; Helicase; DNA-binding protein; ATPase] RecQ protein-like 4, a member of the RecQ family of DNA helicases that may play a role in DNA repair; mutations in the corresponding gene are found in patients with Rothmund-Thomson syndrome, which is associated with genomic instability and cancer Kitao, S. et al. (1999) Nat. Genet. 22:82-84 Mutations in RECQL4 cause a subset of cases of Rothmund-Thomson syndrome.
		662679 Recql	2.2E-22	[Mus musculus] [Hydrolase; Helicase; DNA-binding protein; ATPase] [Nuclear] DNA helicase Q1, has similarity to Escherichia coli RecQ, which is an ATP-dependent DNA helicase, highly expressed in testis and thymus

Table 2

NO: Polypeptide ID 2270608CD1 7502428CD1	ME Score	
7502428CD1		
7502428CD1	1.6E-66	[Mus musculus] Similar to TATA box binding protein (TBP)-associated factor, RNA polymerase II, CI, 130kD
NOTATOR	4.0E-125	[Homo sapiens] testis-specific RING Finger protein
NOTATO ST		Yoshikawa, T. et al. (2000) Biochim. Biophys. Acta 1493:349-355
POLITORE		Isolation of a cDNA for a novel human RING finger protein gene, RNF18, by the
POLITORE		virtual transcribed sequence (VTS) approach(1).
NOTATOR	18 3.5E-126	[Homo sapiens] Ring finger protein 18, a C3HC4 type member of the RING
NOWNER		finger that may be a component of ribonucleoprotein complexes, expressed
NOTATOR		preferentially in testis
140111001	.1 1.1E-33	[Homo sapiens] [DNA-binding protein; RNA-binding protein] Sjogren syndrome
140111001		antigen A1 (52 kD), a member of the Tripartite Motif protein family and
140111001		component of Ro-SSA ribonucleoprotein complexes; recongized by
1401110011		autoantibodies in Sjogren's Syndrome, maternal antibodies are linked to neonatal
1470111031		lupus erythematosus
FO117037		Eftekhari, P. et al. (2000) Eur J Immunol 30:2782-2790
MO141020		Anti-SSA/Ro52 autoantibodies blocking the cardiac 5-HT4 serotoninergic
3C17C07C		receptor could explain neonatal Jupus concenital heart block.
43 308/41CD1 g554/1	7.9E-122	[Mus musculus] Zfp-29
		Denny, P. and Ashworth, A. (1991) A zinc finger protein-encoding gene
		expressed in the post-meiotic phase of spermatogenesis. Gene 106:221-227
584163 Zfp29	29 6.7E-123	[Mus musculus][Transcription factor; DNA-binding protein] Zinc finger protein
		29, a putative transcription factor that may regulate post-meiotic germ cell gene
		expression, expressed specifically in post-meiotic round spermatids
		de Luis, O. et al. (1999) Tex27, a gene containing a zinc-finger domain, is up-
		regulated during the haploid stages of spermatogenesis. Exp. Cell. Res. 249:320-
		770

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
43 cont		432838/ZNF180	I.4E-122	[Homo sapiens] Zinc finger protein 180, a putative zinc-finger protein that may play a role in developmental processes Wang, R. et al. (1996) Identification of a locus of zinc finger genes in human chromosome 19q13.1-q13.3 region by fluorescence in situ hybridization. Somat. Cell Mol. Genet. 22:245-248
4	7506379CD1	g3702270 7.3E-70 625915 RPL18A 6.2E-71	7.3E-70 6.2E-71	[Homo sapiens] ribosomal protein L18a [Homo sapiens] ribosomal protein; RNA-binding protein; Ribosomal subunit [Cytoplasmic] Ribosomal protein L18a, a component of the 60S ribosomal subunit Kenmochi, N. (1998) A map of 75 human ribosomal protein genes. Genome Res. 8:509-523
		715420 E04A4.8 2.6E-41	2.6E-41	[Caenorhabditis elegans][RNA-binding protein][Cytoplasmic] Member of the ribosomal protein L20 protein family Piano, F. (2000) RNAi analysis of genes expressed in the ovary of Caenorhabditis elegans. Curr. Biol. 10:1619-1622
45	7506253CD1	B13543433 1.4E-106 339940 EIF4EL.3 1.2E-107	1.4E-106 1.2E-107	[Homo sapiens] eukaryotic translation initiation factor 4E-like 3 [Homo sapiens][RNA-binding protein; Translation factor] Eukaryotic translation initiation factor 4E-like 3, a putative translation factor that binds capped mRNA Rom, E. et al. (1998) Cloning and characterization of 4EHP, a novel mammalian eIF4E-related cap-binding protein. J. Biol. Chem. 273:13104-13109
		704255 2700069E09Rik	1.5E-92	[Mus musculus][RNA-binding protein; Translation factor] [Cytoplasmic] Eukaryotic translation initiation factor 4E-like 3, a putative translation factor and capped mRNA-binding protein that may play a role in embryogenesis Temeles, G. L. et al. (1994) Expression patterns of novel genes during mouse preimplantation embryogenesis. Mol. Reprod. Dev. 37:121-129
46	7506353CD1	g9651711	2.0E-43	[Mus musculus] arsenite inducible RNA associated protein

Table 2

Polypeptide SEQ Incyte ID NO:	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
46 cont		703161 Airap	1.7E-44	[Mus musculus] Arsenite inducible RNA associated protein, associates with RNA, may function in protecting cells from arsenite toxicity Sok, J. et al. (2001) Arsenite-inducible RNA-associated protein (AIRAP) protects cells from arsenite toxicity. Cell Stress Chaperones 6:6-15
		246742 aip-1	4.0E-25	[Caenorhabditis elegans] Confers C. elegans with resistance to arsenite; weakly similar to S. cerevisiae YNL155W Sok, J. et al. supra
47	7506372CD1	g19484175	0.0	Similar to splicing factor, arginine/serine-rich 8 (suppressor-of-white-apricot homolog, Drosophila) [Mus musculus]
		g508231	0.0	e Dros nn Num 1994) C nal don r-of-w
-		341234 SFRS8	0.0	[Homo sapiens][Spliceosomal subunit; RNA-binding protein] [Nuclear] Splicing factor arginine serine rich 8, a member of the SR protein family, regulates alternative splicing by influencing the selection of alternative 5' splice sites, affects alternative splicing of fibronectin, CD45 (PTPRC), and its own mRNA Sarkissian, M. et al. (1996) The mammalian homolog of suppressor-of-white-apricot regulates alternative mRNA splicing of CD45 exon 4 and fibronectin IIICS. J. Biol. Chem. 271:31106-31114
		276878 swp-1	5.2E-39	[Caenorhabditis elegans][RNA-binding protein] [Nuclear] Putative splicing regulator and member of the SWAP protein family Spikes, D. A. et al. (1994) SWAP pre-mRNA splicing regulators are a novel, ancient protein family sharing a highly conserved sequence motif with the prp21 family of constitutive splicing proteins. Nucleic Acids Res. 22:4510-4519

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
48	7506335CD1	g10121150	7.1E-80	[Homo sapiens] bHLH factor Hes4 Bessho, Y. et al. (2001) Hes7: a bHLH-type repressor gene regulated by Notch and expressed in the presomitic mesoderm. Genes Cells 6:175-185
		617990 LOC5780 6.1E-81 1	6.1E-81	(Homo sapiens) Protein of unknown function, has high similarity to a region of murine Hes1 (hairy and enhancer of split), which is a helix-loop-helix negative regulator of transcription
		583195 Hes1	8.7E-34	[Mus musculus][Inhibitor or repressor; DNA-binding protein; Transcription factor] Hairy and Enhancer of split homolog 1, a helix-loop-helix negative regulator of transcription that participates in differentiation and cell fate determination of neural and other tissues, elevated levels of expression is linked to development of lymphoma
				Cau, E. et al. (2000) Hes genes regulate sequential stages of neurogenesis in the olfactory epithelium. Dev. Suppl. 127:2323-2332
49	5546982CD1	g8648883	0.0	[Homo sapiens] zinc-binding protein
		600182 TRIM36 0.0	0.0	[Homo sapiens] Protein containing fibronectin type III repeats, and a C3HC4 type (RING) zinc finger, which may mediate protein-protein interactions, has low similarity to MID1, which is associated with Opitz syndrome
		430024 Mid2	4.9E-55	[Mus musculus][Cytoplasmic; Cytoskeletal] Midline 2, a member of the B-box family of putative transcriptional regulators, contains a fibronectin type III domain and localizes to microtubules Buchner, G. et al. (1999) MID2, a homologue of the Opitz syndrome gene MID1: similarities in subcellular localization and differences in expression during
50	7507432CD1	g16565963	6.7E-251	[Homo sapiens] (AF380576) SAM-dependent methyltransferase
		692064 MGC245 5.7E-252 4	5.7E-252	[Homo sapiens] Protein of unknown function, has high similarity to uncharacterized mouse 2810025A12Rik

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
SO cont		583139 Gt(ROSA)26asSo r	6.7E-29	[Mus musculus] Gene trap ROSA 26 antisense (Philippe Soriano), a ubiquitously expressed protein that is conserved in humans and in C. elegans Zambrowicz, B. P. et al. (1997) Disruption of overlapping transcripts in the ROSA beta geo 26 gene trap strain leads to widespread expression of betagalactosidase in mouse embryos and hematopoietic cells. Proc. Natl. Acad. Sci. U. S. A. 94:3789-3794
51	5639578CD1	g3688780 581749∏spyl	2.2E-94 1.9E-95	[Mus musculus] testis-specific Y-encoded-like protein Vogel, T. et al. (1998) Murine and human TSPYL genes: novel members of the TSPY-SET-NAP1L1 family. Cytogenet. Cell Genet. 81:265-270 [Mus musculus] TSPY-like, a member of the TSPY-SET-NAP1L1 family Vogel, T. et al. supra
		623732 KIAA072 9.2E-94	9.2E-94	[Homo sapiens] Member of the NAP family of nucleosome assembly proteins
52	7509080CD1	g9857987	0.0	[Homo sapiens] SMCY Shen,P. et al. (2000) Population genetic implications from sequence variation in four Y chromosome genes. Proc. Natl. Acad. Sci. U.S.A. 97:7354-7359
		341282 SMCY	0.0	[Homo sapiens] Selected mouse cDNA on the Y homolog, contains a zinc finger motif and epitopes of the minor histocompatibility antigen, H-Y, a male-specific protein that elicits transplant rejection in female recipients Agulnik, A. I. et al. (1999) Mouse H-Y encoding Smcy gene and its X chromosomal homolog Smcx. Mamm. Genome 10:926-929
		692410 Smcx	0.0	[Mus musculus] SMC homolog X chromosome, encoded by an X-linked gene that escapes X-inactivation Agulnik, A. I. et al. (1994) A novel X gene with a widely transcribed Y-linked homologue escapes X- inactivation in mouse and human. Hum. Mol. Genet. 3:879-884
53	7505899CD1	g13543524	1.8E-44	[Homo sapiens] ribosomal protein, large P2

WO 03/054219

Polypeptide SEQ Incyte ID NO: Polype	ptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
53 cont		337760 RPLP2	1.5E-45	[Homo sapiens][Structural protein; RNA-binding protein; Ribosomal subunit] [Cytoplasmic] Ribosomal protein large P2, an acidic phosphoprotein component of the large 60S ribosomal subunit; contains an antigen which generates autoantibodies in individuals with systemic lupus erythematosus (SLE) Takehara, K. et al. (1990) Systemic lupus erythematosus associated with antiribosomal P protein antibody. Arch. Dermatol. 126:1184-1186
		462464 p2	4.9E-24	[Aspergillus fumigatus][RNA-binding protein; Ribosomal subunit] Acidic ribosomal protein P2, acts as an allergen
24	7505904CD1	g5813803	1.1E-92	[Homo sapiens] double-stranded RNA-binding zinc finger protein JAZ Yang, M. et al. (1999) JAZ requires the double-stranded RNA-binding zinc finger motifs for nuclear localization. J. Biol. Chem. 274:27399-27406
		428812JJAZ	9.2E-94	[Homo sapiens][RNA-binding protein][Nuclear nucleolus; Nuclear] Just another zinc finger protein, contains four C2H2 zinc finger motifs, binds double-stranded RNA and RNA/DNA hybrids, induces apoptosis when overexpressed in mouse fibroblasts Yang, M. et al. <u>supra</u>
		346	3.8E-81	[Mus musculus][RNA-binding protein][Nuclear] Zinc finger protein 346 (just another zinc finger protein), contains four C2H2 zinc finger motifs, binds double-stranded RNA and RNA/DNA hybrids, induces apoptosis when overexpressed in fibroblasts Yang, M. et al. supra
55	7509224CD1	g18307966 g7158880	0.0 4.8E-225	splicing factor, arginine/serine-rich 12 [Homo sapiens] [Rattus norvegicus] serine-arginine-rich splicing regulatory protein SRRP86 Barnard,D.C. and Patton,J.G. (2000) Identification and characterization of a novel serine-arginine-rich splicing regulatory protein. Mol. Cell. Biol. 20:3049-3057

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
55 cont		610045 Ѕπр86	4.1E-226	[Rattus norvegicus][Spliceosomal subunit; RNA-binding protein] [Nuclear] Serine arginine-rich splicing regulatory protein 86, contains an RNA recognition motif and serine-arginine-rich domains, interacts with other serine-arginine-rich splicing factors, involved in splicing regulation and differential splice site selection Barnard, D. C., and Patton, J. G. <u>supra</u>
		241942[D2089.1 4.5E-46	4.5E-46	[Caenorhabditis elegans][RNA-binding protein][Nuclear] An SR protein, thought to be involved in mRNA splicing Longman, D. et al. (2000) Functional characterization of SR and SR-related genes in Caenorhabditis elegans. Embo J. 19:1625-1637
56	7505922CD1	g187291	1.7E-112	[Homo sapiens] MAD3 Haskill,S. et al. (1991) Characterization of an immediate-early gene induced in adherent monocytes which encodes ikB-like activity. Cell 65:1281-1289
		613187 NFKBIA 1.4E-113	1.4E-113	[Homo sapiens] [Inhibitor or repressor] [Nuclear; Cytoplasmic] Nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor alpha, involved in transcription regulation by inhibiting nuclear localization and DNA binding of transcription activator NFkB in unstimulated cells; mutated in Hodgkin lymphoma Jungnickel, B. et al. (2000) Clonal deleterious mutations in the IkappaBalpha gene in the malignant cells in Hodgkin's lymphoma. J. Exp. Med. 191:395-402
		585391 Nfkbia	2.8E-99	[Mus musculus][Inhibitor or repressor][Nuclear; Cytoplasmic] Nuclear factor of kappa light polypeptide gene enhancer in B-cells inhibitor alpha, required for postnatal development and plays a role in transcription regulation by inhibiting transcriptional activator NFkB; human NFKBIA is mutated in Hodgkin lymphoma Huxford, T. et al. (1998) The crystal structure of the IkappaBalpha/NF-kappaB complex reveals mechanisms of NF-kappaB inactivation. Cell 95:759-770
57	7507695CD1	g15488944	6.6E-292	[Homo sapiens] Similar to zinc finger protein 35 (clone HF.10)

Table 2

Polypeptide SEQ Incyte ID NO: Polype	Incyte Polypeptide ID	GenBank ID NO: Probability or PROTEOME Score ID NO:	Probability Score	Annotation
57 cont		338990 ZNF35 4.8E-255	4.8E-255	[Homo sapiens][Activator; Transcription factor; DNA-binding protein] Zinc finger protein 35, a transcriptional activator that may play role in cell cycle control; corresponding gene is down-regulated during terminal differentiation of leukemic myeloid cell lines and commonly deleted in several carcinomas Donti, E. et al. (1990) Localization of the human HF.10 finger gene on a chromosome region (3p21-22) frequently deleted in human cancers. Hum. Genet. 84:391-395
		581901 Zfp105 1.2E-233	1.2E-233	[Mus musculus] Zinc finger protein 105, highly expressed in testis, may function in spermatogenesis Przyborski, S. A. et al. (1998) Differential expression of the zinc finger gene Zfo105 during spermatogenesis. Mamm. Genome 9:758-762

Table (

Incy		Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
	ID Polypeptide		Phosphorylation Sites	Glycosylation Sites		and Databases
. ~	7506140CD1	709	S5 S35 S45 S49	N196 N341 N568	BRAIN PD129722: P467-G655	BLAST_PRODOM
			S86 S91 S103 S152		BRAIN PD105763: M1-E193	
			S164 S169 S170		BRAIN PD129718: R229-A293	
			S181 S190 S247			
			S253 S258 S263			
			S292 S300 S310	٠		
			S367 S376 S380			
			S409 S498 S519			
			S559 S570 S654			
			T57 T198 T240			
			T319 Y707	•		
					do EUKARYOTIC; RNA; RNP-1;	BLAST_DOMO
	. — 				DM07068[P09406 303-470: R657-R708	
l∞	1889415CD1	986	S108 S115 S118	N48	do REGULATORY;	BLAST_DOMO
			S155 S179 S209		DM05091 S54986 1-980: D456-G831; P804-A869	
			S222 S234 S250			
			S322 S354 S371			
		-	S435 S512 S558			
			S564 S617 S696			
			S830 S868 S886			
			S887 S895 S900			
			S957 S972 T188			
			T218 T278 T311			
			T440 T447 T641			
			T683 T762 T982			
			Y409 Y572 Y647			

Table 3

Analytical Methods and Databases	SPSCAN	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	HIMMER_PFAM	BLIMPS_BLOCKS
Signature Sequences, Domains and Motifs	signal_cleavage: M1-A39	HOMEOBOX DNABINDING NUCLEAR PROTEIN BLAST_PRODOM PROSPEROLIKE PROXI PROX TRANSCRIPTION REGULATION DEVELOPMENTAL PD041924: MI-K75; S67-P352	HOMEOBOX PROTEIN NUCLEAR DNABINDING BLAST_PRODOM TRANSCRIPTION REGULATION DEVELOPMENTAL ALTERNATIVE SPLICING PROSPEROLIKE PD017936: S353-L711	PROSPERO-LIKE DOMAIN DM06382 P29617 834- BLAST_DOMO 1402: L557-L711; L325-F348; Q192-E221; S409-P514	PROSPERO-LIKE DOMAIN DM06382[P34522]1-585: D541-L711	Ribosomal protein L11: R16-V130	Ribosomal protein L11 proteins BL00359: M1-E29, L40-K80, 197-V130
Potential Glycosylation Sites	N108 N250 N423						
Potential Phosphorylation Sites	S50 S109 S113 S127 S176 S180 S203 S243 S255 S272 S349 S408 S424 S482 S500 S522 S626 S682 T14 T96 T241 T328 T456 T625 T665 Y235					S121 S134 T78 T87	
Amino Acid Residues	714					991	
SEQ Incyte D Polypeptide NO: D	7506047CD1					7505849CD1	
SEQ ID NO:	3	_				4	

				- -										1	_				- 1		_			_			
Analytical Methods	and Databases	BLAST_PRODOM		01.00 7010	BLASI_DOMO				HMMER_PFAM		BLIMPS_BLOCKS		PROFILESCAN		PROFILESCAN	BLAST_PRODOM				BLAST_PRODOM			BLAST_DOMO				
Signature Sequences, Domains and Motifs		PROTEIN RIBOSOMAL RNABINDING L11 50S 60S L12 CHLOROPLAST L11P	MITOCHONDRION	PDU0136/: E13-V129	RIBOSOMAL PROTEIN LII	DM00681 F34204 10-101. E13-V127 DM00681 P53875 3-157: R16-V130	DM00681 P36250 1-140: E15-V130	DM00681 A64074 2-142: E15-D132	RNA recognition motif. (a.k.a. RRM, RBD, or: L8-	G79	Eukaryotic RNA-binding region RNP-1 proteins	BL00030: L8-F26, R47-N56	Eukaryotic putative RNA-binding region RNP-1	signature: F26-177	Nuclear transition protein 1 signature: G69-S113	PROTEIN RNABINDING NUCLEAR	RIBONUCLEOPROTEIN REPEAT BINDING	SPLICING FACTOR ALTERNATIVE	HETEROGENEOUS PD000013: L8-R78	RNABINDING PROTEIN PUTATIVE	COLDINDUCIBLE GLYCINERICH CIRP A18	HNRNP NUCLEAR RNPL PD050679: Y93-S136	RIBONUCLEOPROTEIN REPEAT	DM00012 P98179 1-83: M1-R78	DM00012 P38159 3-84: A2-R78	DM00012 B49418 3-84: A2-R78	DM00012 S53050 1-83: M1-177
	Glycosylation Sites											•															
Potential	Phosphorylation Sites								S113 S122 S129	T43 T139 Y93																	
Amino Acid	Residues								142									***									
SEQ Incyte	Polypeptide ID								7505972CD1																		
SEQ	βŜ	4 2							5																		

Table 3

SEQ	SEQ Incyte	Amino Acid Potential	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
8 8 9 8	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
5 cont	_				Eukaryotic putative RNA-binding region RNP-1 signature: R47-F54	MOTIFS
9	7505991CD1 317	317	S115 S189 S233 S288 S297 T62 T80 T118 T126 Y285		KH domain: T62-G107	HMMER_PFAM
					KH domain proteins famil PF00013: 173-184	BLIMPS_PFAM
7	7506003CD1	1359	S16 S24 S63 S118 S147 S357 S364 S692 S878 S884 S893 S941 S977 S1040 S1187 T165 T181 T199 T274 T280 T338 T391 T485 T531 T553 T557 T596 T733 T743 T843 T889 T1009 T1017 T1049 T1117 T1124 T1145 T1124 Y446 Y870	N163 N355 N634 N791 N815 N992 N1062 N1075 N1218	RNA polymerase alpha subunit: S248-G906	нммек_рғам
					RNA polymerase A/beta'/A" subunit: 11006-A1335	HIMMER_PFAM

Analytical Methods and Databases	BLMPS_BLOCKS	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM
Signature Sequences, Domains and Motifs	Eukaryotic RNA polymerase II heptapeptide repeat proteins BL00115: H48-186, D87-S118, H157-K178, L226-V266, G300-1328, 1328-G358, K359-V386, A387-N419, G447-D501, G502-A527, N528-A569, C632-N674, A676-C724, E746-M795, I796-A835, N836-T865, A866-S892, S893-C934, P1029-N1062, I1063-I1085, H1182-S1217, N1218-M1263	RNA POLYMERASE DNADIRECTED TRANSCRIPTION TRANSFERASE SUBUNIT CHAIN LARGEST PROTEIN ZINC PD000656: G327-G906; L250-N662	DNADIRECTED RNA POLYMERASE III LARGEST SUBUNIT EC 2.7.7.6 RPC155 TRANSFERASE TRANSCRIPTION ZINC ZINCFINGER NUCLEAR PROTEIN PD097339: L907-G988	RNA POLYMERASE SUBUNIT DNADIRECTED TRANSFERASE TRANSCRIPTION ZINC LARGEST PROTEIN NUCLEAR PD001238: V11-R131	DNADRECTED RNA POLYMERASE LARGEST SUBUNIT III TRANSFERASE TRANSCRIPTION ZINC ZINCFINGER PD020898: P132-D249
Potential Glycosylation Sites					
Potential Phosphorylation Sites					
Amino Acid Residues					
SEQ Incyte ID Polypeptide NO: ID					
SEQ NO:	7 cont				

Table 3

Analytical Methods and Databases	BLAST_DOMO	HMMER_PFAM	BLIMPS_BLOCKS	PROFILESCAN	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	SPSCAN	PROFILESCAN
Signature Sequences, Domains and Motifs	DNA-DIRECTED RNA POLYMERASE II DM00252 P04051 101-654: 1104-Q641 DM00252 P08968 108-641: 1104-V638 DM00252 P31813 53-611: L171-D609; C69-A161 DM00252 P27625 81-751: Q422-V638; L142-I421; C69-L250	RNA recognition motif. (a.k.a. RRM, RBD, or: V13-V79	Protamine P1 proteins BL00048: S123-R149	Eukaryotic putative RNA-binding region RNP-1 signature: F31-V77	SPLICING FACTOR, ARGININE/SERINERICH 7 FACTOR 9G8 NUCLEAR PROTEIN RNABINDING MRNA ALTERNATIVE PHOSPHORYLATION REPEAT PD056023: E80-H120	RIBONUCLEOPROTEIN REPEAT DM00012 P23152 5-82: E10-G84 DM00012 A46398 6-83: K12-G84 DM00012 Q09167 1-72: K12-E80	PAPILLOMAVIRUS E2 PROTEIN DM00171[P36783[1-378: R124-E223	signal_cleavage: M1-G51	ATP synthase gamma subunit signature: D9-G45
Potential Glycosylation Sites								N30	
Potential Phosphorylation Sites		S89 S123 S128 S144 S159 S167 S175 S183 S208 S215 Y14						S16	
Amino Acid Residues		226						76	
Incyte Polypeptide ID		6483977CD1						6301777CD1	
SEQ ID NO:	7 cont	œ						6	

SEO	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
А	ID Polypeptide		Phosphorylation	Glycosylation Sites		and Databases
Ö	ID		Sites			
9 cont					MYC-TYPE, 'HELIX-LOOP-HELIX' DIMERIZATION DOMAIN DM00051[P35429]1-95:	BLAST_DOMO
					MI-E47	
0	7505976CD1	124	S29 S39 T63 T106 T118		signal_cleavage: MI-P19	SPSCAN
=	7506016CD1	488	S158 S267 S291	N2 N95	Interferon regulatory factor transcription factor	HMMER_PFAM
			S297 S342 S430 S436 T71 T238		domain: T10-N123	
					Tryptophan pentad repeat proteins (IRF family) proteins BL00601: R16-G54, D79-P107, N378-G390	BLIMPS_BLOCKS
					Tryptophan pentad repeat (IRF family) signature: T71-PROFILESCAN P127	PROFILESCAN
					Interferon regulatory factor signature PR00267: R16- BLIMPS_PRINTS W35, L42-P55, D60-G77, P83-D105	BLIMPS_PRINTS
					INTERFERON PROTEIN TRANSCRIPTION REGULATION DNABINDING NUCLEAR	BLAST_PRODOM
				-	FACTOR ACTIVATOR REGULATORY	
		_			INDUCTION IRFS	
					PD003535: G195-E418; N123-D162	
					PD119810: G195-I282	
					PD002355: P11-V120	
					PD023740: P194-M419	

SEQ	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
ЭŠ	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
11 cont					RF FAMILY DM02899 Q02556 1-317: R16-V120; P218-E354; DAME E360	BLAST_DOMO
					D343-F300 DM02899[149359]1-355: R16-E354	
					DM02899 S57837 1-355: R16-E354	
					DM02899 Q00978 1-319: R16-D162; S283-A341; P197-L228	
					Tryptophan pentad repeat (IRF family) signature: W35-A68	MOTIFS
12	7506086CD1	576	S3 S143 S195 S244 N378	N378	signal_cleavage: M1-V28	SPSCAN
			S252 S278 S285			
			S306 S319 S332			
	-		S359 S386 S403			
			S416 S510 T79			
			T134 T168 T302			
			T558 Y77			
					bZIP transcription factors basic domain proteins	BLIMPS_BLOCKS
					BL00036: K467-K479	
					Fos transforming protein signature PR00042: N468-	BLIMPS_PRINTS
				1000	L484, L486-F507	
					FACTOR NUCLEAR RELATED DNABINDING	BLAST_PRODOM
					PROTEIN ERYTHROID TRANSCRIPTION	
					REGULATION ACTIVATOR NFE2	
					PD011575: M1-S252, L257-I320	
					FACTOR NUCLEAR PROTEIN DNABINDING	BLAST_PRODOM
					TRANSCRIPTION REGULATION ERYTHROID	
					RELATED ACTIVATOR NFE2	
					PD006162: Q324-1462	

	SEO Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
, Вö	Polypeptide ID		Phosphorylation Sites	Glycosylation Sites		and Databases
12 cont					BZIP TRANSCRIPTION FACTORS BASIC DOMAIN DM00107 A49672 534-692: C368-G527 DM00107 A49671 185-334: S416-F526	BLAST_DOMO
					do TRANSCRIPTION; NRFI; BASIC; ZIPPER; DM04996 A49672 161-441: P161-S224; G267-L337	BLAST_DOMO
					bZIP transcription factors basic domain signature: R463-R478, R464-R478, R465-R478	MOTIFS
13	4176657CD1	573	S116 S136 S256 S358 S391 S445 S463 S510 S546 T55 T79 T95 T183 T189 T218 T225 T485 T538 Y371	N216 N291	Cytosolic domain: T534-H573 Transmembrane domain: I514-L533 Non-cytosolic domain: M1-S513	TMHMMER
4	7506056CD1	501	S35 S37 S65 S77 S128 S242 S246 S315 S332 S482 S489 T96 T115 T149 T224 T318 T464 T492	N240 N298 N490	DNA POLYMERASE EPSILON SUBUNIT B II TRANSFERASE DNADIRECTED REPLICATION DNABINDING PD018390: K248-L498; L83-P303; K47-E80	BLAST_PRODOM
					DNA POLYMERASE EPSILON SUBUNIT B EC 2.7.7.1 II TRANSFERASE DNADIRECTED REPLICATION DNABINDING NUCLEAR PROTEIN PD144164: M1-E46	BLAST_PRODOM

Phosphorylation Glycosylation Sites		Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
Sites	<u>&</u>	lypeptide		Phosphorylation	Glycosylation Sites		and Databases
\$21 \$96 \$192 \$204 N3 N784 N829 DEAD and DEAH box families ATP-dependent \$212 \$291 \$485 \$520 T5 T48 \$1104 T180 T471 \$1104 T180 T471 \$1104 T180 T471 \$12 T556 \$1594 \$1594 \$1600 PDO04168: L563-V869; V748-S890 PDO04168: L563-V869; V748-S890 PELICASE PROTEIN ATPBINDING PNABINDING PNABINDING NUCLEAR DNG PNABINDING PNABINDING PNABINDING PNABINDING PNABINDING PDO06997: M1-D217 PD006997: M1-D217 PD0069997: M1-D217 PD006998: K247-N464 PELICASE CHL 1 PROTEIN POTENTIAL PD0069997: M1-D217 PD00699999999999999999999999999999999999	8		ï	Sites			
### 188 ### 1892 #### 1892 #### 1892 #### 1892 #### 1892 #### 1892 #### 1892 #### 1892 #### 1892 #### 1892 ####################################	1	506185CD1		S21 S96 S192 S204 S212 S291 S485	N3 N784 N829	DEAD and DEAH box families ATP-dependent helicases signatures: L367-Q417	PROFILESCAN
T180 T471 T512 T556 HELICASE PROTEIN ATPBINDING DNABINDING ATPDEPENDENT DNA REPAIR PROBABLE NUCLEAR DING PD004168: L563-V869; V748-S890 HELICASE PROTEIN ATPBINDING DNABINDING NUCLEAR DNA REPAIR CHL1 DNABINDING NUCLEAR DNA REPAIR CHL1 DNAREPAIR COMPLEMENTING PD009997: M1-D217 PD004698: K247-N464 HELICASE CHL1 PROTEIN POTENTIAL PD024600: A435-P562 DM03675[P26659] 180-727: E543-R887; G344-L438 DM03675[P18074] 181-728: A342-R866 DM03675[P06839] 182-728: K345-1865				S505 S520 T5 T48		,	
HELICASE PROTEIN ATPBINDING				T104 T180 T471			
HELICASE PROTEIN ATPBINDING DNABINDING ATPDEPENDENT DNA REPAIR PROBABLE NUCLEAR DING PD004168: L563-V869; V748-S890 HELICASE PROTEIN ATPBINDING DNABINDING NUCLEAR DNA REPAIR CHL1 DNAREPAIR COMPLEMENTING PD009997: M1-D217 PD004698: K247-N464 HELICASE CHL1 PROTEIN POTENTIAL PD004600: A435-P562 DEAH-BOX SUBFAMILY ATP-DEPENDENT HELICASES DM03675 P26659 180-727: E543-R867; G344-L438 DM03675 P26659 181-728: A342-R866 DM03675 P06839 182-728: K345-1865				T477 T512 T556 T594			
NA REPAIR AIR CHL 1 TIAL ENDENT 7; G344-L438 6 5						HELICASE PROTEIN ATPBINDING	BLAST_PRODOM
AIR CHL1 TIAL ENDENT 7; G344-L438 6 5			-			DNABINDING ATPDEPENDENT DNA REPAIR	
AIR CHL1 TIAL ENDENT 7; G344-L438 6 5						PROBABLE NUCLEAR DING	
AIR CHL 1 TIAL ENDENT 7; G344-L438 6 5						PD004168: L563-V869; V748-S890	
EAR DNA REPAIR CHL I EMENTING TEIN POTENTIAL ILY ATP-DEPENDENT 727: E543-R887; G344-L438 728: A342-R866 28: A342-R866 728: K345-1865	_					HELICASE PROTEIN ATPBINDING	BLAST_PRODOM
EMENTING TEIN POTENTIAL ILY ATP-DEPENDENT 727: E543-R887; G344-L438 728: A342-R866 28: A342-R866 728: K345-1865						DNABINDING NUCLEAR DNA REPAIR CHLI	
TEIN POTENTIAL ILY ATP-DEPENDENT 727: E543-R887; G344-L438 728: A342-R866 28: A342-R866 728: K345-I865						DNAREPAIR COMPLEMENTING	
TEIN POTENTIAL ILY ATP-DEPENDENT 127: E543-R887; G344-L438 128: A342-R866 28: A342-R866 128: K345-I865						PD009997: M1-D217	
44-L438						PD004698: K247-N464	
ILY ATP-DEPENDENT 727: E543-R887; G344-L438 728: A342-R866 728: K345-R866						HELICASE CHLI PROTEIN POTENTIAL	BLAST_PRODOM
SUBFAMILY ATP-DEPENDENT 6659 180-727: E543-R887; G344-L438 8074 181-728: A342-R866 6839 182-728: K345-1865						PD024600: A435-P562	
HELICASES DM03675 P26659 180-727: E543-R887; G344-L438 DM03675 P18074 181-728: A342-R866 DM03675 148087 181-728: A342-R866 DM03675 P06839 182-728: K345-1865	_					DEAH-BOX SUBFAMILY ATP-DEPENDENT	BLAST_DOMO
DM03675 P26659 180-727: E543-R887; G344-L438 DM03675 P18074 181-728: A342-R866 DM03675 P06839 182-728: K345-1865						HELICASES	
DM03675 P18074 181-728: A342-R866 DM03675 148087 181-728: A342-R866 DM03675 P06839 182-728: K345-1865						DM03675 P26659 180-727: E543-R887; G344-L438	
DM03675[1807728: A342-R866 DM03675[P06839]182-728: K345-1865						DM03675 P18074 181-728: A342-R866	
DM03675 P06839 182-728: K345-1865						DM03675 148087 181-728: A342-R866	
						DM03675 P06839 182-728: K345-1865	

Table 3

Analytical Methods and Databases		BLAST_PRODOM			BLAST_PRODOM					BLAST_DOMO		HMMER_PFAM				BLAST_PRODOM			
Signature Sequences, Domains and Motifs		PROTEIN REPAIR RAD21 DNA DOUBLESTRANDBREAK DAMAGE NUCLEAR COHESIN HOMOLOG CALCIUMBINDING	PD010068: M1-K86		REPAIR PROTEIN RAD21 DNA	DOUBLESTRANDBREAK HOMOLOG DAMAGE	PHOSPHORYLATION	PD018225: E102-E407	PD015293: E473-I555	do PW29; CALCIUM;	DM08108J0C4248J1-533: M1-D229; D148-1418 DM08108P30776J1-544: M1-D118: D216-T321	Adenosine-deaminase (editase) domain: S396-V579,	A262-R349			DEAMINASE ADENOSINE RNA PROTEIN	DOUBLE-STRANDED EDITING RNA-SPECIFIC	HYDROLASE ZINC RNA-BINDING PD041051:	L395-K550
Potential Glycosylation Sites		N134 N178 N265	•									N489							
Potential Phosphorylation	Sites	S9 S46 S136 S157 S169 S271 S307 S317 S383 S424	S425 S463 S471 S495 S512 T28 T88	T109 T142 T272 T331 T413 T429 T478 V105								S13 S41 S167 S185 N489	S386 S396 S398	S431 S491 T329	T343 T482 T504				
Amino Acid Residues		255										584							
SEQ Incyte ID Polypeptide	Ð	8096611CD1										8174603CD1							
SEQ ID	ÖZ	91										17		•					

Table 3

Residues Phosphorylation Glycosylation Sites Sites 554	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
554		Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
554						BLAST_PRODOM
554					HYDROLASE ZINC RNA-BINDING PD003961: H209-R349	
554						BLAST_DOMO
554					DM04832 F31400 300-710: L393-F377, L210-S304	
554						BLAST_DOMO
554					DM04852 P55265 762-1225: H209-P577, A153-L180	
554						BLAST_DOMO
554					DM04852 P55266 708-1174: H209-P577	
554		-			Leucine zipper pattern: L407-L428	MOTIFS
Y476-H498, Y336-H358, Y504-H326, Y364-H386, Y280-H302, Y420-H442, Y392-H414, Y448-H470, Y308-H330 Zinc finger, C2H2 type, domain proteins BL00028: C478-H494 C2H2-type zinc finger signature: PR00048: P279-S292, L435-G444 PROTEIN ZINC-FINGER METAL PD00066: H354 C366 PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304-E552, G220-F457, K244-F485, P195-H442, G164-F401	CDI	554			Zinc finger, C2H2 type: Y224-H246, F196-H218,	HMMER_PFAM
Y280-H302, Y420-H442, Y392-H414, Y448-H470, Y308-H330 Zinc finger, C2H2 type, domain proteins BL00028: C478-H494 C2H2-type zinc finger signature: PR00048: P279-S292, L435-G444 PROTEIN ZINC-FINGER METAL PD00066: H354 C366 PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304-E552, G220-F457, K244-F485, P195-H442, G164-E401					Y476-H498, Y336-H358, Y504-H526, Y364-H386,	
Y308-H330 Zinc finger, C2H2 type, domain proteins BL00028: C478-H494 C2H2-type zinc finger signature: PR00048: P279-S292, L435-G444 PROTEIN ZINC-FINGER METAL PD00066: H354 C366 PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304-E552, G220-F457, K244-F485, P195-H442, G164-E401					Y280-H302, Y420-H442, Y392-H414, Y448-H470,	
Zinc finger, C2H2 type, domain proteins BL00028: C478-H494					Y308-H330	
C2H2-type zinc finger signature: PR00048: P279- S292, L435-G444 PROTEIN ZINC-FINGER METAL PD00066: H354 C366 PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304- E552, G220-F457, K244-F485, P195-H442, G164-					Zinc finger, C2H2 type, domain proteins BL00028:	BLIMPS_BLOCKS
C2H2-type zinc finger signature: PR00048: P279- S292, L435-G444 PROTEIN ZINC-FINGER METAL PD00066: H354 C366 PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304- E552, G220-F457, K244-F485, P195-H442, G164- F401						
S292, L435-G444						BLIMPS_PRINTS
PROTEIN ZINC-FINGER METAL PD00066: H354 C366 PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304- E552, G220-F457, K244-F485, P195-H442, G164- F401					S292, L435-G444	
C366 PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW 1 PD017719: G304 E552, G220-F457, K244-F485, P195-H442, G164-F401					PROTEIN ZINC-FINGER METAL PD00066: H354 BLIMPS_PRODOM	BLIMPS_PRODOM
PROTEIN ZINC FINGER METAL BINDING DNA BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304- E552, G220-F457, K244-F485, P195-H442, G164- F401					C366	
BINDING ZINC FINGER PATERNALLY EXPRESSED ZN FINGER PW1 PD017719: G304-E552, G220-F457, K244-F485, P195-H442, G164-F401					NG DNA	BLAST_PRODOM
EXPRESSED ZN FINGER PW1 PD017719: G304- E552, G220-F457, K244-F485, P195-H442, G164- F401					BINDING ZINC FINGER PATERNALLY	
ESS2, G220-F457, K244-F485, P195-H442, G164- F401					EXPRESSED ZN FINGER PW1 PD017719: G304-	
F401					ESS2, G220-F457, K244-F485, P195-H442, G164-	
					F401	

			_			
Analytical Methods and Databases	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO	HMMER_PFAM BLIMPS_BLOCKS
Signature Sequences, Domains and Motifs	ZINC FINGER DNA BINDING PROTEIN METAL BINDING NUCLEAR ZINC FINGER TRANSCRIPTION REGULATION REPEAT PD000072: K334-C397, R472-E536, K418-C481, K446-C509, K362-C425, K390-C453	ZINC FINGER, C2H2 TYPE, DOMAIN DM00002 Q05481 789-829: V329-E368, Q355-E396, Q411-E451	ZINC FINGER, C2H2 TYPE, DOMAIN DM00002 Q05481 831-885: C313-E368, C285-E340	ZINC FINGER, C2H2 TYPE, DOMAIN DM00002]P08042[314-358: C341-H386	ZINC FINGER, C2H2 TYPE, DOMAIN DM00002 P52743 31-93: L323-H386	PHD-finger: S58-R105, V273-H321 Forkhead-associated (FHA) domain proteins profile BL50006: V836-A841, S858-T862
Potential Glycosylation Sites						N278 N328 N595 N606 N637 N665 N719 N856
Potential Phosphorylation Sires						S40 S50 S61 S125 S134 S144 S162 S240 S241 S280 S332 S371 S376 S48 S454 S54 S529 S55 S576 S729 S837 S874 S963 S989 T54 T103 T130 T250 T257 T425 T461 T819 T862 Y857
Amino Acid Residues						1004
SEQ Incyte ID Polypeptide NO: ID	ì					4972035CD1
S E SE	cont cont					61

Table (

SEQ	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
a ë	ID Polypeptide NO: ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
19 cont					HOMEODOMAIN; PATHOGENESIS; YMR075W; BLAST_DOMO DM02014 Q09819 49-172: E35-R105	BLAST_DOMO
					YMR075W; DM08124 Q09698 325-538: N221- V322	BLAST_DOMO
					HOMEODOMAIN; PATHOGENESIS; YMR075W; BLAST_DOMO DM02014 S52835 164-321: N55-C102	BLAST_DOMO
20	7506265CD1	123	S120 T18 T110 Y66		Ribosomal protein S13/S18: V35-R113, R14-K34	HMMER_PFAM
					Ribosomal protein S13 proteins BL00646: R14-V61, D81-R113	BLIMPS_BLOCKS
					Ribosomal protein S13 signature: G73-K124	PROFILESCAN
					RIBOSOMAL PROTEIN S13 DM00495 P34788 12- BLAST_DOMO 148: K25-S120, 112-K34	BLAST_DOMO
					RIBOSOMAL PROTEIN S13 DM00495 P25232 12- 148: 15-S120	BLAST_DOMO
					RIBOSOMAL PROTEIN S13 DM00495 P41094 12- BLAST_DOMO 148:K25-S120, 112-K34	BLAST_DOMO
					RIBOSOMAL PROTEIN S13 DM00495 P48151 13- BLAST_DOMO 149:K25-S120, 112-K34	BLAST_DOMO
					Ribosomal protein S13 signature: R92-Q105	MOTIFS
21	7506304CD1	112	S37 S67 S90 S95 T106	N12	Ribosomal L28e protein family: S2-R103	HMMER_PFAM
					PROTEIN 60S RIBOSOMAL L28 F6F22.24	BLAST_PRODOM
					PROBABLE C2E11.04 R11D1.8 PD010767: K47- P98	

Table 3

Analytical Methods	and Databases	HMMER_PFAM		BLIMPS_PFAM		BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM			BLAST_DOMO		BLAST_DOMO		BLAST_DOMO		BLAST_DOMO	
Signature Sequences, Domains and Motifs		Pumilio-family RNA binding repeat: E829-V863, E757-I791, D647-F681, G908-M942, E683-A717, E865-1899, R719-M754, A793-L827		Pumilio-family RNA bindi PF00806: L648-S656,	D768-K777, D840-Q848	KIAA0099 PROTEIN PD125779: N193-F358	KIAA0099 PROTEIN PD134496:A475-E683	KIAA0099 PROTEIN PD062403: V385-1471	PROTEIN MATERNAL PUMILIO	DEVELOPMENTAL REPEAT KIAA0099	PD039905: W45-P192	PUMILIO; 160K; SEGMENT; YLL013C;	DM00365 P25822 1269-1339: G807-F878	PUMILIO; 160K; SEGMENT; YLL013C;	DM00365 A46221 1270-1339:C808-F878	PUMILIO; 160K; SEGMENT; YLL013C;	DM00365 P25822 1197-1267: G733-Y806	PUMILIO; 160K; SEGMENT; YLL013C;	DM00365 A46221 1341-1415: A879-Y957
Potential	Glycosylation Sites	N367 N531 N578 N619																	
Potential	Phosphorylation Sites	S30 S82 S93 S102 N367 N531 N578 S136 S182 S217 N619 S580 S626 S891	T21 T673 T951 T956																
Amino Acid Potential		284																	
Incyte	ID Polypeptide NO: ID	7506198CD1 987																	
SEQ	ΘŻ	22																	

				_						
Analytical Methods and Databases	HMMER_PFAM	HMMER_PFAM	BLIMPS_BLOCKS	BLIMPS_PFAM	BLIMPS_PRODOM	BLAST_DOMO	BLAST_DOMO	MOTIFS	HMMER_PFAM	BLAST_PRODOM
Signature Sequences, Domains and Motifs	BTB/POZ domain: P14-T189	Zinc finger, C2H2 type: Y309-H331, H726-H748, F765-H787, L234-C256, Y365-H388, Y337-H359	Zinc finger, C2H2 type, domain proteins BL00028: C236-H252	BTB (also known as BR-C) PF00651: A43-F55	PROTEIN ZINC-FINGER METAL PD00066: H355- BLIMPS_PRODOM C367	POZ DOMAIN DM00509 S59069 1-171: P10-R56, A133-A161, A777-E817	POZ DOMAIN DM00509 S41647 11-189: D9-E57, V125-L150,	Zinc finger, C2H2 type, domain: C311-H331, C339-H359, C367-H388, C728-H748, C767-H787	Helix-turn-helix: 181-A135	MULTIPROTEIN BRIDGING FACTOR EDF1 PROTEIN mRNA UNKNOWN PD019913: S4-V80
Potential Glycosylation Sites	N937									
Potential Phosphorylation Sites	S145 S196 S261 S323 S328 S332 S397 S619 S629 S636 S669 S706 S754 S762 S827 S846 S853 S893 T353 T411 T552 T553 T571 T605 T722 T737 T836 T1007								S4 S41 S111 S134 T40 T58 T91	
Amino Acid Residues	1013								141	
SEQ Incyte ID Polypeptide NO: ID	1381261CD1								6803876CD1	
SEQ ID NO:	33								24	

Analytical Methods and Databases	BLAST_DOMO	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_DOMO	BLAST_DOMO	BLAST_DOMO
Signature Sequences, Domains and Motifs	REPRESSOR; CONTROL; RESTRICTION; REGULATOR; DM00461 P14327 35-98: V77-V137	Zinc finger, C2H2 type, domain proteins BL00028: C279-H295	PROTEIN TRANSCRIPTION REGULATION ACTIVATOR ZINC FINGER META-BINDING DNA-BINDING NUCLEAR ALTERNATIVE SPLICING PD010982: V54-D182	PROTEIN TRANSCRIPTION REGULATION ACTIVATOR ZINC FINGER METAL-BINDING DNA-BINDING NUCLEAR ALTERNATIVE SPLICING PD013352: S183-K275	PROTEIN DNA-BINDING ZINC FINGER METAL-BLAST_PRODOM BINDING NUCLEAR HUNCHBACK DEVELOPMENTAL GAP REPEAT TRANSCRIPTION PD004213: V276-H329	ZINC-FINGERS II DOMAIN DM03488 A56229 259-BLAST_DOMO 387; R201-R330	ZINC-FINGERS II DOMAIN DM03488 Q03267 302-BLAST_DOMO 426: S208-R330	IKAROS; LYF-1; TRANSCRIPTION; DNA; DM08434[A56229]197-257: G138-E200
Potential Glycosylation Sites		N60 N177						
Potential Phosphorylation Sites		S13 S46 S47 S49 S73 S91 S92 S141 S179 S183 S199 S208 S217 S242 T23 T106 T173 T213 T269 T325 Y108						
Amino Acid Residues		334						
SEQ Incyte ID Polypeptide NO: ID		7506281CD1						
SEQ ID NO:	24 cont	25						

Analytical Methods and Databases	BLAST DOMO		MOTIFS	SPSCAN	HMMER_PFAM	HMMER_PFAM	BLIMPS_PFAM	BLMMPS_PFAM	BLIMPS_PRODOM	BLAST_DOMO		BLAST_DOMO		BLAST_PRODOM							BLAST_PRODOM				HMMER_PFAM	BLIMPS_BLOCKS	PROFILESCAN
Signature Sequences, Domains and Motifs	IKAROS: LYF-1: TRANSCRIPTION: DNA:	DM08434 Q03267 233-289: G138-K195	Zinc finger, C2H2 type, domain: C279-H299	signal_cleavage: M1-A64	BTB/POZ domain: E220-L337, D99-R215	Ank repeat: W35-F67, M1-K34	BTB (also known as BR-C) PF00651: V128-F140	Ankyrin repeat proteins PF00651: L40-L55	Repeat protein ankyrin nucleic acid PD00078	IKAROS LYF-1 TRANSCRIPTION DNA	DM06485 Q10225 44-522: D5-174, H247-Y415	IKAROS LYF-1 TRANSCRIPTION DNA	DM06485 P40560 1-512: D5-V176, C246-V408	PROTEIN SPLICESOME-ASSOCIATED	NOISETTE GENE SAP SPLICING FACTOR	SF3A60 T13H5.4 PD014231: Y11-G300					PROTEIN SPLICESOME-ASSOCIATED	SPLICING FACTOR NOISETTE GENE SAP	SF3A60 T13H5.4 PRE-MRNA PD010787: N321-	L448	HMG14 and HMG17: P2-K90	HMG14 and HMG17 proteins BL00355: K18-K48	HMG14 and HMG17 signature: P2-P53
Potential Glycosylation Sites														N170 N360 N395													
Potential Phosphorylation	Sites			S9 S94 S231 S315 S416										S102 S112 S197	S198 S234 S312	S314 S316 S415 T3	T28 T32 T78 T144	T224 T257 T261	T299 T362 T397	T422 T438 Y107 Y132					S25 S29		
Amino Acid Residues				439										448											<u>8</u>		
Incyte Polypeptide	Ω			7506175CD1										7506303CD1											7353336CD1		
SEQ ID	NO:	Sout Sout		56										27											28		

Table 3

SEO	SEO Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
<u>a</u> ;	Polypeptide	Residues	Phosphorylation	ation Sites		and Databases
ÿ	3		Sites			
78						BLIMPS_PRINTS
cont					signature PR00925: K18-P32, P34-A46, K51-D61,	
					A70-A80	
					PROTEIN CHROMOSOMAL NONHISTONE B	BLAST_PRODOM
					NUCLEAR DNA-BINDING HMG17 HMG14	
					MULTIGENE FAMILY POLYMORPHISM	
					PD008914: P2-K104	
					NONHISTONE CHROMOSOMAL PROTEIN HMG-BLAST_DOMO	3LAST_DOMO
					17 DM07897 P05204 1-88: P2-K90	
			-		NONHISTONE CHROMOSOMAL PROTEIN HMG-BLAST_DOMO	3LAST_DOMO
					17 DM04876 P12902 1-103: P2-D102	
					NONHISTONE CHROMOSOMAL PROTEIN HMG-BLAST_DOMO	3LAST_DOMO
					17 DM04876 P02316 1-99: P2-D92	
					NONHISTONE CHROMOSOMAL PROTEIN HMG-BLAST_DOMO	BLAST_DOMO
					17 DM04876 P12274 1-101: P2-A86	
					HMG14 and HMG17 signature: R23-P32	MOTIFS
53	3001652CD1	692	S30 S39 S79 S173	N180	KRAB box: V519-A581, V29-E91	HMMER_PFAM
			S210 S285 S290			
	_		S329 S364 S366			
			S425 S436 S520			
	· · · · · ·		S529 S569 S693			
			T167 T279 T322			
			T353 T686 Y284			
						HMMER_PFAM
					H172-H194, H676-H698, Y256-H278, Y284-H306,	
					F340-H362, F704-H726, F200-H222	
		i			C2H2 type, domain proteins BL00028:	BLIMPS_BLOCKS
					C230-H246	

Table 3

										_	_	_	_					_	_	_
Analytical Methods	and Databases	BLIMPS_PRINTS	BLIMPS_PRODOM	BLIMPS_PRODOM	BLAST_PRODOM				BLAST_DOMO		BLAST_DOMO		BLAST_DOMO		BLAST_DOMO		MOTIFS	MOTIFS		
Signature Sequences, Domains and Motifs		C2H2-type zinc finger signature PR00048: P339- K352, L299-G308	PROTEIN ZINC-FINGER Metal-binding PD00066: H274-C286	PROTEIN ZINC FINGER ZINC PD01066: F521- G559	HYPOTHETICAL 87.8 KD PROTEIN	HYPOTHETICAL PROTEIN ZINC-FINGER METAL BINDING DNA-BINDING: PD 175969	R363-T518; PD172267: A581-R675; PD170284:	A92-L170; PD169183: R702-G769, R338-E403	KRAB BOX DOMAIN DM00605 P52738 3-77:	G517-W578, G27-W88	KRAB BOX DOMAIN DM00605 Q05481 10-83:	G517-S579, G27-S89	KRAB BOX DOMAIN DM00605 P17097 1-76: V29- BLAST_DOMO	G97, V519-E589	KRAB BOX DOMAIN DM00605 148689 11-85:	T518-W578, A28-Q93	Cell attachment sequence: R596-D598, R749-D751	Zinc finger, C2H2 type, domain: C174-H194, C202-	H222, C230-H250, C258-H278, C286-H306, C314-	H334, C342-H362, C678-H698, C706-H726
Potential	Glycosylation Sites															-				
Potential	Phosphorylation Sites																			
Amino Acid	Residues																			
Incyte	ID Polypeptide NO: ID																			
SEQ	ΩÖ	29 cont							<u> </u>											

Table 3

Analytical Methods	and Databases	HMMER_PFAM													BLIMPS_BLOCKS		BLAST_DOMO		FS.		-S			S		
Analyt	and Da				_						••••			_	BLIM		BLAS		MOTIFS		MOTIFS			MOTIFS	•	
Signature Sequences, Domains and Motifs		Zinc finger, C2H2 type: F817-H839, H402-H424,	F374-H396, F1019-H1041, Y897-H919, Y845-H867,	F1047-H1069, F433-H455, 1929-H952					•••						Zinc finger, C2H2 type, domain proteins BL00028:	C819-H835	ACROSIN DM03631 P12978 10-163: P311-	Q344,L310-P343, S241-P274	Cytochrome c family heme-binding site signature:	C1049-K1054	Immunoglobulins and major histocompatibility	complex proteins signature:	F1047-H1053	Zinc finger, C2H2 type, domain:	C376-H396, C404-H424, C819-H839, C1021-H1041,	
Potential	Glycosylation Sites	N35 N153 N199	N546 N748																							
Potential	Phosphorylation Sites	S17 S27 S44 S55	S107 S115 S131	S186 S257 S261	S320 S364 S366	S370 S387 S442	S447 S570 S648	S663 S728 S740	S792 S809 S868	S980 S987 T39 T65	T229 T234 T754	T767 T796 T874	T932 T938 T953	T959 Y556												
Amino Acid	Residues	1801																								
Incyte	ID Polypeptide NO: ID	1689128CD1 1081								-																
SEQ	ВÖ	ಜ	_																							

SEO	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
Q Ö	Polypeptide TD		Phosphorylation Sires	Glycosylation Sites		and Databases
31	2362969CD1	1007	S7 S8 S91 S112 S155 S182 S229 S302 S324 S373 S397 S448 S459 S465 S529 S544 S653 S692 S712 S716 S765 S785 S800 S886 T36 T131 T216 T220 T274 T383 T563 T617 T675 T720 T725 T891 T904	N6 N173 N223 N660 N670	Zinc finger, C2H2 type, domain proteins BL00028: H52-H68	BLIMPS_BLOCKS
					Cell attachment sequence: R619-D621	MOTIFS
					Zinc finger, C2H2 type, domain: C50-H73	MOTIFS
32	4753527CD1	511	S120 S132 S133 S148 S183 S206 S211 S217 S238 S255 S274 S358 S390 S442 S465 T81 T228	N336 N357 N414 N463	signal_cleavage: M1-G65	SPSCAN
					Zinc finger, C2H2 type: Y261-H286, F328-H352, F295-H322	HMMER_PFAM
					PROTEIN ZINC-FINGER METAL BINDING PD00066: H318-C330	BLIMPS_PRODOM
					PROTEIN NUCLEAR NUCLEOPORIN TRANSPORT PORE REPEAT COMPLEX GLYCOPROTEIN NUP214 P62 PD005717: S110- A260	BLAST_PRODOM

Table 3

Analytical Methods and Databases	BLAST_DOMO	MOTIFS	HMMER_PFAM	HMMER_PFAM	BLIMPS_BLOCKS	BLIMPS_PRINTS	BLIMPS_PRODOM	BLIMPS_PRODOM	BLAST_PRODOM	BLAST_PRODOM
Signature Sequences, Domains and Motifs	ACIDIC SERINE CLUSTER REPEAT DM03496 P32583 57-405: A33-Q249	Zinc finger, C2H2 type, domain: C263-H286, C264-H286, C330-H352	KRAB box: L23-E85	Zinc finger, C2H2 type: H214-H236, Y298-H320, Y326-H348, Y242-H264, Y270-H292, Y354-H376, H186-H208, H158-E180	Zinc finger, C2H2 type, domain proteins BL00028: C244-H260	C2H2-type zinc finger signature PR00048: P213- K226, L313-G322	PROTEIN ZINC-FINGER METAL BINDING PD00066: H232-C244	PROTEIN ZINC FINGER ZINC PD01066: F25-G63 BLIMPS_PRODOM	ZINC-FINGER METAL-BINDING DNA-BINDING BLAST_PRODOM PATERNALLY EXPRESSED ZNFINGER PW1 PD017719: G154-1377, L145-H348, G238-R378, G164-H376, P138-H292	ZINC-FINGER METAL-BINDING DNA-BINDING BLAST_PRODOM NUCLEAR REPEAT TRANSCRIPTION REGULATION PD001562: L23-E85
Potential Glycosylation Sites			N3 N427							
Potential Phosphorylation Sites			S9 S17 S24 S43 S100 S140 S168 S224 S252 S280 S329 S355 S391 S429 T33 T67 T179							
Amino Acid Residues			485							
Incyte Polypeptide ID			6928688CD1							
SEQ NO:	32 cont		33							

SEQ Incyte A	⋉	몆	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
Residues			Phosphorylation Sites	Glycosylation Sites		and Databases
					ZINC-FINGER DNA-BINDING PROTEIN METAL- BLAST_PRODOM	SLAST_PRODOM
					BINDING NUCLEAR TRANSCRIPTION	
•					REGULATION REPEAT PD000072: K212-C275,	
					K184-C247, K240-C303, K156-C219, K296-C359,	
					K268-C331	
					MYELOBLAST KIAA0211 ZINC-FINGER METAL-BLAST_PRODOM	BLAST_PRODOM
					BINDING DNA-BINDING PD149061: Q149-H376	
					KRAB BOX DOMAIN DM00605 P51523 5-79: S22- BLAST_DOMO	3LAST_DOMO
		7			P94	
					KRAB BOX DOMAIN DM00605 148689 11-85: S22- BLAST_DOMO	3LAST_DOMO
					P94	
					KRAB BOX DOMAIN DM00605 P51786 24-86: E20 BLAST_DOMO	3LAST_DOMO
					E79	
					KRAB BOX DOMAIN DM00605 P52736 1-72: L23- BLAST_DOMO	SLAST_DOMO
					P94	
		ı			ATP/GTP-binding site motif A (P-loop): G187-T194	MOTIFS
					Zinc finger, C2H2 type, domain:	MOTIFS
				-	C188-H208, C216-H236, C244-H264, C272-H292,	
		- 1			C300-H320, C328-H348, C356-H376	

Table 3

Analytical Methods	and Databases	HMMER_PFAM	BLIMPS_PFAM	BLAST_PRODOM					BLAST_DOMO		BLAST_DOMO				BLAST_DOMO		MOTIFS
Signature Sequences, Domains and Motifs		PHD-finger: G24-Q74	PHD-finger. PF00628: K159-F173	AF10 PROTEIN NUCLEAR CHROMOSOMAL	TRANSLOCATION PROTOONCOGENE ZINC-	FINGER MAF10 PD041462: P249-V581;	PD041062: T849-D962; PD024590: H595-D833;	PD132318: Q963-K1011	LEUCINE-ZIPPER DOMAIN DM07839 P55197 286-BLAST_DOMO	1026: T286-K1011	YPR031W (includes peregrin, AF-10, AF-17)	DM03695 P55197 18-284: M18-T285	DM03695[P55198]1-286: M18-S208, Q214-S274,	K319-S359, G441-S513, E296-R330	LEUCINE-ZIPPER DOMAIN DM07839 P55198 288-BLAST_DOMO	986: K305-R988	Leucine zipper pattern: L750-L771, L757-L778
Potential	Glycosylation Sites	N116 N134 N209 N272 N384 N442 N489 N525 N645 N648 N661 N703 N763 N810 N941															
Potential	Phosphorylation Sites	S4 S9 S17 S118 S204 S208 S215 S278 S303 S321 S359 S370 S376 S436 S456 S680 S686 S704 S811 T263 T286 T378 T462 T556 T663															
Amino Acid Potential	Residues	1011															
Incyte	D Polypeptide	7506388CD1															
SEQ	ДÖ	34			_												

	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
Dolynosatide		Dhoenhondation	ation Sites		and Databases
900		r nospnory ration Sites	Olycosylation Sites		
7376372CD1	1675	0118 6918 0118	V916 N1097	1414,	HMMER_PFAM
		S175 S180 S290	N1586 N1606	Y125-H147, I1246-H1268, H97-H119, Y1539-	
		S413 S433 S463	•	H1561, Y671-H693, Y66-H88, K1511-H1533, Y699-	
		S600 S740 S842		H722, F316-H338, F208-H230, F790-H813, T753-	
		S883 S903 S1122		H775, L235-H258	
		S1140 S1150 S1174			
		S1219 S1225 S1238			
		S1271 S1279 S1284			
		S1335 S1339 S1373			
		S1400 S1444 S1458			
		S1474 S1478 S1485			
		S1488 S1498 S1504			
		S1522 S1574 S1584			
		S1597 S1616 S1629			
		TS9 T225 T229			
		T243 T246 T395			
		TS14 TS44 TS94			
		T619 T709 T726			
		T952 T1099 T1134			
		T1149 T1190			
		T1199 T1204			
		T1267 T1289			
		T1324 T1350			
		T1354 T1358			
		T1378 T1422			
		T1549 T1649			

CEO	Incute	Amino Acid	Potential	Potential	Signature Sequences. Domains and Motifs	Analytical Methods
y E	TD Polymentide	Decidues	Phoenhorylation	Glycoevlation Sites		and Databases
ğ	rolypeptine ID	vesignes	Sites	control markets		
35 Cont					Zinc finger, C2H2 type, domain proteins BL00028: C1394-H1410	BLIMPS_BLOCKS
					C2H2-type zinc finger signature PR00048: L1526-	BLIMPS_PRINTS
					G1535, P124-N137	
					PROTEIN ZINC-FINGER META PD00066: H1529- BLIMPS_PRODOM	BLIMPS_PRODOM
					C1541	
					TRANSCRIPTION FACTOR RREBI ZINC-	BLAST_PRODOM
					FINGER METAL-BINDING DNA-BINDING	
					PD184063: Q260-S669; PD041346: M932-11246;	
					PD050228: R697-V754; PD058224: M20-D96	
					ATP/GTP-binding site motif A (P-loop): A1320-	MOTIFS
		•			S1327, A1393-S1400	
					Zinc finger, C2H2 type, domain: C68-H88, C99-	MOTIFS
					H119, C127-H147, C210-H230, C237-H258, C318-	
					H338, C645-H665, C701-H722, C755-H775, C792-	
				-	H813, C1248-H1268, C1394-H1414, C1513-H1533,	
					C1541-H1561	
38	2754344CD1	724	S54 S126 S132	N27 N215 N434	TRANSFORMED MOUSE 3T3 CELL DOUBLE	BLAST_PRODOM
			S155 S268 S408	N458	MINUTE I NUCLEAR PROTEIN MDMI	
	_		S502 S517 S577		NUCLEAR PROTEIN PD142497: F213-N667, F684-	
			S586 S652 S658		K723	
			S696 S703 T91			
			T199 T294 T387			
			T418 T436 T459			-
			T484 T525 T537			
			T569 T576 T582			
					NUCLEAR PROTEIN TRANSFORMED MOUSE	BLAST_PRODOM
					3T3 CELL DOUBLE MINUTE MDM1 MDM1A	
_	•				PD037805: M1-V212	

SEO	Incvte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
еŝ	Polypeptide ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
04	7506252CD1	1144	S27 S101 S134 S150 S180 S246 S257 S421 S520 S535 S798 S919 S938 S975 S1033 S1084 S1092 T47 T52 T139 T398 T465 T558 T645 T648 T713 T739		DEAD/DEAH box helicase: Q471-V541, 1604-S679	HMMER_PFAM
					ATP-DEPENDENT DNA HELICASE, PUTATIVE PD183838: L499-A583, E563-R710	BLAST_PRODOM
					DEPENDENT; HELICASE; ATP; DNA; DM01766 P15043 16-384: E474-T722, 1704-E763	BLAST_DOMO
					DEPENDENT; HELICASE; ATP; DNA; DM01766 P50729 8-376: G477-T722	BLAST_DOMO
					DEPENDENT; HELICASE; ATP; DNA; DM01766 S62467 511-893: Q479-E759	BLAST_DOMO
					DEPENDENT; HELICASE; ATP; DNA; DM01766 Q09811 511-893: Q479-E759	BLAST_DOMO
4	2270608CD1	217	S71 S101 S136 S214 T17 T196		Signal Peptide: M24-G49	HMMER
					Signal cleavage: M24-A80	SPSCAN
45	7502428CD1	309	S3 S30 S195 S278 N70 N293 T19 T179	N70 N293	SPRY domain: S195-H308	HMMER_PFAM
					RFP TRANSFORMING PROTEIN DM02346 P19474 59-337: E10-F193	BLAST_DOMO
					REP TRANSFORMING PROTEIN DM01944 P18892 355-477: S195-C305	BLAST_DOMO

SEO	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
<u></u> 自	ID Polypeptide		-	Glycosylation Sites		and Databases
ë	В		Sites			
5					RFP TRANSFORMING PROTEIN	BLAST_DOMO
cont	·-				DM01944 P19474 339-465: S195-F304	
	 				Serpins signature: V280-I290	MOTIFS
43	368741CD1	483	S6 S43 S68 S94	N161 N413 N465	Zinc finger, C2H2 type: Y179-H201, P151-H173,	HIMMER_PFAM
		···	S217 S348 S467		F459-H481, Y291-H313, Y235-H257, Y431-H453,	
	-		T70 T92 T97 T180		Y207-H229, F347-H369, F375-H397, F319-H341,	-
			T196		Y403-H425, Y263-H285	
					Zinc finger, C2H2 type, domain proteins BL00028:	BLIMPS_BLOCKS
					C349-H365	
					C2H2-type zinc finger signature PR00048: P234-	BLIMPS_PRINTS
					K247, L222-G231	
					PROTEIN ZINC-FINGER METAL BINDING	BLIMPS_PRODOM
					PD00066: H197-C209	
					PROTEIN ZINC FINGER METAL BINDING DNA BLAST_PRODOM	BLAST_PRODOM
					BINDING ZINC FINGER PATERNALLY	
	-				EXPRESSED ZNFINGER PW1 PD017719: G175-	
					F412, G148-H397, G343-H481	
				•	ZINC FINGER DNA BINDING PROTEIN METAL	BLAST_PRODOM
					BINDING NUCLEAR ZINC FINGER	
					TRANSCRIPTION REGULATION REPEAT	
					PD000072: K205-C268, R373-C436, R177-C240,	
					K289-C352, K261-C324	
					MYELOBLAST KIAA0211 ZINC FINGER METAL BLAST_PRODOM	BLAST_PRODOM
					BINDING DNA BINDING PD149061: C181-P402,	
		-			C265-H453	

SEQ	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
ΩÖ		Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
43					ZINC FINGER, C2H2 TYPE, DOMAIN DM00002	BLAST_DOMO
					QU3481 831-883: C380-E433, C130-F200, C408- P458	
					Q05481 789-829: Q366-E407, R198-E239, Q282-	
					E323, Q394-E435, R171-K210	
					P08042 272-312: Q226-E267, Q394-E435	
					P08042 314-358: C184-H229, C156-H201, C408-	
					H453, C212-H257, C380-H425	
					Zinc finger, C2H2 type, domain: C153-H173, C181-	MOTIFS
				•	H201, C209-H229, C237-H257, C265-H285, C293-	
					H313, C321-H341, C349-H369, C377-H397, C405-	
					H425, C433-H453, C461-H481	
4	7506379CD1	137	S18 S19 T6 T129 T130 Y24		Ribosomal L18ae protein family: T6-F137	HMMER_PFAM
					PROTEIN RIBOSOMAL L18A 60S L20	BLAST_PRODOM
					MULTIGENE FAMILY WUGSC: H_DJ1107K12.2	
					PUTATIVE YL17 PD008313:K11-V103	
					60S RIBOSOMAL PROTEIN L18A PD021789:	BLAST_PRODOM
					K104-F137	
					RAT RIBOSOMAL PROTEIN L18A DM03166	BLAST_DOMO
					S47353 1-140: Q27-F137	
					P41093 35-176: K11-F137	
					P47913(36-173: K11-F137	
					P05732 30-174: L12-F137	
45	7506253CD1 200	200	S41 S42 S127 S175 N22 N38 N139	N22 N38 N139	Eukaryotic initiation factor 4E: Q18-L192	HMMER_PFAM
			T24 T33 T141			
			T148 T154 T165			
			[T17]			

SEO	SEO Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
	Polypeptide		Phosphorylation	ation Sites	•	and Databases
ÖN	D		Sites			
45					Eukaryotic initiation factor 4E proteins BL00813:	BLIMPS_BLOCKS
Cont					K43-N86, S100-F115, M116-L159	
					INITIATION FACTOR TRANSLATION	BLAST_PRODOM
					EUKARYOTIC PROTEIN CAP-BINDING	
					SUBUNIT 4E EIF4E EIF4E PD003697: K32-K177	
					PROTEIN INITIATION FACTOR CAP-BINDING	BLAST_PRODOM
					4EHP EIF4E-LIKE 4ELP TRANSLATION 4E	
					PD101625: G14-K45	
					EUKARYOTIC INITIATION FACTOR 4E	BLAST_DOMO
					DM02130	
					P48598 52-258: D27-G180	
					P48600 1-205: V46-G183	
					P06730 8-216: E29-L188	
			,		I49644 33-217: E29-L188	
					Eukaryotic initiation factor 4E signature: D67-W90	MOTIFS
77	Т		011111111111111111111111111111111111111			DI PADE DEINITE
40	/506353CD1	123	S11 S102 T14 T110		DksA/17aR zinc finger signature PR00618: C5/-K68	BLIMPS_PRINTS
47	7506372CD1	874	S31 S84 S100 S137 N271		Surp module: H209-E262, Q457-Q508	HMMER_PFAM
			S198 S238 S279			
			S283 S304 S353			
			S477 S519 S528			
			SS62 S635 S665			
			S688 S694 S700			
			S731 S761 S769			
			S779 S816 S823			
			T164 T205 T425			
			TS65 T616 T721			
			T765 Y265			

					l			
Analytical Methods and Databases	BLIMPS_BLOCKS	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLAST_PRODOM	BLIMPS_BLOCKS	PROFILESCAN	BLAST_PRODOM
Signature Sequences, Domains and Motifs	Protamine PI proteins BL00048: R720-R746	SPLICING FACTOR, ARGININE/SERINE RICH 8 SUPPRESSOR OF WHITE APRICOT PROTEIN HOMOLOG TRANSCRIPTION REGULATION RNA BINDING mRNA REPEAT NUCLEAR PD137630: T266-V408	SUPPRESSOR OF WHITE APRICOT PROTEIN TRANSCRIPTION REGULATION RNA BINDING mRNA SPLICING PD011756: R73-Y265	SPLICING FACTOR, ARGININE/SERINE RICH 8 SUPPRESSOR OF WHITE APRICOT PROTEIN HOMOLOG TRANSCRIPTION REGULATION RNA BINDING mRNA REPEAT NUCLEAR PDI28724: N619-R673	SUPPRESSOR OF WHITE APRICOT PROTEIN HOMOLOG TRANSCRIPTION REGULATION RNA BINDING MRNA PD024346:1449-G537	Myc-type, 'helix-loop-helix' dimerization domain proteins BL00038: H40-R60	Myc-type, 'helix-loop-helix' dimenzation domain signature: S30-A80	PROTEIN DNA BINDING NUCLEAR REPRESSOR TRANSCRIPTION REGULATION HAIRY FACTOR HESI DEVELOPMENTAL PD012655: R61-P124
Potential Glycosylation Sites								
Potential Phosphorylation Sites						S30 S37 S38 S41 S58 S121 T52		
Amino Acid Residues						681		
SEQ Incyte ID Polypeptide NO: ID						7506335CD1		
SEQ TO NO.	47 cont					48		

Analytical Methods	and Databases	BLAST_DOMO					MOTIFS	HMMER_PFAM											HIMMER_PFAM	BLAST_PRODOM		٠	BLAST_PRODOM			MOTIFS	
Signature Sequences, Domains and Motifs		MYC-TYPE, 'HELIX-LOOP-HELIX' DIMERIZATION DOMAIN DM00051	P35428 31-119: R35-E88	S29712 1-83: A32-E8	A46231 31-125: A32-E88	F14003[28-110: A32-E88	Myc-type, 'helix-loop-helix' dimerization domain signature: E44-L59	B-box zinc finger.: P195-M237, A140-1180											Zinc finger, C3HC4 type (RING finger): C21-L50	PROTEIN FINGER MIDLINE ZINC FINGER RING BLAST_PRODOM	PUTATIVE TRANSCRIPTION FACTOR XPRF	FETAL PD012462: N123-H232	MIDLINE PROTEIN PUTATIVE	TRANSCRIPTION FACTOR XPRF FETAL	KIDNEY ISOFORM RING PD022167: R233-S403	Zinc finger, C3HC4 type (RING finger), signature:	C30-V43
Potential	Glycosylation Sites							N65 N123 N387	N468																		
Potential	Phosphorylation Sites							S55 S68 S93 S155	S158 S304 S311	S312 S369 S372	S380 S403 S450	S582 S591 S597	S604 S608 S639	T11 T51 T95 T128	T179 T205 T243	T289 T322 T355	T379 T388 T459	1546 1706 Y254									
Amino Acid	Residues							716																			
Incyte	Polypeptide ID							5546982CD1																			,
SEQ	ДÖ	48 cont						49				_															

SEQ	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
e ö	ID Polypeptide NO: ID	Residues	Phosphorylation Sites	Glycosylation Sites		and Databases
20	7507432CD1	503	S10 S47 S126 S181 S210 S292 S357 S431 T41 T54 T178 T179 T206 T222 T417 T456 Y325		Putative RNA methylase family UPF0020: R274-D434	HMMER_PFAM
21	5639578CDI	410	S2 S110 S137 S358 S362 T278		Nucleosome assembly protein (NAP): I248-Y313, S349-H388, L202-I245	HMMER_PFAM
					Nuclesosome assembly protein (NAP) PF00956: 184- BLIMPS_PFAM P94, D243-K283, F296-Y313, C348-S358	BLIMPS_PFAM
					PROTEIN NUCLEOSOME ASSEMBLY NUCLEAR BLAST_PRODOM ILIKE SET NAPI TESTIS SPECIFIC ASPARTIC ACID RICH PD003095: E211-S317, D356-E385	BLAST_PRODOM
					PROTEIN TESTIS SPECIFIC Y-ENCODED Y TSPY Y-ENCODED-LIKE PD035838: S323-Y381	BLAST_PRODOM
					N SIGNAL DOMAIN	BLAST_DOMO
					Q01534 46-240: P191-Q367 Q01105 1-213: L205-W374	
					P53997 -218: L202-W374 P13825 -193: I208-W374	

Table 3

SEO	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
e ë	ID Polypeptide		Phosphorylation Sites	Glycosylation Sites		and Databases
52	7509080CD1	1056	S20 S99 S153 S264 N223 N248 S329 S336 S363 S364 S396 S492 S576 S675 S723 S753 S833 S1010 S1024 T66 T166 T207 T213 T623 T685 T779 T908 T1027	N223 N248	ARID/BRIGHT DNA binding domain: E76-K172	HMMER_PFAM
					PHD-finger: 1259-A307	HMMER_PFAM
					jmjC domain: W434-F550	HMMER_PFAM
					jmjN domain: E13-E59	HMMER_PFAM
					C5HC2 zinc finger: C640-L693	HMMER_PFAM
					PHD-finger PF00628: C275-P289	BLIMPS_PFAM
					PROTEIN ZK593.4 MYELOBLAST KIAA0234 SMCY XE169 SMCX ALTERNATIVE SPLICING RETINOBLASTOMA PD014548: A651-V1031	BLAST_PRODOM
					PROTEIN INTERGENIC REGION XE169 NUCLEAR ZINC FINGER ZINC FINGER METAL	BLAST_PRODOM
					BINDING DNA BINDING PD005470: F316-L618, D592-L649	
					MYELOBLAST KIAA0234 SMCY XE169 PROTEIN SMCX ALTERNATIVE SPLICING	BLAST_PRODOM
					PD022457: Q118-Y258	A COCOCA TO A SA
					PROTEIN XE169 JUMONJI DEVELOPMENTAL ZKS93.4 MYELOBLAST KIAA0234 SMCY SMCX	BLASI_PRODOM
					ALTERNATIVE PD006843: M1-L75	

Table

				_	_	_			_			_	_			_								_
Analytical Methods and Databases	BLAST_DOMO	BLAST_DOMO		SPSCAN	HIMMER	HMMER_PFAM	BLIMPS_PRINTS	BLAST_PRODOM			BLAST_DOMO						HIMMER_SMART			HMMER_SMART		BLIMPS_BLOCKS		BLAST PRODOM
Signature Sequences, Domains and Motifs	SMCX_HUMAN XE169 DM08128 P41229 671- 1190: F604-V1031	FINGER; SMCX; SMCY; YDR096W; DM01930 IP412291377-669; K310-M603		signal_cleavage: M1-A68	Signal Peptide: M1-S17	60s Acidic ribosomal protein: S32-D103, M1-V30	=	RIBOSOMAL PROTEIN ACIDIC 60S	PHOSPHORYLATION P2 P1 L12 MULTIGENE	FAMILY PD001928: M1-D103	RAT ACIDIC RIBOSOMAL PROTEIN PI	DM00632	[P42899 1-114: M1-D103	S54179 1-112: M1-D103	S43109 1-113: M1-D103	P41099[1-113: M1-D103	Zinc finger, C2H2 type: T73-H97, Q134-H158, Y177- HMMER_SMART	H201		Zinc finger, U1 domain: F70-Y104, D131-K165,	G174-T208	Zinc finger, C2H2 type, domain proteins BL00028:	C136-H152	DSRBPZFA PD127377: P53-Q167, C75-P217
Potential Glycosylation Sites																	N140							
Potential Phosphorylation Sites				S19 S93													S21 S86 S94 S126	S168 S205 T112	T117 T169 T181		-			
Amino Acid Residues				103								-				- -	232							
SEQ Incyte ID Polypeptide NO: ID			_	7505899CD1													7505904CD1							
SEQ RO	S2 cont			53													24							

Analytical Methods	and Databases	MOTIFS	SPSCAN	HMMER_PFAM	BLIMPS_BLOCKS	BLAST_PRODOM		BLAST_PRODOM	·	BLAST_PRODOM	
Signature Sequences, Domains and Motifs		Zinc finger, C2H2 type, domain: C75-H97, C135-H158, C136-H158, C179-H201	signal_cleavage: M1-A31	RNA recognition motif. (a.k.a. RRM, RBD, or RNP domain): V68-I137	Protamine P1 proteins BL00048: R366-S392	PROTEIN DNA BINDING CODED FOR BY C. ELEGANS CDNA CHROMOSOME HOMOLOG	PD001830: R212-G447, K199-K417, K236-K465, P179-R375, E149-P365, D259-N493	PROTEIN TOPOISOMERASE I DNA ISOMERASE BLAST_PRODOM REPEAT DNA BINDING INTERMEDIATE	FILAMENT HEPTAD PD000422: D259-E483, K226- D424, K183-P407, E161-P364, K303-K487, Q170- K323, E286-K506	PROTEIN REPEAT TROPOMYOSIN COILED COIL ALTERNATIVE SPLICING SIGNAL	PRECURSOR CHAIN PD000023: K260-K465, K262 D479, A155-K360, R283-K506, K230-R434
Potential	Glycosylation Sites		N138 N223 N369 N454 N500				-				
Potential	Phosphorylation Sites		S59 S181 S203 S208 S213 S235 S243 S251 S371 S377 S386 S389 S392 S403 S404 S408 S427 S437 S441 S442 S457 S502 T43 T78								
Amino Acid	Residues		808								
SEQ Incyte	Polypeptide ID		7509224CD1								
SEQ.	ДÖ	S4 cont	55				-				

							_			_		_	_				_	_	
Analytical Methods and Databases	BLAST_DOMO	HMMER_PFAM	HMMER_SMART	BLIMPS_PFAM	BLAST_PRODOM		BLAST_PRODOM			RI AST DOMO					HMMER_PFAM				BLIMPS_BLOCKS
Signature Sequences, Domains and Motifs	TYPE B REPEAT REPEAT DM05511 S26650 1-1203: P148-R423, R197-K461, S198-A480 P18583 113-1296: P148-R423, R165-A480	Ankyrin repeat: N182-T214, L110-F142, R143-1175	Ankyrin repeat: D73-F103, L110-L139, L143-L172, N182-A211	Ankyrin repeat proteins PF00023B: G144-E153	PROTEIN MAJOR HISTOCOMPATIBILITY COMPLEX ENHANCER BINDING MAD3 PHOSPHORYLATION ANK REPEAT ECIGIKBA	PD015498: M1-Q111	PROTEIN MAJOR HISTOCOMPATIBILITY	COMPLEX ENHANCER BINDING MAD3	PHOSPHORYLATION ANK REPEAT ECIGIKBA	ANK VEIN PEPEAT DMOONE	P25963 167-201: C167-L202	P25963 131-165: L131-S166	P25963 58-93: E58-194	P25963 95-129: R95-L130	Zinc finger, C2H2 type: Y390-H412, Y502-H524,	Y334-H356, Y278-H300, Y474-H496, F362-H384,	F250-H272, F222-H244, F306-H328, Y418-H440,	Y446-H468	Zinc finger, C2H2 type, domain proteins BL00028: C336-H352
Potential Glycosylation Sites															N67 N292				
Potential Phosphorylation Sites		S32 S36 S219 S240 T248 T256									_				SS9 S139 S187 T54 N67 N292	T305 T330 T475			
Amino Acid Residues		274													527				
Incyte Polypeptide ID		7505922CD1													7507695CD1				
Ια	55 cont	26													57				

SEQ	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
A	ID Polypeptide	Residues	Phosphorylation	Glycosylation Sites		and Databases
ö	<u>a</u>		Sites			
23					C2H2-type zinc finger signature PR00048: P249.	BLIMPS_PRINTS
cont					S262, L489-G498	
					Zinc-finger metal binding protein PD00066: H492-	BLIMPS_PRODOM
					C504	
					ZINC FINGER PROTEIN ZINC FINGER METAL	BLAST_PRODOM
					BINDING DNA BINDING HF.10	
					DIFFERENTIATION NUCLEAR PD106077: Q32-	
					K220	
					PROTEIN ZINC FINGER METAL BINDING DNA BLAST_PRODOM	BLAST_PRODOM
_					BINDING ZINC FINGER PATERNALLY	
					EXPRESSED ZN FINGER PW1 PD017719: K214-	
					F455, G274-S515, Y334-H524, C196-1439	
		Į.			ZINC FINGER DNA BINDING PROTEIN METAL BLAST_PRODOM	BLAST_PRODOM
					BINDING NUCLEAR ZINC FINGER	
					TRANSCRIPTION REGULATION REPEAT	•
					PD000072: K276-C339, P445-C507, K248-C311	
					ZINC FINGER PROTEIN ZINC FINGER METAL	BLAST_PRODOM
	· ···				BINDING DNA BINDING PUTATIVE REX2	
					TRANSCRIPTION REGULATION PD033163:	
		_			K285-K416	
					ZINC FINGER, C2H2 TYPE, DOMAIN DM00002	BLAST_DOMO
					P52743 31-93: L265-H328, L377-H440	
					P13682 485-515: Q493-H524	
					P08042 314-358: C367-H412, C255-H300, C227-	
					H272, C451-H496	
					P13682 201-231: A209-H240	

Table 3

SEQ	Incyte	Amino Acid	Potential	Potential	Signature Sequences, Domains and Motifs	Analytical Methods
<u>e</u>	Polypeptide	Residues	Phosphorylation	Glycosylation Sites		and Databases
Ö	<u>O</u>		Sites			
57					Zinc finger, C2H2 type, domain: C224-H244, C252- MOTIFS	MOTIFS
cont					H272, C280-H300, C308-H328, C336-H356, C364-	
					H384, C392-H412, C420-H440, C448-H468, C476-	
					H496	

Polynucleotide SEQ ID NO:/	Sequence Fragments
Incyte ID/ Sequence Length	
58/7506140CB1/	1-251, 1-311, 3-240, 3-379, 12-273, 18-335, 19-286, 24-253, 29-348, 30-342, 30-822, 30-2615, 34-379, 35-379, 35-
2629	452, 41-351, 41-371, 43-372, 46-371, 55-261, 73-392, 77-371, 80-379, 84-371, 84-379, 86-374, 139-335, 141-624,
	161-379, 167-396, 241-906, 255-371, 261-371, 314-361, 355-1038, 368-601, 368-851, 421-716, 465-867, 470-906,
	485-904, 489-855, 551-790, 672-903, 676-1392, 677-1479, 864-1070, 892-1130, 892-1196, 892-1501, 892-1543,
	892-1754, 893-1078, 893-1088, 893-1381, 902-1115, 902-1624, 907-1185, 910-1164, 917-1683, 922-1389, 925-
	1521, 929-1552, 930-1189, 931-1173, 931-1490, 933-1520, 934-1504, 934-1516, 934-1569, 936-1198, 936-1225,
	938-1161, 938-1170, 940-1048, 941-1160, 942-1800, 943-1537, 944-1179, 948-1185, 949-1160, 949-1616, 949-
	1618, 952-1350, 954-1498, 955-1417, 957-1425, 957-1794, 958-1227, 958-1228, 959-1503, 967-1260, 969-1304,
	1010-1613, 1012-1509, 1019-1239, 1103-1777, 1107-1542, 1126-1386, 1137-1638, 1138-1784, 1146-1700, 1180-
	1452, 1209-1709, 1230-1748, 1254-1906, 1289-1868, 1293-1409, 1327-1820, 1345-1598, 1366-1674, 1372-1698,
	1380-1843, 1399-1899, 1405-1693, 1406-1634, 1411-1675, 1413-1717, 1419-1650, 1419-1742,
	1432-2164, 1451-1697, 1474-2091, 1481-1728, 1481-1766, 1494-1781, 1501-1736, 1502-1720, 1504-2109, 1516-
	2048, 1538-1788, 1563-1839, 1565-1823, 1568-1842, 1569-2304, 1585-1829, 1610-2084, 1612-2233, 1616-1908,
	1667-2124, 1703-1908, 1718-1908, 1746-2043, 1748-2227, 1755-2115, 1791-2081, 1804-2053, 1930-1985, 1933-
	1953, 1933-2006, 1940-2517, 1956-2179, 1958-2224, 1958-2233, 1958-2253, 1982-2198, 1982-2321, 1983-2201,
	1983-2322, 1992-2261, 1994-2234, 2015-2270, 2021-2285, 2036-2171, 2036-2293, 2144-2432, 2155-2414, 2201-
	2490, 2276-2613, 2328-2614, 2343-2629, 2352-2586, 2354-2496, 2365-2629, 2366-2616, 2367-2625, 2372-2595,
	2390-2501, 2390-2615, 2430-2609, 2443-2591
59/1889415CB1/	1-647, 300-984, 481-842, 481-1248, 726-1202, 989-1047, 1046-1267, 1046-1490, 1064-1490, 1122-1463, 1317-
3902	1405, 1317-1412, 1317-1490, 1317-1843, 1317-1992, 1376-1992, 1390-1992, 1425-1945, 1626-2193, 1878-2160,
	1878-2162, 1878-2342, 1902-1981, 1927-2193, 2280-2549, 2291-2698, 2349-2629, 2349-2866, 2431-2955, 2462-
	2703, 2462-2747, 2474-3114, 2826-3536, 2911-3141, 2911-3528, 3033-3238, 3090-3695, 3113-3757, 3172-3456,
	3172-3460, 3191-3580, 3249-3752, 3253-3544, 3267-3765, 3276-3520, 3310-3761, 3402-3762, 3509-3741, 3616-
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Polynucleotide	Sequence Fragments
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Incyte ID/ Sequence Length	
60/7506047CB1/	1-482, 1-2993, 176-552, 176-659, 176-672, 176-691, 176-719, 176-730, 176-769, 176-771, 176-876, 196-905, 473-
2993	867, 473-1026, 474-752, 474-1026, 497-989, 498-956, 499-889, 536-983, 560-1016, 567-1140, 584-1159, 596-1056,
	648-1159, 686-1332, 688-1159, 693-1173, 720-1165, 721-1339, 730-1270, 790-1339, 823-1436, 852-1395, 864-
	1339, 871-1339, 899-1340, 917-1339, 917-1340, 1023-1635, 1060-1575, 1113-1571, 1118-1635, 1199-1804, 1222-
	1632, 1339-1633, 1417-1633, 1631-2180, 1848-2215, 2005-2591, 2103-2805, 2369-2993, 2443-2717, 2534-2974,
	2582-2967, 2771-2942
61/7505849CB1/	1-252, 2-315, 2-334, 4-277, 8-411, 8-660, 9-272, 10-131, 13-131, 15-288, 15-453, 15-467, 15-517, 16-286, 18-256,
619	19-281, 19-306, 19-590, 23-533, 24-311, 25-597, 26-445, 26-622, 27-268, 28-408, 29-281, 33-624, 35-256, 37-232,
	130-679, 131-368, 132-363, 152-621, 157-428, 158-574, 160-430, 171-659, 172-431, 175-443, 175-477, 175-483,
	179-432, 181-427, 186-474, 197-375, 214-441, 214-447, 223-467, 227-313, 235-679, 243-664, 250-677, 250-679,
	252-657, 258-496, 265-558, 278-659, 282-516, 294-523, 297-558, 300-561, 302-518, 308-563, 315-635, 334-653,
	336-679, 342-631, 361-614, 363-491, 363-679, 376-656, 381-585, 394-610, 401-655, 404-651, 404-674, 422-599,
	449-661
62/7505972CB1/	1-283, 1-285, 1-294, 1-300, 1-304, 2-272, 2-283, 2-305, 3-273, 3-311, 4-219, 4-282, 4-283, 4-771, 5-223, 5-239, 5-
918	251, 5-259, 5-283, 5-287, 6-205, 6-285, 6-300, 6-301, 6-304, 6-309, 7-178, 7-228, 7-238, 7-244, 7-252, 7-277, 7-282,
_	7-283, 7-284, 7-285, 7-289, 7-290, 7-293, 7-394, 7-305, 7-306, 7-308, 7-310, 7-311, 7-511, 7-590, 7-606, 7-
	628, 8-254, 8-307, 8-311, 9-288, 9-300, 10-95, 10-134, 10-179, 10-219, 10-249, 10-250, 10-261, 10-269, 10-278, 10-
	280, 10-282, 10-283, 10-285, 10-286, 10-287, 10-288, 10-289, 10-290, 10-291, 10-293, 10-294, 10-296, 10-297, 10-
	298, 10-300, 10-302, 10-308, 10-309, 10-310, 10-311, 11-105, 11-222, 11-251, 11-257, 11-260, 11-261, 11-262, 11-
	271, 11-277, 11-292, 11-294, 11-297, 11-301, 11-310, 11-313, 11-643, 12-184, 12-206, 12-219, 12-228, 12-231, 12-
	238, 12-241, 12-245, 12-250, 12-253, 12-256, 12-259, 12-262, 12-266, 12-272, 12-275, 12-278, 12-280, 12-281, 12-
	282, 12-283, 12-284, 12-285, 12-288, 12-289, 12-294, 12-295, 12-296, 12-297, 12-298, 12-299, 12-300, 12-302, 12-
	307, 12-311, 13-101, 13-209, 13-264, 13-268, 13-269, 13-284, 13-298, 13-308, 13-311, 14-219, 14-254, 14-281, 14-
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Polynucleotide	Sequence Fragments
SEQ ID NO:/	
Incyte ID/ Sequence	
Length	
62	16-270, 16-280, 16-284, 16-291, 16-301, 16-307, 16-311, 17-223, 17-253, 17-266, 17-297, 17-311, 18-209, 19-269,
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	585, 307-804, 308-544, 311-587, 314-615, 315-558, 315-572, 318-730, 319-417, 334-582, 337-604, 338-543, 340-
	606, 340-620, 341-605, 342-621, 344-563, 344-575, 344-633, 347-593, 347-625, 348-605, 350-591, 351-635, 356-
	599, 357-514, 357-620, 358-583, 358-590, 358-614, 358-637, 359-813, 360-604, 360-613, 363-625, 363-626, 364-
	647, 364-699, 370-643, 373-651, 376-628, 381-634, 383-666, 387-662, 391-654, 391-711, 392-671, 392-672, 392-
	674, 392-680, 399-679, 400-557, 400-649, 400-665, 403-664, 406-632, 406-653, 413-706, 413-804, 414-711, 416-
	680, 419-670, 419-683, 423-624, 423-644, 423-645, 423-676, 431-666, 431-709, 431-726, 432-690,
	432-712, 436-684, 436-700, 436-727, 441-706, 442-738, 444-918, 445-708, 445-727, 446-672, 450-708, 451-736,
	453-655, 456-649, 464-666, 466-754, 468-703, 468-716, 474-734, 475-671, 478-746, 482-752, 483-739, 485-728,
	488-671, 488-734, 488-750, 488-752, 488-757, 489-758, 490-699, 492-730, 492-745, 495-702, 496-754, 496-759,
	500-736, 500-757, 502-742, 507-707, 508-739, 510-733, 510-744, 520-685, 520-723, 520-743, 520-753, 520-757,
	549-757, 552-806, 559-735, 559-755, 563-718, 567-806, 575-733, 587-759, 629-759
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114/7507695CB1/	1-628, 1-2617, 18-578, 42-498, 42-590, 70-717, 206-1862, 337-713, 854-1063, 874-1562, 875-1056, 914-1031, 946-
2852	1269, 950-1029, 958-1646, 1198-1521, 1201-1280, 1202-1383, 1211-1392, 1274-1483, 1538-1617, 1538-1719, 1586-
	1703, 1621-1700, 1748-2046, 1748-2105, 1748-2107, 1748-2242, 1774-2319, 1919-2453, 1923-2541, 1963-2329,
	1984-2528, 2000-2520, 2053-2556, 2059-2529, 2079-2528, 2083-2608, 2104-2577, 2149-2614, 2179-2619, 2180-
	2617, 2199-2616, 2270-2616, 2280-2622, 2325-2852

Table 5

ary																														
Representative Library	MPHGNOT03	FTUBTUE01	KIDNFET02	OVARTDT01	BRSTNOT23	DRGTNOT01	EPIPUNA01	LUNGTUT03	UTREDIT07	ADRETUT01	BRSTNOT04	LIVRTUE01	BRADDIR01	SKINBITOI	BRSTNOT24	TESTNOF01	EYERNON01	TMLR2DT01	THYMNOR02	OVARNON03	CONNTUTOI	EPIPUNA01	FIBPFEN06	PROSTUT12	TLYMNOT03	FIBRTXS07	KIDETXF04	BRAIFEF01	BMARTXE01	RRAIFFIM
Incyte Project ID:	7506140CB1	1889415CB1	7506047CB1	7505849CB1	7505972CB1	7505991CB1	7506003CB1	6483977CB1	6301777CB1	7505976CB1	7506016CB1	7506086CB1	4176657CB1	7506056CB1	7506185CB1	8096611CB1	8174603CB1	3101042CB1	4972035CB1	7506265CB1	7506304CB1	7506198CB1	1381261CB1	6803876CB1	7506281CB1	7506175CB1	7506303CB1	7353336CB1	3001652CB1	1689128CR1
Polynucleotide SEQ Incyte Project ID:	ID NO:	59	95	19	62	63	8	65	99	29	89	69	70	71	72	73	74	75	76	77	78	62	80	81	82	83	84	85	86	87

Table 5

Polynucleotide SEQ ID NO: 88	Incyte Project ID: 2362969CB1 4753527CB1	Polynucleotide SEQ Incyte Project ID: Representative Library
	0928088CB1 7506388CB1 7376372CB1	BRAFNONUZ TESTNOC01 FTUBTUR01
	2754344CB1 8268822CB1	LNODNON02 EPIPNOT01
	1814553CB1 71217830CB1	THP INOTO I BRSTTUTO2
	7506252CB1 2270608CB1	PROSTUS19 EPIMNON05
	368741CB1	SYNORAT01
	7506253CB1	MPHGNOT03
	7506353CB1	THPITXT04
	7506335CB1	UTREDITO?
	5546982CB1	TESTNOC01
	7507432CB1	KIDETXJ01
	5639578CB1	UTRSTMR01
	7509080CB1	THYMFET03
	7505899CB1	PANCNOT04
	7505904CB1	PKINDNV32
	7509224CB1	SINTNOR01
	7505922CB1	NEUTLPT01
	7507695CB1	DRGCNOT01

Library	Vector	Library Description
TUTOI	PSPORT	Library was constructed using RNA isolated from right adrenal tumor tissue removed from a 50-year-old Turkish male during aunilateral adrenalectomy. Pathology indicated a metastatic renal cell carcinoma that formed a circumscribed, spongy, hemorrhagic nodule situated in the region of the medulla. The patient presented with corticoadrenal insufficiency, incisional hemia, and non-alcoholic steato hepatitis. Patient history included renal cell carcinoma. Family history included liver cancer.
BMARTXE01	pINCY	This 5' biased random primed library was constructed using RNA isolated from treated SH-SY5Y cells derived from a metastatic bone marrow neuroblastoma, removed from a 4-year-old Caucasian female (Schering AG). The medium was MEM/HAM'S F12 with 10% fetal calf serum. After reaching about 80% confluency cells were treated with 6-Hydroxydopamine (6-OHDA) at 100 microM for 8 hours.
BRADDIR01	pINCY	Library was constructed using RNA isolated from diseased choroid plexus tissue of the lateral ventricle, removed from the brain of a 57-year-old Caucasian male, who died from a cerebrovascular accident.
BRAFNON02	pINCY	This normalized frontal cortex tissue library was constructed from 10.6 million independent clones from a frontal cortex tissue library. Starting RNA was made from superior frontal cortex tissue removed from a 35-year-old Caucasian male who died from cardiac failure. Pathology indicated moderate leptomeningeal fibrosis and multiple microinfarctions of the cerebral neocortex. Grossly, the brain regions examined and cranial nerves were unremarkable. No atherosclerosis of the major vessels was noted. Microscopically, the cerebral hemisphere revealed moderate fibrosis of the leptomeninges with focal calcifications. There was evidence of shrunken and slightly eosinophilic pyramidal neurons throughout the cerebral hemispheres. There were also multiple small microscopic areas of cavitation with surrounding gliosis scattered throughout the cerebral cortex. Patient history included dilated cardiomyopathy, congestive heart failure, cardiomegaly, and an enlarged spleen and liver. Patient medications included simethicone, Lasix, Digoxin, Colace, Zantac, captopril, and Vasotec.
		The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
BRAIFEF01	PCMV-ICIS	This full-length enriched library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus who was stillborn with a hypoplastic left heart at 23 weeks' gestation. Serologies were negative.
BRAIFET02	pINCY	Library was constructed using RNA isolated from brain tissue removed from a Caucasian male fetus, who was stillborn with a hypoplastic left heart at 23 weeks' gestation.

Table (

Library	Vector	Library Description
BRSTNOT03	PSPORT1	Library was constructed using RNA isolated from diseased breast tissue removed from a 54-year-old Caucasian female during a bilateral radical mastectomy. Pathology for the associated tumor tissue indicated residual invasive grade 3 mammary ductal adenocarcinoma. Patient history included kidney infection and condyloma acuminatum. Family history included benign hypertension, hyperlipidemia and a malignant neoplasm of the colon.
BRSTNOT04	PSPORT1	Library was constructed using RNA isolated from breast tissue removed from a 62-year-old East Indian female during a unilateral extended simple mastectomy. Pathology for the associated tumor tissue indicated an invasive grade 3 ductal carcinoma. Patient history included benign hypertension, hyperlipidemia, and hematuria. Family history included cerebrovascular and cardiovascular disease, hyperlipidemia, and liver cancer.
BRSTNOT23	pINCY	Library was constructed using RNA isolated from diseased breast tissue removed from a 35-year-old Caucasian female during a bilateral reduction mammoplasty. Pathology indicated nonproliferative fibrocystic disease. Family history included type II diabetes, atherosclerotic coronary artery disease, acute myocardial infarction, hyperlipidemia, and coronary artery bypass.
BRSTNOT24	pINCY	Library was constructed using RNA isolated from diseased breast tissue removed from a 46-year-old Caucasian female during bilateral subcutaneous mammectomy. Pathology indicated nonproliferative fibrocystic disease. Family history included breast cancer and cardiovascular disease.
BRSTTUT02	PSPORTI	Library was constructed using RNA isolated from breast tumor tissue removed from a 54-year-old Caucasian female during a bilateral radical mastectomy with reconstruction. Pathology indicated residual invasive grade 3 mammary ductal adenocarcinoma. The remaining breast parenchyma exhibited proliferative fibrocystic changes without atypia. One of 10 axillary lymph nodes had metastatic tumor as a microscopic intranodal focus. Patient history included kidney infection and condyloma acuminatum. Family history included benign hypertension, hyperlipidemia, and a malignant colon neoplasm.
CONNTUTO	pINCY	Library was constructed using RNA isolated from a soft tissue tumor removed from the clival area of the skull of a 30-year-old Caucasian female. Pathology indicated chondroid chordoma with neoplastic cells reactive for keratin.

Library	Vector	Library Description
OTO		Library was constructed using RNA isolated from dorsal root ganglion tissue removed from the cervical spine of a 32-year-
		old Caucasian male who died from acute pulmonary edema and bronchopneumonia, bilateral pleural and pericardial effusions, and malignant lymphoma (natural killer cell type). Patient history included probable cytomegalovirus infection,
		hepatic congestion and steatosis, splenomegaly, hemorrhagic cystitis, thyroid hemorrhage, and Bell's palsy. Surgeries
		included radiation therapy.
EPIMNON05	pincy	This normalized mammary epithelial cell tissue library was constructed from 3.28 million independent clones from an epithelial cell tissue library. Starting RNA was made from untreated mammary epithelial cell tissue removed from a 21-year-old female. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 -hours/round) reannealing hybridization was used.
EPIPNOT01	pINCY	Library was constructed using RNA isolated from prostatic epithelial cells removed from a 17-year-old Hispanic male.
EPIPUNA01	PSPORTI	Library was constructed using RNA isolated from untreated prostatic epithelial cell tissue removed from a 17-year-old Hispanic male. Serologies were negative.
EYERNONOI	PSPORT1	This normalized pooled retina tissue library was constructed from independent clones from a pooled retina tissue library. Starting RNA was made from pooled retina tissue removed from 34 male and female donors, aged 9 to 80-years-old. The library was normalized in one round using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
FIBPFEN06	pINCY	The normalized prostate stromal fibroblast tissue libraries were constructed from 1.56 million independent clones from a prostate fibroblast library. Starting RNA was made from fibroblasts of prostate stroma removed from a male fetus, who died after 26 weeks' gestation. The libraries were normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48-hours/round)reannealing hybridization was used. The library was then linearized and recircularized to select for insert containing clones as follows: plasmid DNA was prepped from approximately 1 million clones from the normalized prostate stromal fibroblast tissue libraries following soft agar transformation.

Table (

Library	Vector	Library Description
2007	pINCY	This subtracted library was constructed using 1.3 million clones from a dermal fibroblast library and was subjected to two rounds of subtraction hybridization with 2.8 million clones from an untreated dermal fibroblast tissue library. The starting library for subtraction was constructed using RNA isolated from treated dermal fibroblast tissue removed from the breast of a 31-year-old Caucasian female. The cells were treated with 9CIS retinoic acid. The hybridization probe for subtraction was derived from a similarly constructed library from RNA isolated from untreated dermal fibroblast tissue from the same donor. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991) 19:1954 and Bonaldo, et al., Genome Research (1996) 6:791.
FTUBTUE01	pINCY	This 5' biased random primed library was constructed using RNA isolated from right fallopian tube tumor tissue removed from an 85-year-old Caucasian female during bilateral salpingo-oophorectomy and hysterectomy. Pathology indicated poorly differentiated mixed endometrioid (80%) and serous (20%) adenocarcinoma of the right fallopian tube, which was confined to the mucosa without mural involvement. Endometrioid carcinoma in situ was also present. Pathology for the associated uterus tumor indicated focal endometrioid adenocarcinoma in situ and moderately differentiated invasive adenocarcinoma arising in an endometrial polyp. A metastatic endometrioid and serous adenocarcinoma was present in the cul-de-sac tumor. The patient presented with a pelvic mass and ascites. Patient history included medullary carcinoma of the thyroid and myocardial infarction. Patient medications included Nitro-Dur, Lescol, Lasix and Cardizem.
FTUBTUR01	PCDNA2.1	This random primed library was constructed using RNA isolated from fallopian tube tumor tissue removed from an 85-year old Caucasian female during bilateral salpingo-oophorectomy and hysterectomy. Pathology indicated poorly differentiated mixed endometrioid (80%) and serous (20%) adenocarcinoma, which was confined to the mucosa without mural involvement. Endometrioid carcinoma in situ was also present. Pathology for the associated uterus tumor indicated focal endometrioid adenocarcinoma in situ and moderately differentiated invasive adenocarcinoma arising in an endometrial polyp. Metastatic endometrioid and serous adenocarcinoma was present at the cul-de-sac tumor. Patient history included medullary carcinoma of the thyroid and myocardial infarction.
KIDETXF04	PCMV-ICIS	Library was constructed using RNA isolated from a treated, transformed embryonal cell line (293-EBNA) derived from kidney epithelial tissue. The cells were treated with 5-aza-2*-deoxycytidine (5AZA) for 72 hours and Trichostatin A for 24 hours and transformed with adenovirus 5 DNA.

Table (

Library	Vector	Library Description
KIDETXJ01	:	This random primed 5' cap isolated library was constructed using RNA isolated from a treated, transformed embryonal cell line (293-EBNA) derived from kidney epithelial tissue (Invitrogen). The cells were treated with 5-aza-2'-deoxycytidine (5AZA) for 72 hours and Trichostatin A for 24 hours and transformed with adenovirus 5 DNA.
KIDNFET02	pINCY	Library was constructed using RNA isolated from kidney tissue removed from a Caucasian male fetus, who was stillborn with a hypoplastic left heart and died at 23 weeks' gestation.
LIVRTUE01	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from liver tumor tissue removed from a 72-year-old Caucasian male during partial hepatectomy. Pathology indicated metastatic grade 2 (of 4) neuroendocrine carcinoma forming a mass. The patient presented with metastatic liver cancer. Patient history included benign hypertension, type I diabetes, prostatic hyperplasia, prostate cancer, alcohol abuse in remission, and tobacco abuse in remission. Previous surgeries included destruction of a pancreatic lesion, closed prostatic biopsy, transurethral prostatectomy, removal of bilateral testes and total splenectomy. Patient medications included Eulexin, Hytrin, Proscar, Ecotrin, and insulin. Family history included atherosclerotic coronary artery disease and acute myocardial infarction in the mother; atherosclerotic coronary artery disease and type II diabetes in the father.
LNODNON02	pINCY	This normalized lymph node tissue library was constructed from .56 million independent clones from a lymph node tissue library. Starting RNA was made from lymph node tissue removed from a 16-month-old Caucasian male who died from head trauma. Serologies were negative. Patient history included bronchitis. Patient medications included Dopamine, Dobutamine, Vancomycin, Vasopressin, Proventil, and Atarax. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228-9932 and Bonaldo et al., Genome Research 6 (1996):791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
LUNGTUT03	PSPORT1	Library was constructed using RNA isolated from lung tumor tissue removed from the left lower lobe of a 69-year-old Caucasian male during segmental lung resection. Pathology indicated residual grade 3 invasive squamous cell carcinoma. Patient history included acute myocardial infarction, prostatic hyperplasia, malignant skin neoplasm, and tobacco use.

Library	Vector	Library Description
XT04	pINCY	Library was constructed using RNA isolated from treated, derived dendritic cells from umbilical cord blood CD34+ precursor cells removed from a male. The cells were derived with granulocyte/macrophage colony stimulating factor (GM-CSF), tumor necrosis factor alpha (TNF alpha), and stem cell factor (SCF), then treated with phorbol myristate acetate (PMA), and Ionomycin. The GM-CSF was added at time 0 at 100 ng/ml, the TNF alpha was added at time 0 at 25 ng/ml. The PMA and Ionomycin were added at 13 days for five hours. Incubation time was 13 days.
MPHGNOT03	PBLUESCRIPT I	Library was constructed using RNA isolated from plastic adherent mononuclear cells isolated from buffy coat units obtained from unrelated male and female donors.
NEUTLPTOI	PBLUESCRIPT L	Library was constructed using RNA isolated from peripheral blood granulocytes collected by density gradient centrifugation through Ficoll-Hypaque. The cells were isolated from buffy coat units obtained from unrelated male and female donors. Cells were cultured in 100 ng/ml E. coli LPS for 30 minutes, lysed in GuSCN, and spun through CsCl to obtain RNA for library construction.
OVARNON03	pINCY	This normalized ovarian tissue library was constructed from 5 million independent clones from an ovary library. Starting RNA was made from ovarian tissue removed from a 36-year-old Caucasian female during total abdominal hysterectomy, bilateral salpingo-oophorectomy, soft tissue excision, and an incidental appendectomy. Pathology for the associated tumor tissue indicated one intramural and one subserosal leiomyomata of the myometrium. The endometrium was proliferative phase. Patient history included deficiency anemia, calculus of the kidney, and a kidney anomaly. Family history included hyperlipidemia, acute myocardial infarction, atherosclerotic coronary artery disease, type II diabetes, and chronic liver disease. The library was normalized in two rounds using conditions adapted from Soares et al., PNAS (1994) 91:9228 and Bonaldo et al., Genome Research (1996) 6:791, except that a significantly longer (48 hours/round) reannealing hybridization was used.
OVARTDT01	piNCY	Library was constructed using RNA isolated from right ovary tissue removed from a 47-year-old Caucasian female during a total abdominal hysterectomy with bilateral salpingo-oophorectomy. Pathology for the associated tumor tissue indicated two intramural leiomyomas. The endometrium was in the secretory phase. The patient presented with abnormal blood chemistry. Patient history included infertility, abnormal blood chemistry, abnormal heart sounds, andextrinsic asthma. Family history included benign hypertension, atheroscleroticcoronary artery disease, an aortic valve disorder, acute myocardial infarction, cerebrovascular disease, and pancreatic cancer.
PANCNOT04	PSPORTI	Library was constructed using RNA isolated from the pancreatic tissue of a 5-year-old Caucasian male who died in a motor vehicle accident.

Library	Vector	Library Description
PKINDNV32	PCR2-TOPOTA	PCR2-TOPOTA Library was constructed using pooled cDNA from different donors. cDNA was generated using mRNA isolated from pooled skeletal muscle tissue removed from ten 21 to 57-year-old Caucasian male and female donors who died from sudden death; from pooled thymus tissue removed from nine 18 to 32-year-old Caucasian male and female donors who died from sudden death; from pooled liver tissue removed from 32 Caucasian male and female fetuses who died at 18-24 weeks gestation due to spontaneous abortion; from kidney tissue removed from 59 Caucasian male and female fetuses who died at 20-33 weeks gestation due to spontaneous abortion; and from brain tissue removed from a Caucasian male fetus who died at 23 weeks gestation due to fetal demise.
PROSTMY01	pINCY	This large size-fractionated cDNA and normalized library was constructed using RNA isolated from diseased prostate tissue removed from a 55-year-old Caucasian male during closed prostatic biopsy, radical prostatectomy, and regional lymph node excision. Pathology indicated adenofibromatous hyperplasia. Pathology for the matched tumor tissue indicated adenocarcinoma Gleason grade 4 forming a predominant mass involving the left side peripherally with extension into the right posterior superior region. The tumor invaded the capsule and perforated the capsule to involve periprostatic tissue in the left posterior superior region. The left inferior posterior and left superior posterior surgical margins are positive. One left pelvic lymph node is metastatically involved. Patient history included calculus of the kidney. Family history included lung cancer and breast cancer. The size-selected library was normalized in 1 round using conditions adapted from Soares et al., PNAS (1994) 91:9228-9232 and Bonaldo et al., Genome Research (1996) 6:791.
PROSTUS19	pINCY	This subtracted prostate tumor tissue library was constructed using 2.36 million clones from a prostate tumor library and was subjected to two rounds of subtraction hybridization with 2.36 million clones from a prostate epithelium library. The starting library for subtraction was constructed using RNA isolated from prostate tumor tissue removed from a 59-year-old Caucasian male during a radical prostatectomy with regional lymph node excision. Pathology indicated adenocarcinoma (Gleason grade 3+3) involving the prostate peripherally with invasion of the capsule. Adenofibromatous hyperplasia was present. The patient presented with elevated prostate-specific antigen (PSA). Patient history included diverticulitis of colon, asbestosis, and thrombophlebitis. Family history included benign hypertension, multiple myeloma, hyperlipidemia, and rheumatoid arthritis. Subtractive hybridization conditions were based on the methodologies of Swaroop et al., NAR (1991) 19:1954 and Bonaldo, et al. Genome Research (1996) 6:791.

Library	Vector	Library Description
PROSTUT12	pINCY	Library was constructed using RNA isolated from prostate tumor tissue removed from a 65-year-old Caucasian male during a radical prostatectomy. Pathology indicated an adenocarcinoma (Gleason grade 2+2). Adenofibromatous hyperplasia was also present. The patient presented with elevated prostate specific antigen (PSA).
SINTNOROI	PCDNA2.1	This random primed library was constructed using RNA isolated from small intestine tissue removed from a 31-year-old Caucasian female during Roux-en-Y gastric bypass. Patient history included clinical obesity.
SKINBITOI	pINCY	Library was constructed using RNA isolated from diseased skin tissue of the left lower leg. Patient history included erythema nodosum of the left lower leg.
SYNORATO	PSPORTI	Library was constructed using RNA isolated from synovial membrane tissue removed from the elbow of a 51-year-old Asian female with rheumatoid arthritis.
TESTNOC01	PBLUESCRIPT	PBLUESCRIPT This large size fractionated library was constructed using RNA isolated from testicular tissue removed from a pool of eleven, 10 to 61-year-old Caucasian males.
TESTNOF01	PSPORTI	This 5' cap isolated full-length library was constructed using RNA isolated from testis tissue removed from a 26-year-old Caucasian male who died from head trauma due to a motor vehicle accident. Serologies were negative. Patient history included a hernia at birth, tobacco use (1 1/2 ppd), marijuana use, and daily alcohol use (beer and hard liquor).
TESTTUE02	PCDNA2.1	This 5' biased random primed library was constructed using RNA isolated from testicular tumor removed from a 31-year-old Caucasian male during unilateral orchiectomy. Pathology indicated embryonal carcinoma forming a largely necrotic mass involving the entire testicle. Rare foci of residual testicle showed intralobular germ cell neoplasia and tumor was identified at the spermatic cord margin. The patient presented with backache. Patient history included tobacco use. Previous surgeries included a needle biopsy of testis. Patient medications included Colace and antacids.
THPINOTOI	pINCY	Library was constructed using RNA isolated from untreated THP-1 cells. THP-1 is a human promonocyte line derived from the peripheral blood of a 1-year-old Caucasian male with acute monocytic leukemia (ref: Int. J. Cancer (1980) 26:171).
THPITXT04	pINCY	Library was constructed using RNA isolated from stimulated THP-1 cells. THP-1 is a human promonocyte line derived from the peripheral blood of a 1-year-old male (Abbott Sample) with acute monocytic leukemia (Int. J. Cancer 26 (1980):171).
THYMFET03	pINCY	Library was constructed using RNA isolated from thymus tissue removed from a Caucasian male fetus.

Table (

Library	Vector	Library Description
THYMNOR02 pINCY	pINCY	The library was constructed using RNA isolated from thymus tissue removed from a 2-year-old Caucasian female during a
_		thymectomy and patch closure of left atrioventricular fistula. Pathology indicated there was no gross abnormality of the
		thymus. The patient presented with congenital heart abnormalities. Patient history included double inlet left ventricle and a
		rudimentary right ventricle, pulmonary hypertension, cyanosis, subaortic stenosis, seizures, and a fracture of the skull base.
		Family history included reflux neuropathy.
TLYMNOT03 pINCY	pINCY	Library was constructed using RNA isolated from nonactivated Th1 cells. These cells were differentiated from umbilical cord CD4 T cells with IL-12 and B7-transfected COS cells.
TMLR2DT01	PBLUESCRIPT	Library was constructed using RNA isolated from non-adherent peripheral blood mononuclear cells. The blood was
	_	obtained from unrelated male and female donors. Cells from each donor were purified on Ficoll Hypaque, then co-cultured
		for 24 hours in medium containing normal human serum at a cell density of 2million cells/ml.
UTREDITO7	pINCY	Library was constructed using RNA isolated from diseased endometrial tissue removed from a female during endometrial
		biopsy. Pathology indicated in phase endometrium with missing beta 3, Type II defects.
UTRSTMR01	pINCY	Library was constructed using RNA isolated from uterine myometrial tissue removed from a 41-year-old Caucasian female
_		during a vaginal hysterectomy. The endometrium was secretory and contained fragments of endometrial polyps. Pathology
		for associated tumor tissue indicated uterine leiomyoma. Patient history included ventral hemia and a benign ovarian
		neoplasm.

Program	Description	Reference	Parameter Threshold
ABIFACTURA	A program that removes vector sequences and masks Applied Biosystems, Foster City, CA ambiguous bases in nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
ABI/PARACEL FDF	A Fast Data Finder useful in comparing and annotating amino acid or nucleic acid sequences.	Applied Biosystems, Foster City, CA; Paracel Inc., Pasadena, CA.	Mismatch <50%
ABI AutoAssembler	A program that assembles nucleic acid sequences.	Applied Biosystems, Foster City, CA.	
BLAST	A Basic Local Alignment Search Tool useful in sequence similarity search for amino acid and nucleic acid sequences. BLAST includes five functions: blastp, blastn, blastn, tblastn, and tblastx.	al Alignment Search Tool useful in Altschul, S.F. et al. (1990) J. Mol. Biol. illarity search for amino acid and nucleic 215:403-410; Altschul, S.F. et al. (1997) es. BLAST includes five functions: Nucleic Acids Res. 25:3389-3402. Nucleic Acids Res. 25:3389-3402.	ESTs: Probability value = 1.0E-8 or less; Full Length sequences: Probability value = 1.0E-10 or less
FASTA	A Pearson and Lipman algorithm that searches for similarity between a query sequence and a group of sequences of the same type. FASTA comprises as least five functions: fasta, tfasta, fastx, tfastx, and ssearch.	Pearson, W.R. and D.J. Lipman (1988) Proc. ESTs: fasta E value = 1.06E-6; Natl. Acad Sci. USA 85:2444-2448; Pearson, Assembled ESTs: fasta Identity W.R. (1990) Methods Enzymol. 183:63-98; = 95% or greater and Match and Smith, T.F. and M.S. Waterman (1981) length = 200 bases or greater; fastx Adv. Appl. Math. 2:482-489. Full Length sequences: fastx score = 100 or greater	ESTs: fasta E value = 1.06E-6; Assembled ESTs: fasta Identity = 95% or greater and Match length = 200 bases or greater; fastx E value = 1.0E-8 or less; Full Length sequences: fastx score = 100 or greater
BLIMPS	A BLocks IMProved Searcher that matches a sequence against those in BLOCKS, PRINTS, DOMO, PRODOM, and PFAM databases to search for gene families, sequence homology, and structural fingerprint regions. Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff I.G. and S. Henikoff (1996) Methods I.G. and S. Henikoff (1991) I.G. and J.G. Henikoff (1991)	Henikoff, S. and J.G. Henikoff (1991) Nucleic Acids Res. 19:6565-6572; Henikoff, less J.G. and S. Henikoff (1996) Methods Enzymol. 266:88-105; and Attwood, T.K. et al. (1997) J. Chem. Inf. Comput. Sci. 37:417-	Probability value = 1.0E-3 or less

Drogen			Dogumeter Threshold
rrogram		Reletence	rainiciei Illication
HMMER	An algorithm for searching a query sequence against	Krogh, A. et al. (1994) J. Mol. Biol.	PFAM, INCY, SMART or
		235:1501-1531; Sonnhammer, E.L.L. et al.	TIGRFAM hits: Probability
	÷	(1988) Nucleic Acids Res. 26:320-322;	value = 1.0E-3 or less; Signal
	INCY, SMART and TIGREAM.	Durbin, R. et al. (1998) Our World View, in	peptide hits: Score = 0 or greater
		a Nutshell, Cambridge Univ. Press, pp. 1-	
		350.	
ProfileScan	An algorithm that searches for structural and	Gribskov, M. et al. (1988) CABIOS 4:61-66; Normalized quality score ≥ GCG	Normalized quality score ≥ GCG
	sequence motifs in protein sequences that match	Gribskov, M. et al. (1989) Methods	specified "HIGH" value for that
	sequence patterns defined in Prosite.	Enzymol. 183:146-159; Bairoch, A. et al.	particular Prosite motif.
		(1997) Nucleic Acids Res. 25:217-221.	Generally, score = 1.4-2.1.
Phred	A base-calling algorithm that examines automated	Ewing, B. et al. (1998) Genome Res. 8:175-	
	sequencer traces with high sensitivity and probability.	aces with high sensitivity and probability. 185; Ewing, B. and P. Green (1998) Genome	
		Res. 8:186-194.	
Phrap	A Phils Revised Assembly Program including	Smith, T.F. and M.S. Waterman (1981) Adv. Score = 120 or greater; Match	Score = 120 or greater; Match
•	SWAT and CrossMatch, programs based on efficient Appl. Math. 2:482-489; Smith, T.F. and	Appl. Math. 2:482-489; Smith, T.F. and	length = 56 or greater
	implementation of the Smith-Waterman algorithm,	M.S. Waterman (1981) J. Mol. Biol. 147:195-	
	useful in searching sequence homology and	197; and Green, P., University of	
	assembling DNA sequences.	Washington, Seattle, WA.	
Consed	A graphical tool for viewing and editing Phrap	Gordon, D. et al. (1998) Genome Res. 8:195-	
	assemblies.	202.	
SPScan	A weight matrix analysis program that scans protein	Nielson, H. et al. (1997) Protein Engineering Score = 3.5 or greater	Score = 3.5 or greater
	sequences for the presence of secretory signal	10:1-6; Claverie, J.M. and S. Audic (1997)	
	peptides.	CABIOS 12:431-439.	
TMAP	A program that uses weight matrices to delineate	Persson, B. and P. Argos (1994) J. Mol. Biol.	
	transmembrane segments on protein sequences and	237:182-192; Persson, B. and P. Argos	
	determine orientation.	(1996) Protein Sci. 5:363-371.	

Table '

Program	Description	Reference	Parameter Threshold
CER .		hat uses a hidden Markov model (HMM) Sonnhammer, E.L. et al. (1998) Proc. Sixth transmembrane segments on protein Intl. Conf. On Intelligent Systems for Mol. Biol., Glasgow et al., eds., The Am. Assoc. for Artificial Intelligence (AAAI) Press, Menlo Park, CA, and MIT Press, Cambridge, MA, pp. 175-182.	
Motifs	A program that searches amino acid sequences for patterns that matched those defined in Prosite.	Bairoch, A. et al. (1997) Nucleic Acids Res. 25:217-221; Wisconsin Package Program Manual, version 9, page M51-59, Genetics Computer Group, Madison, WI.	

															_														
Hispanic Allele 1 frequency		n/a	n/a	n/a	0.80	n/a	0.23	n/d	n/đ	n/a																			
Asian Allele 1 frequency	•	n/a	n/a	n/a	0.84	n/a	0.10	n/d	n/d	n/a	n/a	n/a	n/a	n/a	п/а	п/а	n/a	n/a	r/a										
African Allele 1 frequency		n/a	n/a	n/a	98.0	n/a	0.08	0.99	p/u	n/a																			
Caucasian Allele 1 frequency	,	n/a	n/a	n/a	0.81	n/a	n/a	n/a	n/a	n/d	n/a	n/a	n/a	p/u	0.33	0.70	0.27	p/u	p/u	n/a	n/a	p/u	p/u	p/u	n/a	n/a	n/d	n/a	p/u
Amino Acid		noncoding	141	noncoding	noncoding	noncoding	noncoding	R61	noncoding	noncoding	N277	noncoding	N216	W264	P65	A429	L562	T48	H508	L979	S83	P62	D67	noncoding	Q172	S597	F500	noncoding	K24
Allele 2		G	၁	Α	Т	g	υ	Т	g	၁	۲	Т	ပ	V	٧	V	ပ	T	T	Т	g	Y	¥	Т	Т	Т	C	C	g
Allele 1		Т	Т	g	၁	Α	H	၁	၁	g	ပ	ပ	4	ຽ	g	ŋ	۲	ပ	ပ	၁	A	ပ	ပ	Ŋ	၁	g	Τ	Т	C
EST Allele		T	Т	G	Т	A	Т	Т	ပ	Ð	T	Т	A	ŋ	A	ŋ	Т	ပ	ပ	Ţ	9	ပ	ပ	Ŋ	С	g	H	Т	G
CB1 SNP		4007 !	182	108	5187	224	2873	287	1454	295	906	3838	1103	854	451	1855	1987	316	1687	2950	386	210	276	199	643	2097	1525	4778	181
EST	•	8	62	13	12	92	162	981	105	256	153	131	14	183	254	210	270	195	212	31	389	=	8	151	157	35	170	360	125
SNP ID		SNP00015185	SNP00020849	SNP00039835	SNP00012943	SNP00066548	SNP00107537	SNP00014990	SNP00009517	SNP00032569	SNP00049926	SNP00009056	SNP00047603	SNP00112423	SNP00024799	SNP00060994	SNP00000294	SNP00031965	SNP00105341	SNP00051840	SNP00129354	SNP00058616	SNP00029137	SNP00036352	SNP00029300	SNP00074077	SNP00044438	SNP00015247	SNP00047347
EST ID		2427525H1	007882H1	1294222H1	1002021H1	1210089HI	2686636H1	1856272H1	3937378H1	031320H1	6712119HI	1910984HI	119060169	3597648H1	4674120H1	2600624H1	2745172H1	3383569H1	1870195H1	2006992H1	7667604H1	1432222H1	1322352H1	1831292H1	3166112H1	5822041H1	3390140H1	6500745H1	6031045H1
OIA	•	4972035	7506265	7506304	7506198	6803876	7506281	7506175	7506303	7353336	3001652	1689128	2362969	4753527	7506388	7376372	2754344	8268822	1814553	7506252	368741	7506379	7506253	7506353	7506372	5546982	7507432	7509080	7505899
) Eg e g	<u>. </u>	9/	11	78	79	81	82	83	æ	85	98	87	88	68	91	33	93	8	95	6	001	101	102	103	104	901	101	601	110

Table 8

SEQ	PID	ESTID	SNPID	EST	CBI	EST	Allele	Allele	Amino Acid	Caucasian	African	Asian	Hispanic
£				SNP	SNP	Allele	-	7	SNP SNP Allele 1 2 Allele 1	Allele 1	Allele 1	Allele 1	Allele 1
Ö										frequency	frequency	frequency	frequency
Ξ	7505904	5836174H1	SNP00015294	241	896	S 896	CT	T	noncoding	n/a	n/a	n/a	n/a
112	7509224	2702219HI	SNP00037452	861	1509	A	g	4	E322	n/a	n/a	n/a	n/a
113	7505922	4184173HI	SNP00013088	126	301	ပ	ပ	T	L25	0.75	86.0	0.76	0.77
114	5692052	1274378H1	SNP00003360	117	2079	Ö	4	ŋ	noncoding	0.74	n/a	n/a	n/a

What is claimed is:

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- 1. An isolated polypeptide selected from the group consisting of:
- a) a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57,
- b) a polypeptide comprising a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:1-2, SEQ ID NO:13, SEQ ID NO:16, SEQ ID NO:18-20, SEQ ID NO:22-23, SEQ ID NO:31-32, SEQ ID NO:36-39, SEQ ID NO:41, SEQ ID NO:43, SEQ ID NO:46, SEQ ID NO:51, and SEQ ID NO:55,
- a polypeptide comprising a naturally occurring amino acid sequence at least 91% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:21, and SEQ ID NO:27,
- a polypeptide comprising a naturally occurring amino acid sequence at least 94% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:17, SEQ ID NO:20, and SEQ ID NO:42,
- e) a polypeptide comprising a naturally occurring amino acid sequence at least 95% identical to the amino acid sequence of SEQ ID NO:50,
- f) a polypeptide comprising a naturally occurring amino acid sequence at least 96% identical to the amino acid sequence of SEQ ID NO:19,
- g) a polypeptide comprising a naturally occurring amino acid sequence at least 98% identical to the amino acid sequence of SEQ ID NO:24,
- a polypeptide comprising a naturally occurring amino acid sequence at least 99% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:9, SEQ ID NO:33, SEQ ID NO:35, SEQ ID NO:52, and SEQ ID NO:57,
- i) a polypeptide consisting essentially of a naturally occurring amino acid sequence at least 90% identical to an amino acid sequence selected from the group consisting of SEQ ID NO:3-8, SEQ ID NO:10-12, SEQ ID NO:14-15, SEQ ID NO:24-27, SEQ ID NO:29, SEQ ID NO:34, SEQ ID NO:40, SEQ ID NO:42, SEQ ID NO:44-45, SEQ ID NO:47-49, SEQ ID NO:53-54, and SEQ ID NO:56,
- a biologically active fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, and

k) an immunogenic fragment of a polypeptide having an amino acid sequence selected from the group consisting of SEQ ID NO:1-57.

- An isolated polypeptide of claim 1 comprising an amino acid sequence selected from the
 group consisting of SEQ ID NO:1-57.
 - 3. An isolated polynucleotide encoding a polypeptide of claim 1.
 - 4. An isolated polynucleotide encoding a polypeptide of claim 2.

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- 5. An isolated polynucleotide of claim 4 comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114.
- A recombinant polynucleotide comprising a promoter sequence operably linked to a
 polynucleotide of claim 3.
 - 7. A cell transformed with a recombinant polynucleotide of claim 6.
 - 8. A transgenic organism comprising a recombinant polynucleotide of claim 6.

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- 9. A method of producing a polypeptide of claim 1, the method comprising:
- a) culturing a cell under conditions suitable for expression of the polypeptide, wherein said cell is transformed with a recombinant polynucleotide, and said recombinant polynucleotide comprises a promoter sequence operably linked to a polynucleotide encoding the polypeptide of claim 1, and

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- b) recovering the polypeptide so expressed.
- 10. A method of claim 9, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-57.

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11. An isolated antibody which specifically binds to a polypeptide of claim 1.

12. An isolated polynucleotide selected from the group consisting of:

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- a) a polynucleotide comprising a polynucleotide sequence selected from the group consisting of SEQ ID NO:58-114,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 90% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:58-67, SEQ ID NO:69-86, SEQ ID NO:88, SEQ ID NO:93, SEQ ID NO:95 96, SEQ ID NO:98, SEQ ID NO:100, SEQ ID NO:108, and SEQ ID NO:110-112,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 91% identical to the polynucleotide sequence of SEO ID NO:94,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 93% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:106, and SEQ ID NO:114,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 94% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:87, and SEQ ID NO:103,
- f) a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 96% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:89-90, SEQ ID NO:92, and SEQ ID NO:101,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 97% identical to the polynucleotide sequence of SEQ ID NO:99,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 98% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:68, SEQ ID NO:91, and SEQ ID NO:109,
- a polynucleotide comprising a naturally occurring polynucleotide sequence at least
 99% identical to a polynucleotide sequence selected from the group consisting of SEQ
 ID NO:102, and SEQ ID NO:107,
- j) a polynucleotide consisting essentially of a naturally occurring polynucleotide sequence at least 90% identical to a polynucleotide sequence selected from the group consisting of SEO ID NO:97, SEO ID NO:104-105, and SEO ID NO:113,
- k) a polynucleotide complementary to a polynucleotide of a),
 - 1) a polynucleotide complementary to a polynucleotide of b),
 - m) a polynucleotide complementary to a polynucleotide of c),

- n) a polynucleotide complementary to a polynucleotide of d),
- o) a polynucleotide complementary to a polynucleotide of e),
- p) a polynucleotide complementary to a polynucleotide of f),
- q) a polynucleotide complementary to a polynucleotide of g),
- r) a polynucleotide complementary to a polynucleotide of h),
- s) a polynucleotide complementary to a polynucleotide of i),
- t) a polynucleotide complementary to a polynucleotide of j), and
- u) an RNA equivalent of a)-t).

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- 13. An isolated polynucleotide comprising at least 60 contiguous nucleotides of a polynucleotide of claim 12.
 - 14. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
- 15 a) hybridizing the sample with a probe comprising at least 20 contiguous nucleotides comprising a sequence complementary to said target polynucleotide in the sample, and which probe specifically hybridizes to said target polynucleotide, under conditions whereby a hybridization complex is formed between said probe and said target polynucleotide or fragments thereof, and
- 20 b) detecting the presence or absence of said hybridization complex, and, optionally, if present, the amount thereof.
 - 15. A method of claim 14, wherein the probe comprises at least 60 contiguous nucleotides.
- 25 16. A method of detecting a target polynucleotide in a sample, said target polynucleotide having a sequence of a polynucleotide of claim 12, the method comprising:
 - amplifying said target polynucleotide or fragment thereof using polymerase chain reaction amplification, and
 - b) detecting the presence or absence of said amplified target polynucleotide or fragment thereof, and, optionally, if present, the amount thereof.
 - 17. A composition comprising a polypeptide of claim 1 and a pharmaceutically acceptable

excipient.

18. A composition of claim 17, wherein the polypeptide comprises an amino acid sequence selected from the group consisting of SEQ ID NO:1-57.

- 19. A method for treating a disease or condition associated with decreased expression of functional NAAP, comprising administering to a patient in need of such treatment the composition of claim 17.
- 20. A method of screening a compound for effectiveness as an agonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
 - b) detecting agonist activity in the sample.
- 15 21. A composition comprising an agonist compound identified by a method of claim 20 and a pharmaceutically acceptable excipient.
- 22. A method for treating a disease or condition associated with decreased expression of functional NAAP, comprising administering to a patient in need of such treatment a composition of
 claim 21.
 - 23. A method of screening a compound for effectiveness as an antagonist of a polypeptide of claim 1, the method comprising:
 - a) exposing a sample comprising a polypeptide of claim 1 to a compound, and
- 25 b) detecting antagonist activity in the sample.
 - 24. A composition comprising an antagonist compound identified by a method of claim 23 and a pharmaceutically acceptable excipient.
- 30 25. A method for treating a disease or condition associated with overexpression of functional NAAP, comprising administering to a patient in need of such treatment a composition of claim 24.

26. A method of screening for a compound that specifically binds to the polypeptide of claim 1, the method comprising:

- a) combining the polypeptide of claim 1 with at least one test compound under suitable conditions, and
- b) detecting binding of the polypeptide of claim 1 to the test compound, thereby identifying a compound that specifically binds to the polypeptide of claim 1.

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- 27. A method of screening for a compound that modulates the activity of the polypeptide of claim 1, the method comprising:
- a) combining the polypeptide of claim 1 with at least one test compound under conditions permissive for the activity of the polypeptide of claim 1,
 - b) assessing the activity of the polypeptide of claim 1 in the presence of the test compound, and
 - c) comparing the activity of the polypeptide of claim 1 in the presence of the test compound with the activity of the polypeptide of claim 1 in the absence of the test compound, wherein a change in the activity of the polypeptide of claim 1 in the presence of the test compound is indicative of a compound that modulates the activity of the polypeptide of claim 1.
- 28. A method of screening a compound for effectiveness in altering expression of a target polynucleotide, wherein said target polynucleotide comprises a sequence of claim 5, the method comprising:
 - exposing a sample comprising the target polynucleotide to a compound, under conditions suitable for the expression of the target polynucleotide,
 - b) detecting altered expression of the target polynucleotide, and
 - c) comparing the expression of the target polynucleotide in the presence of varying amounts of the compound and in the absence of the compound.
 - 29. A method of assessing toxicity of a test compound, the method comprising:
 - a) treating a biological sample containing nucleic acids with the test compound,
 - b) hybridizing the nucleic acids of the treated biological sample with a probe comprising at least 20 contiguous nucleotides of a polynucleotide of claim 12 under conditions

whereby a specific hybridization complex is formed between said probe and a target polynucleotide in the biological sample, said target polynucleotide comprising a polynucleotide sequence of a polynucleotide of claim 12 or fragment thereof,

- c) quantifying the amount of hybridization complex, and
- 5 d) comparing the amount of hybridization complex in the treated biological sample with the amount of hybridization complex in an untreated biological sample, wherein a difference in the amount of hybridization complex in the treated biological sample is indicative of toxicity of the test compound.
- 30. A method for a diagnostic test for a condition or disease associated with the expression of NAAP in a biological sample, the method comprising:
 - a) combining the biological sample with an antibody of claim 11, under conditions suitable for the antibody to bind the polypeptide and form an antibody:polypeptide complex, and
- b) detecting the complex, wherein the presence of the complex correlates with the presence of the polypeptide in the biological sample.
 - 31. The antibody of claim 11, wherein the antibody is:
 - a) a chimeric antibody,
 - b) a single chain antibody,
 - c) a Fab fragment,

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- d) a F(ab')₂ fragment, or
- e) a humanized antibody.
- 25 32. A composition comprising an antibody of claim 11 and an acceptable excipient.
 - 33. A method of diagnosing a condition or disease associated with the expression of NAAPin a subject, comprising administering to said subject an effective amount of the composition of claim32.

34. A composition of claim 32, further comprising a label.

35. A method of diagnosing a condition or disease associated with the expression of NAAP

in a subject, comprising administering to said subject an effective amount of the composition of claim 34.

- 36. A method of preparing a polyclonal antibody with the specificity of the antibody of claim
 11, the method comprising:
 - immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
 - b) isolating antibodies from the animal, and

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- c) screening the isolated antibodies with the polypeptide, thereby identifying a polyclonal antibody which specifically binds to a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57.
 - 37. A polyclonal antibody produced by a method of claim 36.
 - 38. A composition comprising the polyclonal antibody of claim 37 and a suitable carrier.
 - 39. A method of making a monoclonal antibody with the specificity of the antibody of claim 11, the method comprising:
 - a) immunizing an animal with a polypeptide consisting of an amino acid sequence selected from the group consisting of SEQ ID NO:1-57, or an immunogenic fragment thereof, under conditions to elicit an antibody response,
 - b) isolating antibody producing cells from the animal,
 - c) fusing the antibody producing cells with immortalized cells to form monoclonal antibody-producing hybridoma cells,
 - d) culturing the hybridoma cells, and
 - e) isolating from the culture monoclonal antibody which specifically binds to a
 polypeptide comprising an amino acid sequence selected from the group consisting of
 SEQ ID NO:1-57.
 - 40. A monoclonal antibody produced by a method of claim 39.
 - 41. A composition comprising the monoclonal antibody of claim 40 and a suitable carrier.

42. The antibody of claim 11, wherein the antibody is produced by screening a Fab expression library.

- 43. The antibody of claim 11, wherein the antibody is produced by screening a recombinant immunoglobulin library.
 - 44. A method of detecting a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57 in a sample, the method comprising:
 - a) incubating the antibody of claim 11 with the sample under conditions to allow specific binding of the antibody and the polypeptide, and
 - b) detecting specific binding, wherein specific binding indicates the presence of a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57 in the sample.
- 45. A method of purifying a polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57 from a sample, the method comprising:
 - incubating the antibody of claim 11 with the sample under conditions to allow specific binding of the antibody and the polypeptide, and
 - separating the antibody from the sample and obtaining the purified polypeptide comprising an amino acid sequence selected from the group consisting of SEQ ID NO:1-57.
 - 46. A microarray wherein at least one element of the microarray is a polynucleotide of claim 13.
 - 47. A method of generating an expression profile of a sample which contains polynucleotides, the method comprising:
 - a) labeling the polynucleotides of the sample,

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- b) contacting the elements of the microarray of claim 46 with the labeled polynucleotides
 of the sample under conditions suitable for the formation of a hybridization complex,
 and
 - c) quantifying the expression of the polynucleotides in the sample.

48. An array comprising different nucleotide molecules affixed in distinct physical locations on a solid substrate, wherein at least one of said nucleotide molecules comprises a first oligonucleotide or polynucleotide sequence specifically hybridizable with at least 30 contiguous nucleotides of a target polynucleotide, and wherein said target polynucleotide is a polynucleotide of claim 12.

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- 49. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 30 contiguous nucleotides of said target polynucleotide.
- 50. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to at least 60 contiguous nucleotides of said target polynucleotide.
 - 51. An array of claim 48, wherein said first oligonucleotide or polynucleotide sequence is completely complementary to said target polynucleotide.
- 52. An array of claim 48, which is a microarray.
 - 53. An array of claim 48, further comprising said target polynucleotide hybridized to a nucleotide molecule comprising said first oligonucleotide or polynucleotide sequence.
- 54. An array of claim 48, wherein a linker joins at least one of said nucleotide molecules to said solid substrate.
 - 55. An array of claim 48, wherein each distinct physical location on the substrate contains multiple nucleotide molecules, and the multiple nucleotide molecules at any single distinct physical location have the same sequence, and each distinct physical location on the substrate contains nucleotide molecules having a sequence which differs from the sequence of nucleotide molecules at another distinct physical location on the substrate.
 - 56. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:1.

- 57. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:2.
- 58. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:3.

59. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:4. 60. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:5. 61. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:6. 5 62. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:7. 63. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:8. 10 64. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:9. 65. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:10. 66. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:11. 15 67. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:12. 68. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:13. 20 69. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:14. 70. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:15. 71. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:16. 25 72. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:17. 73. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:18. 30 74. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:19. 75. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:20.

76. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:21. 77. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:22. 78. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:23. 5 79. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:24. 80. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:25. 10 81. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:26. 82. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:27. 83. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:28. 15 84. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:29. 85. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:30. 20 86. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:31. 87. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:32. 88. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:33. 25 89. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:34. 90. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:35. 30 91. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:36. 92. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:37.

93. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:38. 94. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:39. 95. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:40. 5 96. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:41. 97. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:42. 10 98. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:43. 99. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:44. 100. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:45. 15 101. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:46. 102. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:47. 20 103. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:48. 104. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:49. 25 105. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:50. 106. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:51. 107. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:52. 30 108. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:53. 109. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:54.

110. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:55.

- 111. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:56.
- 5 112. A polypeptide of claim 1, comprising the amino acid sequence of SEQ ID NO:57.
 - 113. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:58.
- 10 114. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:59.
 - 115. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:60.
 - 116. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:61.

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- 117. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:62.
 - 118. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:63.
- 25 119. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:64.
 - 120. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:65.
 - 121. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:66.

122. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:67.

- 123. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:68.
 - 124. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:69.
- 10 125. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:70.
 - 126. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:71.

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- 127. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:72.
- 128. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID 20 NO:73.
 - 129. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:74.
- 25 130. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:75.
 - 131. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:76.
 - 132. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:77.

133. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:78.

- 134. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:79.
 - 135. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:80.
- 136. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:81.
 - 137. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:82.
- 138. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:83.

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- 139. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:84.
 - 140. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:85.
- 25 141. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:86.
 - 142. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:87.
 - 143. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID . NO:88.

144. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:89.

- 145. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:90.
 - 146. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:91.
- 10 147. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:92.
 - 148. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:93.
 - 149. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:94.

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- 150. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:95.
 - 151. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:96.
- 25 152. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:97.
 - 153. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:98.
 - 154. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:99.

155. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:100.

- 156. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:101.
 - 157. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:102.
- 10 158. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:103.
 - 159. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:104.
 - 160. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:105.

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- 161. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:106.
 - 162. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:107.
- 25 163. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:108.
 - 164. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:109.
 - 165. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:110.

166. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:111.

- 167. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ IDNO:112.
 - 168. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:113.
- 10 169. A polynucleotide of claim 12, comprising the polynucleotide sequence of SEQ ID NO:114.

PCT/US02/41115 WO 03/054219

<110> INCYTE GENOMICS, INC. AZIMZAI, Yalda BAUGHN, Mariah R. BECHA, Shanya D. BOROWSKY, Mark L. BURFORD, Neil ELLIOTT, Vicki S. EMERLING, Brooke M. FORSYTHE, Ian J. GORVAD, Ann E. GRIFFIN, Jennifer A. KABLE, Amy E. KHARE, Reena LAL, Preeti G. LEE, Ernestine A. LEE, Soo Yeun LI, Joana X. MARQUIS, Joseph P. RAMKUMAR, Jayalaxmi RICHARDSON, Thomas W. SPRAGUE, William W. SWARNAKER, Anita TANG, Y. Tom CHAWLA, Narinder K. WARREN, Bridget A. YUE, Henry

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Lys	Ser	Gly	Thr		Glu	Asp	Asp	Leu		Met	Lys	Leu	Thr	Glu
T10	Tla	Dhe	Leu	275	Aen	Val	T16	Twa	280	ui.	7	71.	C.~	285
116	116	FIIG	цец	290	Asp	vai	116	гуs	295	nis	Arg	116	ser	300
Ala	Lys	Thr	Gln	Met	Ile	Met	Glu	Asp	Trp	Asp	Phe	Leu	Gln	
O1	~	31.	•	305	-1-			~1	310				_	315
GIN	Сув	ATA	Leu	320	TIE	Asn	ser	GIU	325	ser	GIĀ	TIE	Pro	330
Asn	Met	Ala	Pro		Lys	Trp	Thr	Arg		Phe	Val	Gln	Arg	
•	01			335	_	_,	_		340	_	_		_	345
гуs	GIĀ	rys	GIn	320 GTA	Arg	Phe	Arg	Gly	Asn 355	Leu	Ser	Gly	Lys	Arg 360
Val	Asp	Phe	Ser		Arg	Thr	Val	Ile		Pro	Asp	Pro	Asn	
_		_		365				_	370					375
Arg	Ile	Asp	GIu	Val 380	Ala	Val	Pro	Val	His 385	Val	Ala	Lys	Ile	Leu 390
Thr	Phe	Pro	Glu		Val	Asn	Lys	Ala		Ile	Asn	Phe	Leu	
				395					400					405
ГÀЗ	Leu	Val	Gln	Asn 410	Gly	Pro	Glu	Val	His 415	Pro	Gly	Ala	Asn	
Ile	Gln	Gln	Arg		Thr	Gln	Met	Lys		Phe	Leu	Lys	Tyr	420 Glv
				425					430					435
Asn	Arg	Glu	Lys	Met 440	Ala	Gln	Glu	Leu	Lys 445	Tyr	Gly	Asp	Ile	
Glu	Arg	His	Leu		Asp	Gly	Asp	Val	_	Leu	Phe	Asn	Arg	450 Gln
				455					460					465
Pro	Ser	Leu	His	Lys 470	Leu	Ser	Ile		Ala 475	His	Leu	Ala	Arg	
Lvs	Pro	His	Arq		Phe	Arg	Phe		4,5	Cvs	Va1	Cvs	Thr	480 Pro
				485					490					495
Tyr	Asn	Ala	Asp		Asp	Gly	qaA	Glu		Asn	Leu	His	Leu	
Gln	Thr	Glu	Glu	500 Ala	Lvs	Ala	Glu	Ala	505	Va 1	Leu	Met	Glv	510
				515					520		202		O1,	525
Lys	Ala	Asn	Leu		Thr	Pro	Arg	Asn		Glu	Pro	Leu	Ile	
Ala	Tle	Gln	Aen	530 Phe	T.em	Thr	alv	מ 1 מ	535	Lou	T ou	ωb~	T 011	540
				545	_6u	- +++	7	nta	550	nen	ned	IIII	neu	ьув 555
Asp	Thr	Phe	Phe		Arg	Ala	Lys	Ala		Gln	Ile	Ile	Ala	
11=	T.e.u	1 <u>م</u> ري	alv	560	Ac-	Glu	Lare	T1a	565	17-1	>	T	D	570
110	Deu	497	Gry	575	vaħ	GIU	пλя	118	Lуs 580	VAI	Arg	Leu	Pro	Pro 585
Pro	Thr	Ile	Leu	Lys	Pro	Val	Thr	Leu		Thr	Gly	Lys	Gln	

				590					595					600
				605		Pro			610					615
				620		Lys			625					630
Leu	Сув	Ala	Asn	Asp 635	Ser	Tyr	Va1	Thr	Ile 640	Gln	Asn	Ser	Glu	Leu 645
Met	Ser	Gly	Ser	Met 650	Asp	Lys	Gly	Thr	Leu 655	Gly	Ser	Gly	Ser	Lys 660
Asn	Asn	Ile	Phe	Tyr 665	Ile	Leu	Leu	Arg	Asp 670	Trp	Gly	Gln	Asn	Glu 675
Ala	Ala	Asp	Ala	Met 680	Ser	Arg	Leu	Ala	Arg 685	Leu	Ala	Pro	Val	Tyr 690
Leu	Ser	Asn	Arg	Gly 695	Phe	Ser	Ile	Gly	Ile 700	Gly	Asp	Val	Thr	Pro 705
Gly	Gln	Gly	Leu	Leu 710	Lys	Ala	Lys	Tyr	Glu 715	Leu	Leu	Asn	Ala	Gly 720
Tyr	Lys	Lys	Суз	Asp 725	Glu	Tyr	Ile	Glu	Ala 730	Leu	Asn	Thr	Gly	Lys 735
Leu	Gln	Gln	Gln	Pro 740	Gly	Cys	Thr	Ala	Glu 745	Glu	Thr	Leu	Glu	Ala 750
Leu	Ile	Leu	Lys	Glu 755	Leu	Ser	Val	Ile	Arg 760	Asp	His	Ala	Gly	Ser 765
Ala	Сув	Leu	Arg	Glu 770	Leu	qaA	Lys	Ser	Asn 775	Ser	Pro	Leu	Thr	Met 780
Ala	Leu	Cys	Gly	Ser 785	Lys	Gly	Ser	Phe	Ile 790	Asn	Ile	Ser	Gln	Met 795
Ile	Ala	Cys	Val	Gly 800	Gln	Gln	Ala	Ile	Ser 805	Gly	Ser	Arg	Val	Pro 810
Asp	Gly	Phe	Glu	Asn 815	Arg	Ser	Leu	Pro	His 820	Phe	Glu	Lys	His	Ser 825
Lys	Leu	Pro	Ala	Ala 830	Lys	Gly	Phe	Val	Ala 835	Asn	Ser	Phe	Tyr	Ser 840
Gly	Leu	Thr	Pro	Thr 845	Glu	Phe	Phe	Phe	His 850	Thr	Met	Ala	G1y	Arg 855
Glu	Gly	Leu	Val	Asp 860	Thr	Ala	Val	Lys	Thr 865	Ala	Glu	Thr	Gly	Tyr 870
Met	Gln	Arg	Arg	Leu 875	Val	Lys	Ser	Leu	Glu 880	Aap	Leu	Cys	Ser	Gln 885
Tyr	Asp	Leu	Thr	Val 890	Arg	Ser	Ser	Thr	Gly 895	Asp	Ile	Ile	Gln	Phe 900
Ile	Tyr	Gly	Gly	Asp 905	Gly	Leu	Asp	Pro	Ala 910	Ala	Met	Glu	Gly	Lys 915
Asp	Glu	Pro	Leu	Glu 920	Phe	Lys	Arg	Val	Leu 925	Asp	Asn	Ile	Lys	Ala 930
Va1	Phe	Pro	Cys	Pro 935	Ser	Glu	Pro	Ala	Leu 940	Ser	Lys	Asn	Glu	Leu 945
Ile	Leu	Thr	Thr	Glu 950	Ser	Ile	Met	Lys	Lys 955	Ser	Glu	Phe	Leu	Сув 960
Сув	Gln	Asp	Ser	Phe 965	Leu	Gln	Glu	Ile	Lys 970	ГУз	Phe	Ile	Lys	Gly 975
Val	Ser	Glu	Lys	Ile 980	Lys	Lys	Thr	Arg	Asp 985	ГЛа	Tyr	Gly	Ile	Asn 990
qaA	Asn	Gly	Thr	Thr 995	Glu	Pro	Arg		Leu 1000	Tyr	Gln	Leu		Arg 1005
Ile	Thr	Pro	Thr	Gln	Val	Glu	ГЛя	Phe	Leu	Glu	Thr	Сув	Arg	Asp

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1015
              1010
Lys Tyr Met Arg Ala Gln Met Glu Pro Gly Ser Ala Val Gly Ala
              1025
                               1030
Leu Cys Ala Gln Ser Ile Gly Glu Pro Gly Thr Gln Met Thr Leu
                                1045
              1040
Lys Thr Phe His Phe Ala Gly Val Ala Ser Met Asn Ile Thr Leu
                                1060
              1055
Gly Val Pro Arg Ile Lys Glu Ile Ile Asn Ala Ser Lys Ala Ile
                                1075
              1070
Ser Thr Pro Ile Ile Thr Ala Gln Leu Asp Lys Asp Asp Asp Ala
              1085
                                1090
Asp Tyr Ala Arg Leu Val Lys Gly Arg Ile Glu Lys Thr Leu Leu
                                1105
             1100
Gly Glu Val Asn Ala Glu Thr Val Arg Tyr Ser Ile Cys Thr Ser
                               1120
             1115
Lys Leu Arg Val Lys Pro Gly Asp Val Ala Val His Gly Glu Ala
             1130
                               1135
Val Val Cys Val Thr Pro Arg Glu Asn Ser Lys Ser Ser Met Tyr
                               1150
             1145
Tyr Val Leu Gln Phe Leu Lys Glu Asp Leu Pro Lys Val Val Val
             1160
                               1165
                                                   1170
Gln Gly Ile Pro Glu Val Ser Arg Ala Val Ile His Ile Asp Glu
             1175
                               1180
Gln Ser Gly Lys Glu Lys Tyr Lys Leu Leu Val Glu Gly Asp Asn
             1190
                                1195
Leu Arg Ala Val Met Ala Thr His Gly Val Lys Gly Thr Arg Thr
             1205
                                1210
Thr Ser Asn Asn Thr Tyr Glu Val Glu Lys Thr Leu Gly Ile Glu
             1220
                                1225
Ala Ala Arg Thr Thr Ile Ile Asn Glu Ile Gln Tyr Thr Met Val
             1235
                               1240
Asn His Gly Met Ser Ile Asp Arg Arg His Val Met Leu Leu Ser
             1250
                                1255
Asp Leu Met Thr Tyr Lys Gly Glu Val Leu Gly Ile Thr Arg Phe
             1265
                               1270
                                                   1275
Gly Leu Ala Lys Met Lys Glu Ser Val Leu Met Leu Ala Ser Phe
             1280
                               1285
Glu Lys Thr Ala Asp His Leu Phe Asp Ala Ala Tyr Phe Gly Gln
             1295
                               1300
Lys Asp Ser Val Cys Gly Val Ser Glu Cys Ile Ile Met Gly Ile
             1310
                               1315
Pro Met Asn Ile Gly Thr Gly Leu Phe Lys Leu Leu His Lys Ala
                   1330
             1325
Asp Arg Asp Pro Asn Pro Pro Lys Arg Pro Leu Ile Phe Asp Thr
             1340 1345
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Asn Glu Phe His Ile Pro Leu Val Thr
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Gly Asn Leu Gly Thr Gly Ala Gly Lys Gly Glu Leu Glu Arg Ala
                                    25
Phe Ser Tyr Tyr Gly Pro Leu Arg Thr Val Trp Ile Ala Arg Asn
                                    40
Pro Pro Gly Phe Ala Phe Val Glu Phe Glu Asp Pro Arg Asp Ala
                                    55
Glu Asp Ala Val Arg Gly Leu Asp Gly Lys Val Ile Cys Gly Ser
                                    70
                65
Arg Val Arg Val Glu Leu Ser Thr Gly Met Pro Arg Arg Ser Arg
                80
                                     85
Phe Asp Arg Pro Pro Ala Arg Arg Pro Phe Asp Pro Asn Asp Arg
                95
                                    100
Cys Tyr Glu Cys Gly Glu Lys Gly His Tyr Ala Tyr Asp Cys His
                                   115
               110
Arg Tyr Ser Arg Arg Arg Ser Arg Ser Arg Ser Arg Ser His
                                   130
               125
Ser Arg Ser Arg Gly Arg Arg Tyr Ser Arg Ser Arg Ser Arg Ser
                140
                                   145
Arg Gly Arg Arg Ser Arg Ser Ala Ser Pro Arg Arg Ser Arg Ser
                                    160
Ile Ser Leu Arg Arg Ser Arg Ser Ala Ser Leu Arg Arg Ser Arg
                170
                                    175
Ser Gly Ser Ile Lys Gly Ser Arg Tyr Phe Gln Ser Pro Ser Arg
                185
                                    190
Ser Arg Ser Arg Ser Arg Ser Ile Ser Arg Pro Arg Ser Ser Arg
                                    205
Ser Pro Ser Gly Ser Pro Arg Arg Ser Ala Ser Pro Glu Arg Met
                                    220
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Asp
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Met Gly Leu Pro Arg Arg Ala Gly Asp Ala Ala Glu Leu Arg Lys
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Ser Leu Lys Pro Leu Leu Glu Lys Arg Arg Arg Ala Arg Ile Asn
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                                     25
Gln Ser Leu Ser Gln Leu Lys Gly Leu Ile Leu Pro Leu Leu Gly
                                     40
Arg Glu Asp Ala Ser Gly Trp His Thr Trp Leu Pro Leu His Ala
Gln Asn Cys Leu Leu Tyr Ile Gln Ala Pro Glu Gln Pro Pro
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Ala
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Ala Val Pro Pro Glu Lys Leu Glu Gly Ala Gly Ser Ser Ser Ala
                 20
                                     25
Pro Glu Arg Asn Cys Val Gly Ser Ser Leu Pro Glu Ala Ser Pro
                 35
                                     40
Pro Ala Pro Glu Pro Ser Ser Pro Asn Ala Ala Val Pro Glu Ala
                 50
                                     55
Ile Pro Thr Pro Arg Ala Ala Ala Ser Ala Ala Leu Glu Leu Pro
                 65
                                     70
Leu Gly Pro Ala Pro Val Ser Val Ala Pro Gln Ala Glu Ala Glu
                 80
                                     85
Ala Arg Ser Thr Pro Gly Pro Ala Gly Ser Arg Leu Gly Pro Glu
                                    100
Thr Phe Arg Gln Arg Phe Arg Gln Phe Arg Arg Arg Thr Asp Val
                                    115
                                                        120
Arg Ile Thr Gly
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Arg Leu Lys Pro Trp Leu Val Ala Gln Val Asn Ser Cys Gln Tyr
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                                     25
Pro Gly Leu Gln Trp Val Asn Gly Glu Lys Lys Leu Phe Cys Ile
                 35
                                     40
Pro Trp Arg His Ala Thr Arg His Gly Pro Ser Gln Asp Gly Asp
                 50
                                     55
Asn Thr Ile Phe Lys Ala Trp Ala Lys Glu Thr Gly Lys Tyr Thr
                 65
                                     70
Glu Gly Val Asp Glu Ala Asp Pro Ala Lys Trp Lys Ala Asn Leu
                 80
                                     85
Arg Cys Ala Leu Asn Lys Ser Arg Asp Phe Arg Leu Ile Tyr Asp
                 95
                                    100
Gly Pro Arg Asp Met Pro Pro Gln Pro Tyr Lys Ile Tyr Glu Val
                                    115
Cys Ser Asn Gly Pro Ala Pro Thr Asp Ser Gln Pro Pro Glu Asp
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130
Tyr Ser Phe Gly Ala Gly Glu Glu Glu Glu Glu Glu Glu Leu
                140
                                    145
Gln Arg Met Leu Pro Ser Leu Ser Leu Thr Glu Asp Val Lys Trp
                155
                                    160
Pro Pro Thr Leu Gln Pro Pro Thr Leu Gln Pro Pro Val Val Leu
                170
                                    175
Gly Pro Pro Ala Pro Asp Pro Ser Pro Leu Ala Pro Pro Pro Gly
                185
                                    190
Asn Pro Ala Gly Phe Arg Glu Leu Leu Ser Glu Val Leu Glu Pro
                200
                                    205
Gly Pro Leu Pro Ala Ser Leu Pro Pro Ala Gly Glu Gln Leu Leu
                215
                                    220
Pro Asp Leu Leu Ile Ser Pro His Met Leu Pro Leu Thr Asp Leu
                230
                                    235
Glu Ile Lys Phe Gln Tyr Arg Gly Arg Pro Pro Arg Ala Leu Thr
                245
                                    250
Ile Ser Asn Pro His Gly Cys Arg Leu Phe Tyr Ser Gln Leu Glu
                260
                                    265
Ala Thr Gln Glu Gln Val Glu Leu Phe Gly Pro Ile Ser Leu Glu
                275
                                    280
Gln Val Arg Phe Pro Ser Pro Glu Asp Ile Pro Ser Asp Lys Gln
                290
                                    295
Arg Phe Tyr Thr Asn Gln Leu Leu Asp Val Leu Asp Arg Gly Leu
                                    310
Ile Leu Gln Leu Gln Gly Gln Asp Leu Tyr Ala Ile Arg Leu Cys
                320
                                    325
Gln Cys Lys Val Phe Trp Ser Gly Pro Cys Ala Ser Ala His Asp
                335
                                    340
Ser Cys Pro Asn Pro Ile Gln Arg Glu Val Lys Thr Lys Leu Phe
Ser Leu Glu His Phe Leu Asn Glu Leu Ile Leu Phe Gln Lys Gly
                                    370
                365
                                                        375
Gln Thr Asn Thr Pro Pro Pro Phe Glu Ile Phe Phe Cys Phe Gly
                380
                                    385
Glu Glu Trp Pro Asp Arg Lys Pro Arg Glu Lys Lys Leu Ile Thr
                395
                                    400
Val Gln Val Val Pro Val Ala Ala Arg Leu Leu Glu Met Phe
                410
                                    415
Ser Gly Glu Leu Ser Trp Ser Ala Asp Ser Ile Arg Leu Gln Ile
                425
                                    430
Ser Asn Pro Asp Leu Lys Asp Arg Met Val Glu Gln Phe Lys Glu
                440
                                    445
Leu His His Ile Trp Gln Ser Gln Gln Arg Leu Gln Pro Val Ala
                455
                                    460
Gln Ala Pro Pro Gly Ala Gly Leu Gly Val Gly Gln Gly Pro Trp
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                                    475
Pro Met His Pro Ala Gly Met Gln
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Ala Asp Leu Pro Pro Pro Ser Ala Leu Lys Lys Gly Ser Lys Glu
                                    400
Lys Gln Ala Asp Phe Leu Asp Lys Gln Met Ser Arg Asp Glu His
                                    415
                410
Arg Ala Arg Ala Met Lys Ile Pro Phe Thr Asn Asp Lys Ile Ile
                                    430
                425
Asn Leu Pro Val Glu Glu Phe Asn Glu Leu Leu Ser Lys Tyr Gln
                                    445
                440
Leu Ser Glu Ala Gln Leu Ser Leu Ile Arg Asp Ile Arg Arg
                455
                                    460
Gly Lys Asn Lys Met Ala Ala Gln Asn Cys Arg Lys Arg Lys Leu
                                    475
                470
Asp Thr Ile Leu Asn Leu Glu Arg Asp Val Glu Asp Leu Gln Arg
                485
                                    490
Asp Lys Ala Arg Leu Leu Arg Glu Lys Val Glu Phe Leu Arg Ser
                500
                                    505
Leu Arg Gln Met Lys Gln Lys Val Gln Ser Leu Tyr Gln Glu Val
                515
                                    520
Phe Gly Arg Leu Arg Asp Glu Asn Gly Arg Pro Tyr Ser Pro Ser
                530
                                    535
Gln Tyr Ala Leu Gln Tyr Ala Gly Asp Gly Ser Val Leu Leu Ile
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Pro Arg Thr Met Ala Asp Gln Gln Ala Arg Arg Gln Glu Arg Lys
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Pro Lys Asp Arg Arg Lys
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Thr Gln Glu Met Glu Gly Ile Val Ile Val Lys Val Glu Glu Glu
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Asp Glu Glu Asp His Phe Gln Lys Glu Arg Asn Lys Val Glu Ser
                 35
                                     40
Ser Pro Gln Val Leu Ser Arg Ser Thr Thr Met Asn Glu Arg Ala
                 50
                                     55
Leu Leu Ser Ser Tyr Leu Val Ala Tyr Arg Val Ala Lys Glu Lys
                 65
                                     70
Met Ala His Thr Ala Ala Glu Lys Ile Ile Leu Pro Ala Cys Met
                 80
                                     85
Asp Met Val Arg Thr Ile Phe Asp Asp Lys Ser Ala Asp Lys Leu
                 95
                                    100
Arg Thr Ile Pro Leu Ser Asp Asn Thr Ile Ser Arg Arg Ile Cys
Thr Ile Ala Lys His Leu Glu Ala Met Leu Ile Thr Arg Leu Gln
                125
                                    130
Ser Gly Ile Asp Phe Ala Ile Gln Leu Asp Glu Ser Thr Asp Ile
```

				140					145					150
310	Com	٥	Dro	140	Len	T au	Va 1	Туг	Val	Ara	ጥላተ	Va1	Ттъ	
Ala	261	Cys	PIO	155	Dea	Dea	val	-7-	160	9	-3-			165
) en	Agn	Phe	Va1		Asp	Leu	Leu	Cvs	Сув	Leu	Asn	Leu	Asn	
Vab	nop	1110	74.	170					175					180
His	Tle	Thr	G1v		Asp	Leu	Phe	Thr	Glu	Leu	Glu	Asn	Cys	
			,	185					190				-	195
Leu	Gly	Gln	Tyr	Lys	Leu	Asn	Trp	Lys	His	Суз	Lys	Gly	Ile	Ser
			•	200			_	_	205					210
Ser	qaA	Gly	Thr	Ala	Asn	Met	Thr	Gly	Lys	His	Ser	Arg	Leu	Thr
	_	_		215					220					225
Glu	Lys	Leu	Leu	Glu	Ala	Thr	His	Asn	Asn	Ala	Val	Trp	Asn	His
				230					235					240
Сув	Phe	Ile	His		Glu	Ala	Leu	Val	Ser	Lys	Glu	Ile	Ser	
				245				_	250	_				255
Ser	Leu	Met	Asp		Leu	Lys	Asn	Ala	Val	Lys	Thr	Val	Asn	
	_	~-	_	260			a	•	265	T	01	7 1 -	mh -	270
He	Lys	GΙΆ	Ser		Leu	Asn	Ser	Arg	Leu 280	Leu	GIU	шe	Pne	285
C	~ 1	T1_	a1	275	۸	ui.		นเล	Leu	T ou	Dha	Wie	Thr	
Ser	GIU	110	GIY	290	ASII	mis	1111	1113	295	Dea	1110			300
Val	Ara	ТТО	Leu		G1n	Glv	Lvs	Val	Leu	Ser	Ara	Val	Tvr	
	••• 9			305		3	-3 -		310				•	315
Leu	Arg	Asn	Glu	Ile	Tyr	Ile	Phe	Leu	Val	Glu	Lys	Gln	Ser	His
	_			320					325					330
Leu	Ala	Asn	Ile	Phe	Glu	Asp	Asp	Ile	\mathtt{Trp}	Val	Thr	Lys	Leu	Ala
				335					340					345
Tyr	Leu	Ser	Asp	Ile	Phe	Gly	Ile	Leu	Asn	Glu	Leu	Ser	Leu	
	_			350	_	_			355	_	_			360
Met	Gln	Gly	Lys		Asn	Asp	Ile	Phe	Gln	Tyr	Leu	GIu	His	11e 375
•	01	D L -	01 -	365	mb	T	T	7	370 Trp	015	21-	λ ~~~	Lou	
Leu	GIY	Pile	GIII	380	1111	Leu	Deu	Leu	385	GIII	nia	nry	Lou	390
Ser	Δan	Δτα	Pro		Tvr	Tvr	Met	Phe	Pro	Thr	Leu	Leu	Gln	
-				395	-3-	-,-			400					405
Ile	Glu	Glu	Asn		Ile	Asn	Glu	Asp	Сув	Leu	Lys	Glu	Ile	Lys
				410					415					420
Leu	Glu	Ile	Leu	Leu	His	Leu	Thr	Ser	Leu	Ser	Gln	Thr	Phe	Asn
				425					430					435
Tyr	Tyr	Phe	Pro		Glu	Lys	Phe	Glu	Ser	Leu	Lys	Glu	Asn	
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Trp	Met	Lys	Asp			Ala	Pne	GIN	460	Pro	GIu	ser	TIE	Ile 465
01	T 011	λαυ	T ou	455		G 111	G1.,	Glu	Asn	Glu	T.eu	T.em	Gln	
GIU	rea	ASII	Leu	470	FIO	GIU	GIU	GIG	475	GIU	ПСС	БСС	G 1	480
Ser	Ser	Ser	Phe		Leu	Lvs	Asn	Tvr	Tyr	Lvs	Ile	Leu	Ser	
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Ser	Ala	Phe	Trp	Ile	Lys	Ile	Lys	Asp	Asp	Phe	Pro	Leu	Leu	Ser
			_	500	_				505					510
Arg	Lys	Ser	Ile	Leu	Leu	Leu	Leu	Pro	Phe	Thr	Thr	Thr	Tyr	Leu
				515					520					525
Сув	Glu	Leu	Gly			Ile	Leu	Thr	Arg		Lys	Thr	Lys	
	_			530			_		535		7.F. 7		T	540
Arg	Asn	Arg	Leu			Ala	Pro	qaA		Arg	vai	Ala	Leu	Ser 555
Sa-	رب. د	v-1	Dra	545		Lare	Gl.	Lev	550	λor	Ara	G1n	alه	His
261	cys	AGI	110	ოინ	ιτb	пλя	GIU	₽ e u	1.100	Vall		0111	nia	

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Pro Ser His

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Leu Lys Thr Leu Ala Asp Ile Ile Cys Glu Tyr Pro Asp Ile His
                                    325
Gln Ser Ser Arg Phe Val Phe Val Pro Gly Pro Glu Asp Pro Gly
                335
                                    340
Phe Gly Ser Ile Leu Pro Arg Pro Pro Leu Ala Glu Ser Ile Thr
                                    355
Asn Glu Phe Arg Gln Arg Val Pro Phe Ser Val Phe Thr Thr Asn
                365
                                    370
Pro Cys Arg Ile Gln Tyr Cys Thr Gln Glu Ile Thr Val Phe Arg
                380
                                    385
Glu Asp Leu Val Asn Lys Met Cys Arg Asn Cys Val Arg Phe Pro
                395
                                    400
Ser Ser Asn Leu Ala Ile Pro Asn His Phe Val Lys Thr Ile Leu
                410
                                    415
                                                         420
Ser Gln Gly His Leu Thr Pro Leu Pro Leu Tyr Val Cys Pro Val
                425
                                    430
Tyr Trp Ala Tyr Asp Tyr Ala Leu Arg Val Tyr Pro Val Pro Asp
                440
                                    445
Leu Leu Val Ile Ala Asp Lys Tyr Asp Pro Phe Thr Thr Asn
                455
                                    460
Thr Glu Cys Leu Cys Ile Asn Pro Gly Ser Phe Pro Arg Ser Gly
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Phe Ser Phe Lys Val Phe Tyr Pro Ser Asn Lys Thr Val Glu Asp
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Ser Lys Leu Gln Gly Phe
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Tyr Arg Val Leu Glu Ala Gly Lys Ile Gly Ile Phe Glu Ser Pro
                 35
                                     40
Thr Gly Thr Gly Lys Ser Leu Ser Leu Ile Cys Gly Ala Leu Ser
                 50
                                     55
Trp Leu Arg Asp Phe Glu Gln Lys Lys Arg Glu Glu Glu Ala Arg
                 65
                                     70
Leu Leu Glu Thr Gly Thr Gly Pro Leu His Asp Glu Lys Asp Glu
                 80
                                     85
Ser Leu Cys Leu Ser Ser Ser Cys Glu Gly Ala Ala Gly Thr Pro
                 95
                                    100
Arg Pro Ala Gly Glu Pro Ala Trp Val Thr Gln Phe Val Gln Lys
                110
                                    115
Lys Glu Glu Arg Asp Leu Val Asp Arg Leu Lys Ala Glu Gln Ala
                125
                                    130
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Arg Arg Lys Gln Arg Glu Glu Arg Leu Gln Gln Leu Gln His Arg

				140					145					150
Val	Gln	Leu	Lys		Ala	Ala	Lys	Arg	145 Leu	Arg	Gln	Glu	Glu	
				155			_	_	160					165
Glu	Arg	Glu	Asn	Leu 170	Leu	Arg	Leu	Ser	Arg 175	Glu	Met	Leu	Glu	Thr 180
Gly	Pro	Glu	Ala	_	Arg	Leu	Glu	Gln		Glu	Ser	Gly	Glu	Glu
-1				185	01	_	-1	_	190	a1	01	•	·	195
GIU	ren	Val	Leu	200	Giu	ıyr	GIU	ser	205	GIU	GIU	гÀв	ràs	210
Ala	Ser	Arg	Val	Asp 215	Glu	Asp	Glu	Asp	Asp 220	Leu	Glu	Glu	Glu	His 225
Ile	Thr	Lys	Ile	Tyr	Tyr	Cys	Ser	Arg	Thr	His	Ser	Gln	Leu	Ala
01=	Dho	17.01	wi-	230	vo 1	7.40	T	S	235	Pho	01	Tuo	N ==	240
GIII	FIIG	Val	UIS	245	Vai	Буs	Lys	261	250	rne	GLY	цув	App	255
Arg	Leu	Va1	Ser	Leu	Gly	Ser	Arg	Gln	Asn	Leu	Cys	Val	Asn	Glu
		_	_	260	~1	_	1	-1	265	-1.				270
Asp	vaı	Lys	Ser	275	GIÀ	Ser	val	GIN	280	IIe	Asn	Asp	Arg	285
Val	yab	Met	Gln		Ser	Arg	His	Glu		Lys	Lys	Gly	Ala	
Glu	Glu	Lys	Pro	290	Ara	Ara	Ara	G1n	295 Glu	Lvs	G1n	λla	Ala	300 Cvs
		,		305					310	_,_				315
Pro	Phe	Tyr	Asn	His 320	Glu	Gln	Met	Gly	Leu 325	Leu	Arg	Asp	Glu	Ala 330
Leu	Ala	G1u	Val		Asp	Met	Glu	Gln		Leu	Ala	Leu	Gly	
				335	_				340				_	345
Glu	Ala	Arg	Ala	Cys 350	Pro	Tyr	Tyr	Gly	Ser	Arg	Leu	Ala	Ile	Pro 360
Ala	Ala	Gln	Leu		Val	Leu	Pro	Tyr		Met	Leu	Leu	His	
				365					370					375
Ala	Thr	Arg	Gln	Ala 380	Ala	Gly	Ile	Arg	Leu 385	Gln	Asp	Gln	Val	Val 390
Ile	Ile	Asp	Glu	Ala	His	Asn	Leu	Ile	Asp	Thr	Ile	Thr	Gly	Met
uic	Ser.	Val	Gl.	395	Ser	Gly	Ser	Gln	400	Ove.	Gln	λl =	น่าย	405
1113	561	vai	GIU	410	561	OLY	561	O ₂ II	415	Cys	0411	nia	114.5	420
Gln	Leu	Leu	Gln		Val	Glu	Arg	Tyr		Lys	Arg	Leu	Lys	
Lvs	Asn	Leu	Met	425 Tvr	Leu	Lvs	Gln	Ile	430 Leu	Tvr	Leu	Leu	Glu	435 Lvs
-3-				440		-2 -			445					450
Phe	Val	Ala	Val	Leu 455	Gly	Gly	Asn	Ile	Lys 460	Gln	Asn	Pro	Asn	Thr 465
Gln	Ser	Leu	Ser		Thr	Gly	Thr	Glu		Lys	Thr	Ile	Asn	
Phe	Len	Phe	Gln	470 Ser	G1n	Tle	Agn	Asn	475	Asn	Leu	Phe	Lvs	480 Val
1110	Dea	1110	01	485	U		nop	71011	490	non	200		2,0	495
Gln	Arg	Tyr	Сув	Glu 500	Lys	Ser	Met	Ile	Ser 505	Arg	Lys	Leu	Phe	Gly 510
Phe	Thr	Glu	Arg	Tyr	Gly	Ala	Val	Phe	Ser	Ser	Arg	Glu	Gln	Pro
I.ve	Leu	Ala	Glv	515 Phe	G1n	G1 n	Pho	יים.	520 Gln	Ser	T.em	Gln	Pro	525 Ara
~y3		u	OLY	530	J 111	0111	. 116	Lau	535	201	Dou	0111	110	540
Thr	Thr	Glu	Ala		Ala	Ala	Pro	Ala	_	Glu	Ser	Gln	Ala	
Thr	Leu	Arg	Pro	545 Ala	Ser	Pro	Leu	Met	550 His	Ile	Gln	G1v	Phe	555 Leu
												3		

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560
                                    565
Ala Ala Leu Thr Thr Ala Asn Gln Asp Gly Arg Val Ile Leu Ser
                                    580
Arg Gln Gly Ser Leu Ser Gln Ser Thr Leu Lys Phe Leu Leu Leu
                590
                                    595
Asn Pro Ala Val His Phe Ala Gln Val Val Lys Glu Cys Arg Ala
                605
                                     610
Val Val Ile Ala Gly Gly Thr Met Gln Pro Val Ser Asp Phe Arg
                620
                                    625
Gln Gln Leu Leu Ala Cys Ala Gly Val Glu Ala Glu Arg Val Val
                635
                                    640
Glu Phe Ser Cys Gly His Val Ile Pro Pro Asp Asn Ile Leu Pro
                650
                                    655
Leu Val Ile Cys Ser Gly Ile Ser Asn Gln Pro Leu Glu Phe Thr
                665
                                    670
Phe Gln Lys Arg Glu Leu Pro Gln Met Met Asp Glu Val Gly Arg
                                     685
                680
Ile Leu Cys Asn Leu Cys Gly Val Val Pro Gly Gly Val Val Cys
                                    700
                695
Phe Phe Pro Ser Tyr Glu Tyr Leu Arg Gln Val His Ala His Trp
                                    715
                710
Glu Lys Gly Gly Leu Leu Gly Arg Leu Ala Ala Arg Lys Lys Ile
                725
                                    730
Phe Gln Glu Pro Lys Ser Ala His Gln Val Glu Gln Val Leu Leu
                                    745
Ala Tyr Ser Arg Cys Ile Gln Ala Cys Gly Gln Glu Arg Gly Gln
                                    760
Val Thr Gly Ala Leu Leu Ser Val Val Gly Gly Lys Met Ser
                770
                                     775
Glu Gly Ile Asn Phe Ser Asp Asn Leu Gly Arg Ser Ala Glu Leu
Gln Glu Lys Met Ala Tyr Leu Asp Gln Thr Leu Pro Arg Ala Pro
                                     805
                800
Gly Gln Ala Pro Pro Gly Lys Ala Leu Val Glu Asn Leu Cys Met
                                     820
                815
Lys Ala Val Asn Gln Ser Ile Gly Arg Ala Ile Arg His Gln Lys
                830
                                     835
Asp Phe Ala Ser Val Val Leu Leu Asp Gln Arg Tyr Ala Arg Pro
                845
                                     850
Pro Val Leu Ala Lys Leu Pro Ala Trp Ile Arg Ala Arg Val Glu
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                860
Val Lys Ala Thr Phe Gly Pro Ala Ile Ala Ala Val Gln Lys Phe
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                                     880
His Arg Glu Lys Ser Ala Ser Ser
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<223> Incyte ID No: 8096611CD1

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Lys	Ile	Trp	Leu	Ala 20	Ala	His	Trp	Glu		Lys	Leu	Thr	Lys	Ala 30
His	Val	Phe	Glu	Cys 35	Asn	Leu	Glu	Ile		Ile	Glu	Lys	Ile	Leu 45
Ser	Pro	Lys	Val	Lys 50	Ile	Ala	Leu	Arg	Thr 55	Ser	Gly	His	Leu	Leu 60
Leu	Gly	Val	Val	Arg 65	Ile	Tyr	Asn	Arg	Lys 70	Ala	Lys	Tyr	Leu	Leu 75
				80		Phe			85					90
				95					100					Tyr 105
				110		Glu			115					120
				125		Val			130					135
				140		Thr			145					150
				155		Ser			160					165
				170		Asp			175					180
				185		Ser			190			_		195
				200		Gly			205					210
				215		Asn			220					225
				230		Leu			235					240
				245		Ala			250					255
				260		Glu			265					270
				275		Phe			280					285
				290		Lys			295					300
				305		Ser			310					315
				320		Leu			325					330
				335		Trp			340					345
				350		Gln			355					360
				365		Phe			370					375
				380		Glu			385					390
				395		Ser			400				_	405
GIII	GIU	neu	ser	1ys 410	PTO	Gln	ınr	ı.tb	Lys 415	Asp	val	TIE	GŢĀ	Gly 420

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Ser Gln His Ser Ser His Glu Asp Thr Asn Lys Asn Ile Asn Ser
                                    430
Glu Asp Ile Val Glu Met Val Ser Leu Ala Ala Glu Glu Ser Ser
                                    445
                440
Leu Met Asn Asp Leu Phe Ala Gln Glu Ile Glu Tyr Ser Pro Val
                                    460
Glu Leu Glu Ser Leu Ser Asn Glu Glu Asn Ile Glu Thr Glu Arg
                470
Trp Asn Gly Arg Ile Leu Gln Met Leu Asn Arg Leu Arg Glu Ser
                                    490
                485
Asn Lys Met Gly Met Gln Ser Phe Ser Leu Met Lys Leu Cys Arg
                500
                                    505
Asn Ser Asp Arg Lys Gln Ala Ala Ala Lys Phe Tyr Ser Phe Leu
                515
                                    520
Val Leu Lys Lys Gln Leu Ala Ile Glu Leu Ser Gln Ser Ala Pro
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               530
Tyr Ala Asp Ile Ile Ala Thr Met Gly Pro Met Phe Tyr Asn Ile
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Pro Ala Val Ser Arg Pro Gly Ala Glu Asn Ile Leu Thr His Glu
                200
                                     205
Gln Arg Cys Ala Ala Leu Val Ser Ala Gly Phe Asp Leu Leu Leu
                215
                                     220
Asp Glu Arg Ser Pro Tyr Trp Ala Cys Lys Gly Thr Val Ala Gly
Val Ile Leu Glu Arg Glu Ile Pro Arg Ala Arg Gly His Val Lys
                245
                                     250
Glu Ile Tyr Lys Leu Val Ala Leu Gly Thr Gly Ser Ser Cys Cys
                260
                                     265
Ala Gly Trp Leu Glu Phe Ser Gly Gln Gln Leu His Asp Cys His
                275
                                     280
Gly Leu Val Ile Ala Arg Arg Ala Leu Leu Arg Phe Leu Phe Arg
                290
                                     295
Gln Leu Leu Ala Thr Gln Gly Gly Pro Lys Gly Lys Glu Gln
                305
                                     310
Ser Val Leu Ala Pro Gln Pro Gly Pro Gly Pro Pro Phe Thr Leu
                320
                                    325
Lys Pro Arg Val Phe Leu His Leu Tyr Ile Ser Asn Thr Pro Lys
                335
                                    340
Gly Ala Ala Arg Asp Ile Tyr Leu Pro Pro Thr Ser Glu Gly Gly
                350
                                    355
Leu Pro His Ser Pro Pro Met Arg Leu Gln Ala His Val Leu Gly
                365
                                    370
Gln Leu Lys Pro Val Cys Tyr Val Ala Pro Ser Leu Cys Asp Thr
                380
                                    385
His Val Gly Cys Leu Ser Ala Ser Asp Lys Leu Ala Arg Trp Ala
                                     400
Val Leu Gly Leu Gly Gly Ala Leu Leu Ala His Leu Val Ser Pro
                410
                                    415
Leu Tyr Ser Thr Ser Leu Ile Leu Ala Asp Ser Cys His Asp Pro
                                     430
Pro Thr Leu Ser Arg Ala Ile His Thr Arg Pro Cys Leu Asp Ser
                                     445
Val Leu Gly Pro Cys Leu Pro Pro Pro Tyr Val Arg Thr Ala Leu
                455
                                    460
His Leu Phe Ala Gly Pro Pro Val Ala Pro Ser Glu Pro Thr Pro
                470
                                    475
Asp Thr Cys Arg Gly Leu Ser Leu Asn Trp Ser Leu Gly Asp Pro
                485
                                    490
Gly Ile Glu Val Val Asp Val Ala Thr Gly Arg Val Lys Ala Asn
                500
                                    505
Ala Ala Leu Gly Pro Pro Ser Arg Leu Cys Lys Ala Ser Phe Leu
                515
                                    520
Arg Ala Phe His Gln Ala Ala Arg Ala Val Gly Lys Pro Tyr Leu
                530
                                    535
Leu Ala Leu Lys Thr Tyr Glu Ala Ala Lys Ala Gly Pro Tyr Gln
                545
                                    550
Glu Ala Arg Arg Gln Leu Ser Leu Leu Leu Asp Gln Gln Gly Leu
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Gly Ala Trp Pro Ser Lys Pro Leu Val Gly Lys Phe Arg Asn
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<212> PRT

<213> Homo sapiens

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<223> Incyte ID No: 3101042CD1

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370
Ser Ser Val Leu Gln Val His Trp Arg Phe His Thr Gly Glu Lys
                                    385
                380
Pro Tyr Arg Cys Gly Glu Cys Gly Lys Gly Phe Ser Gln Cys Thr
                                    400
His Leu His Ile His Gln Arg Val His Thr Gly Glu Lys Pro Tyr
                                    415
                410
Lys Cys Asn Val Cys Gly Lys Asp Phe Ala Tyr Ser Ser Val Leu
                                                         435
                425
                                    430
His Thr His Gln Arg Val His Thr Gly Glu Lys Pro Tyr Lys Cys
                440
                                    445
Glu Val Cys Gly Lys Cys Phe Ser Tyr Ser Ser Tyr Phe His Leu
                455
                                    460
His Gln Arg Asp His Ile Arg Glu Lys Pro Tyr Lys Cys Asp Glu
                                    475
                470
Cys Gly Lys Gly Phe Ser Arg Asn Ser Asp Leu Asn Val His Leu
                                    490
                485
Arg Val His Thr Arg Glu Arg Pro Tyr Lys Cys Lys Ala Cys Gly
                500
                                    505
Lys Gly Phe Ser Arg Asn Ser Tyr Leu Leu Ala His Gln Arg Val
                515
                                    520
His Ile Asp Glu Thr Gln Tyr Thr His Cys Glu Arg Gly Lys Asp
                530
                                    535
Leu Leu Thr His Gln Arg Leu His Glu Gln Arg Glu Thr Leu
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                                     25
Pro Lys Thr Asp Glu Ala Glu Lys Arg Ser Arg Lys Pro Glu Lys
                 35
                                     40
Glu Pro Arg Arg Ser Gly Arg Ala Thr Asn His Asp Ser Cys Asp
                                     55
                 50
Ser Cys Lys Glu Gly Gly Asp Leu Cys Cys Asp His Cys Pro
                 65
                                     70
Ala Ala Phe His Leu Gln Cys Cys Asn Pro Pro Leu Ser Glu Glu
                 80
                                     85
Met Leu Pro Pro Gly Glu Trp Met Cys His Arg Cys Thr Val Arg
                 95
                                    100
Arg Lys Lys Arg Glu Gln Lys Lys Glu Leu Gly His Val Asn Gly
                110
                                    115
Leu Val Asp Lys Ser Gly Lys Arg Thr Thr Ser Pro Ser Ser Asp
                125
                                    130
Thr Asp Leu Leu Asp Arg Ser Ala Ser Lys Thr Glu Leu Lys Ala
                                    145
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Ile	Ala	His	Ala	Arg 155	Ile	Leu	Glu	Arg	Arg 160	Ala	Ser	Arg	Pro	Gly 165
Thr	Pro	Thr	Ser	Ser 170	Ala	Ser	Thr	Glu		Pro	Thr	Ser	Glu	_
Asn	Asp	Val	Asp	Glu 185	Asp	Ile	Ile	Asp	Val 190	Asp	Glu	Glu	Pro	Val 195
Ala	Ala	G1u	Pro	Asp 200	Tyr	Val	Gln	Pro	Gln 205	Leu	Arg	Arg	Pro	Phe 210
Glu	Leu	Leu	Ile	Ala 215	Ala	Ala	Met	Glu	Arg 220	Asn	Pro	Thr	Gln	Phe 225
Gln	Leu	Pro	Asn	Glu 230	Leu	Thr	Сув	Thr	Thr 235	Ala	Leu	Pro	Gly	Ser 240
Ser	Lys	Arg	Arg	Arg 245	ГЛа	Glu	Glu	Thr	Thr 250	Gly	Lys	Asn	Val	Lys 255
Lys	Thr	Gln	His	Glu 260	Leu	Asp	His	Asn	Gly 265	Leu	Val	Pro	Leu	Pro 270
	_			Phe 275					280					285
			_	Asp 290	_	_			295				_	300
				Leu 305					310	_		_		315
				G1u 320			_		325		_			330
			_	Cys 335				_	340					345
				Va1 350					355					360
_				Asn 365	_				370			_		375
		_		Pro 380				_	385		_			390
				Ala 395					400					405
		_		Pro 410					415					420
				Thr 425					430				_	435
				Gln 440					445					450
				His 455		_			460					465
				11e 470					475					480
			_	Lys 485					490		_			495
_			_	11e 500					505			_		510
				Pro 515					520	_	_			525
				Lys 530					535					540
				Val 545					550					555
Pro	Thr	Asp	Ser	Thr 560	Asp	Pro	Arg	Arg	Leu 565	Pro ·	Gly	Ala	Asn	Thr 570

				575					580			Arg		585
				590					595			Val	_	600
Ile	Val	Lys	Thr	Glu 605	Asn	Ala	Thr	Gly	Pro 610	Ser	Ser	Сув	Pro	Gln 615
				620					625			Ile		630
				635					640			Arg		645
				650					655			Arg		660
				665					670			Pro		675
				680					685			Arg		690
				695					700			Gly		705
				710					715			Ala		720
				725					730			Phe		735
				740					745			Lys		750
				755					760			Pro		765
				770					775			Leu		780
				785					790			Gln		795
				800					805			Val		810
				815					820			Met	-	825
				830					835			Gly		840
				845					850			Glu		855
				860					865			Leu		870
				875					880			Ser		885
				890					895			His Ser		900
				905					910			Ala		915
				920					925			Gly		930
				935					940			Leu		945
				950					955			Lys		960
				965					970			Leu		975
				980		~13	val	Dou	985	Jiu	-uy s	Tea	261	990

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Thr Arg Pro Thr Lys Ser Ser 110

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Ala	Ala	Ala	Thr		Ala	Gly	Pro	Ala		Val	Pro	Pro	Gln	-
				335					340					345
Tyr	Gly	Val	Pro	Trp	Gly	Val	Tyr	Pro	Ala	Asn	Leu	Phe	Gln	Gln
				350					355					360
Gln	Ala	Ala	Ala	Ala	Ala	Asn	Asn	Thr	Ala	Ser	Gln	Gln	Ala	Ala
•				365			•		370					375
_		- 1	~1		01	~ 1	01	01		•	.		01	
Ser	Gln	Ala	Gin		GIY	GIN	GIN	GIN		Leu	Arg	Ата	GIĀ	
				380					385					390
Gly	Gln	Arg	Pro	Leu	Thr	Pro	Asn	Gln	Gly	Gln	Gln	Gly	Gln	Gln
				395					400					405
Ala	Glu	Ser	Leu	Ala	Ala	Ala	Ala	Ala	Ala	Asn	Pro	Thr	Leu	Ala
				410					415					420
Dha	Gly	01-	C1	_	21.	mb~	01.	Wat		01	т	01-	17-1	
FIIE	GIA	GIII	GIY		MIG	1111	GIY	Mec		GIY	IYI	GIII	Val	
_		_	_	425			_		430	_		_	_	435
Ala	Pro	Thr	Ala	Tyr	Tyr	Asp	Gln	Thr	Gly	Ala	Leu	Val	Val	Gly
				440					445					450
Pro	Gly	Ala	Arg	Thr	Gly	Leu	Gly	Ala	Pro	Val	Arg	Leu	Met	Ala
				455					460					465
Pro	Thr	Pro	Va 1		T1a	Ser	Ser	Δla	Δla	Δla	Gln	Δla	Δla	
110		110	V41	470	110	501	501	7,14	475	mru	U 111		niu	480
					_,	_,		_		_	1	_,	_	
Ala	Ala	Ala	Ala	-	GIA	Thr	Ala	Ser		Leu	Thr	GIÀ	Ser	
				485					490					495
Asn	Gly	Leu	Phe	Arg	Pro	Ile	Gly	Thr	Gln	Pro	Pro	Gln	Gln	Gln
				500					505					510
G1n	Gln	Gln	Pro	Ser	Thr	Asn	Leu	Gln	Ser	Asn	Ser	Phe	Tvr	Glv
				515					520				•	525
C	C	C	T 011	_) an	C ~ ~	°-~	01-		C	Ca=	Lon	Dho	
261	Ser	Ser	red		ASII	ser	Ser	GIII		Ser	Ser	ьеи	rne	
				530	_				535	_				540
His	Gly	Pro	Gly		Pro	Gly	Ser	Thr		Leu	Gly	Phe	Gly	
				545					550					555
Gly	Asn	Ser	Leu	Gly	Ala	Ala	Ile	Gly	Ser	Ala	Leu	Ser	Gly	Phe
				560					565					570
Glv	Ser	Ser	Glv	Glv	Leu	Thr	Asn	Glv	Ser	Glv	Ara	Tvr	Ile	Ser
,			2	575				2	580	3	5	-1-		585
	.1.	D	01		01	N1	T			C		C	C	
AIA	Ala	PIO	GIY		GIU	ALA	гЛя	IÀT		Set	AIA	Ser	261	
				590				_	595	_				600
Ser	Ser	Leu	Phe	Ser	Ser	Ser	Ser	Gln	Leu	Phe	Pro	Pro	Ser	Arg
				605					610					615
Leu	Arg	Tyr	Asn	Arg	Ser	Asp	Ile	Met	Pro	Ser	Gly	Arg	Ser	Arg
				620					625					630
Leu	Leu	Glu	Asp	Phe	Ara	Asn	Asn	Ara	Phe	Pro	Asn	Leu	Gln	Leu
				635	9				640					645
.		•	-1 -		***	-1 -	*** 1	~1			01	3	O1	
Arg	Asp	Leu	TIE		HIS	TIE	val	GIU		ser	GIN	ASD	GIN	
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Gly	Ser	Arg	Phe	Ile	Gln	Gln	Lys	Leu	Glu	Arg	Ala	Thr	Pro	
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Glu	Arg	Gln	Met	Val	Phe	Asn	Glu	Ile	Leu	Gln	Ala	Ala	Tyr	Gln
	•			680					685				-	690
T.em	Met	Thr	Agn		Dhe	Glv	Δen	ጥረታ		T۱۵	G1n	Tare	Dhe	
Dea	Mec	1111	иор	695	FIIG	GLY	VOII	1 Y L	700	110	G111	ry 5	FIIG	
	_,		_		_		_	_		_		_,	_	705
Glu	Phe	GIA	ser		Asp	GIn	гàг	Leu		Leu	Ala	Thr	Arg	
				710					715					720
Arg	Gly	His	Val	Leu	Pro	Leu	Ala	Leu	Gln	Met	Tyr	Gly	Сув	Arg
				725					730					735
Val	Ile	Gln	Lvs	Ala	Leu	Glu	Ser	Ile	Ser	Ser	Asp	Gln	Gln	

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745
Ile Ser Glu Met Val Lys Glu Leu Asp Gly His Val Leu Lys Cys
                                    760
                755
Val Lys Asp Gln Asn Gly Asn His Val Val Gln Lys Cys Ile Glu
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Cys Val Gln Pro Gln Ser Leu Gln Phe Ile Ile Asp Ala Phe Lys
Gly Gln Val Phe Val Leu Ser Thr His Pro Tyr Gly Cys Arg Val
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                                    805
Ile Gln Arg Ile Leu Glu His Cys Thr Ala Glu Gln Thr Leu Pro
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Ile Leu Glu Glu Leu His Gln His Thr Glu Gln Leu Val Gln Asp
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                                    835
Gln Tyr Gly Asn Tyr Val Ile Gln His Val Leu Glu His Gly Arg
                                    850
                845
Pro Glu Asp Lys Ser Lys Ile Val Ser Glu Ile Arg Gly Lys Val
                860
                                    865
Leu Ala Leu Ser Gln His Lys Phe Ala Ser Asn Val Val Glu Lys
                875
                                    880
Cys Val Thr His Ala Ser Arg Ala Glu Arg Ala Leu Leu Ile Asp
                890
                                    895
Glu Val Cys Cys Gln Asn Asp Gly Pro His Ser Ala Leu Tyr Thr
                905
                                    910
Met Met Lys Asp Gln Tyr Ala Asn Tyr Val Val Gln Lys Met Ile
                920
                                    925
Asp Met Ala Glu Pro Ala Gln Arg Lys Ile Ile Met His Lys Ile
                                    940
Arg Pro His Ile Thr Thr Leu Arg Lys Tyr Thr Tyr Gly Lys His
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Ile Leu Ala Lys Leu Glu Lys Tyr Tyr Leu Lys Asn Ser Pro Asp
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Leu Gly Pro Ile Gly Gly Pro Pro Asn Gly Met Leu
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Val Leu Arg Gln Leu Asn Glu Gln Arg Leu Arg Gly Leu Phe Cys
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Asp Val Thr Leu Ile Ala Gly Asp Thr Lys Phe Pro Ala His Arg
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35

50

65

40

55

70

Ser Val Leu Ala Ala Ser Ser Pro Phe Phe Arg Glu Ala Leu Leu

Thr Ser Ala Pro Leu Pro Leu Pro Pro Ala Thr Gly Gly Ala Ala

Pro Asn Pro Ala Thr Thr Ala Ala Ser Ser Ser Ser Ser Ser

Ser	Ser	Ser	Ser	Ser 95	Ser	Ser	Ser	Ser	Ser 100	Ala	Ser	Ser	Ser	Ser 105
Ser	Ser	Ser	Ser	Ser 110	Ser	Pro	Pro	Pro	Ala 115	Ser	Pro	Pro	Ala	Ser 120
Ser	Pro	Pro	Arg	Val 125	Leu	Glu	Leu	Pro	Gly 130	Val	Pro	Ala	Ala	Ala 135
Phe	Ser	Asp	Val	Leu 140	Asn	Phe	Ile	Tyr	Ser 145	Ala	Arg	Leu	Ala	Leu 150
Pro	Gly	Gly	Gly	Gly 155	Asp	Gly	Ala	Ala	Val 160	Ala	Glu	Ile	Gly	Ala 165
Leu	Gly	Arg	Arg	Leu 170	Gly	Ile	Ser	Arg	Leu 175	Gln	Gly	Leu	Gly	Glu 180
Gly	Gly	Asp	Ala	Trp 185	Val	Pro	Pro	Thr	Pro 190	Ala	Pro	Met	Ala	Thr 195
				Glu 200	_			_	205			_		210
				Gly 215					220					225
				Pro 230					235					240
				His 245					250					255
				Ala 260					265				_	270
				Pro 275					280					285
				Val 290 Gly					295					300
				305 Thr				_	310	-			_	315
		_		320 Arg					325					330
				335 Glu	_			_	340				_	345
				350 Tyr					355					360
				365 Leu					370					375
				380 Leu					385					390
Lys	Pro	Lys	Leu	395 Asn	Thr	Leu	Lys	Leu	400 Tyr	Arg	Leu	Leu	Pro	405 Met
Arg	Ala	Ala	Lys	410 Arg	Pro	Tyr	Lys	Thr	415 Tyr	Ser	Gln	Gly	Ala	420 Pro
Glu	Ala	Pro	Leu	425 Ser	Pro	Thr	Leu	Asn	430 Thr	Pro	Ala	Pro	Val	435 Ala
Met	Pro	Ala	Ser	440 Pro	Pro	Pro	Gly	Pro	445 Pro	Pro	Ala	Pro	Glu	450 Pro
Gly	Pro	Pro	Pro	455 Ser	Val	Ile	Thr	Phe		His	Pro	Ala	Pro	
Val	Ile	Val	His	470 Gly	Gly	Ser	Ser	Ser		Gly	Gly	Gly	Ser	
Thr	Ala	Ser	Thr	485 Gly	Gly	Ser	Gln	Ala		Ser	Val	Ile	Thr	
				500					505					510

Thr	Ala	Pro	Pro	Arg 515	Pro	Pro	Lys	Lys	Arg 520	Glu	Tyr	Pro	Pro	Pro 525
Pro	Pro	Glu	Pro	Ala 530	Ala	Thr	Pro	Thr	Ser 535	Pro	Ala	Thr	Ala	Val 540
Ser	Pro	Ala	Thr	Ala 545	Ala	Gly	Pro	Ala	Met 550	Ala	Thr	Thr	Thr	Glu 555
Glu	Ala	Lys	Gly	Arg 560	Asn	Pro	Arg	Ala	Gly 565	Arg	Thr	Leu	Thr	Tyr 570
Thr	Ala	Lys	Pro	Val 575	Gly	Gly	Ile	Gly	Gly 580	Gly	Gly	Gly	Pro	Pro 585
Thr	Gly	Ala	Gly	Arg 590	Gly	Pro	Ser	Gln	Leu 595	Gln	Ala	Pro	Pro	Pro 600
	-		Ile	605		_			610					615
_	_		Ser	620					625					630
			Glu	635					640					645
			Glu	650	_				655	_		_	_	660
			Trp	665					670					675
	_		Ala	680					685					690
_	_	_	Arg	695		_	_		700	_				705
			Glu	710					715					720
_			Arg	725					730					735
			Arg	740					745					750
			Ser	755					760					765
			His	770					775					780
			Gly	785					790					795
			Val	800					805					810
				815					820					Ser 825
				830					835					Glu 840
			Thr	845					850					855
			Pro	860				_	865		_		_	870
			Glu	875					880					885
			Gly	890					895					900
			Gly	905					910					915
Thr	Phe	Tyr	Pro	Glu 920	Pro	Tyr	Pro	Leu	Val 925	Tyr	Gly	Pro	Gln	Leu 930

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Leu Ala Ala Tyr Pro Tyr Asn Phe Ser Asn Leu Ala Ala Leu Pro
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Val Ala Leu Asn Met Val Leu Pro Asp Glu Lys Gly Ala Gly Ala
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                                    955
Leu Pro Phe Leu Pro Gly Val Phe Gly Tyr Ala Val Asn Pro Gln
                                    970
Ala Ala Pro Pro Ala Pro Pro Thr Pro Pro Pro Pro Thr Leu Pro
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Pro Pro Ile Pro Pro Lys Gly Glu Gly Glu Arg Ala Gly Val Glu
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Arg Thr Gln Lys Gly Asp Val Gly
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Ala Gln Arg Arg Gly Glu Asp Val Glu Thr Ser Lys Lys Trp Ala
Ala Gly Gln Asn Lys Gln His Ser Ile Thr Lys Asn Thr Ala Lys
Leu Asp Arg Glu Thr Glu Glu Leu His His Asp Arg Val Thr Leu
                 65
Glu Val Gly Lys Val Ile Gln Gln Gly Arg Gln Ser Lys Gly Leu
                80
                                     85
Thr Gln Lys Asp Leu Ala Thr Lys Ile Asn Glu Lys Pro Gln Val
                95
                                   100
Ile Ala Asp Tyr Glu Ser Gly Arg Ala Ile Pro Asn Asn Gln Val
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Leu Gly Lys Ile Glu Arg Ala Ile Asp Val Gly Thr Arg Ser Ala
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Met Pro Ile Pro Glu Asp Leu Ser Thr Thr Ser Gly Gly Gln Gln
Ser Ser Lys Ser Asp Arg Val Val Val Ile Lys Glu Glu Thr Asn
                 50
                                     55
His Ser Glu Met Ala Glu Asp Leu Cys Lys Ile Gly Ser Glu Arg
                 65
                                     70
Ser Leu Val Leu Asp Arg Leu Ala Ser Asn Val Ala Lys Arg Lys
                                     85
Ser Ser Met Pro Gln Lys Phe Leu Gly Asp Lys Gly Leu Ser Asp
                                    100
                 95
Thr Pro Tyr Asp Ser Ser Ala Ser Tyr Glu Lys Glu Asn Glu Met
                110
                                    115
Met Lys Ser His Val Met Asp Gln Ala Ile Asn Asn Ala Ile Asn
                                    130
                125
Tyr Leu Gly Ala Glu Ser Leu Arg Pro Leu Val Gln Thr Pro Pro
                                    145
                140
Gly Gly Ser Glu Val Val Pro Val Ile Ser Pro Met Tyr Gln Leu
                                    160
                155
His Lys Pro Leu Ala Glu Gly Thr Pro Arg Ser Asn His Ser Ala
                                    175
                170
Gln Asp Ser Ala Val Glu Asn Leu Leu Leu Leu Ser Lys Ala Lys
                                    190
Leu Val Pro Ser Glu Arg Glu Ala Ser Pro Ser Asn Ser Cys Gln
                200
                                    205
Asp Ser Thr Asp Thr Glu Ser Asn Asn Glu Glu Gln Arg Ser Gly
                215
                                    220
                                                         225
Leu Ile Tyr Leu Thr Asn His Ile Ala Pro His Ala Arg Asn Gly
                230
                                    235
Leu Ser Leu Lys Glu Glu His Arg Ala Tyr Asp Leu Leu Arg Ala
                                    250
                245
Ala Ser Glu Asn Ser Gln Asp Ala Leu Arg Val Val Ser Thr Ser
                                                         270
                                    265
                260
Gly Glu Gln Met Lys Val Tyr Lys Cys Glu His Cys Arg Val Leu
                275
                                    280
Phe Leu Asp His Val Met Tyr Thr Ile His Met Gly Cys His Gly
                290
                                    295
Phe Arg Asp Pro Phe Glu Cys Asn Met Cys Gly Tyr His Ser Gln
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Phe His Met Ser
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Gry	ary	vai	ALG	20	Dea	Dea	O14	U1	25		V 41			30
Va 1	Ara	Asp	LVS		Asp	Ser	Thr	Pro		Tvr	Tvr	Ala	Cvs	
•	9	nop	<i>D</i> , 0	35				• • •	40	-3-	,- -		-1-	45
Cys	Glv	His	Glu	Glu	Leu	Val	Leu	Tyr	Leu	Leu	Ala	Asn	Gly	Ala
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Arg	Cys	Glu	Ala	Asn	Thr	Phe	Asp	Gly	Glu	Arg	Суз	Leu	Tyr	Gly
				65					70					75
Ala	Leu	Ser	Asp	Pro	Ile	Arg	Arg	Ala	Leu	Arg	Asp	Tyr	Lys	Gln
				80					85					90
Val	Thr	Ala	Ser	Сув	Arg	Arg	Arg	Asp	Tyr	Tyr	Asp	Asp	Phe	
_				95					100	_	•			105
Gln	Arg	Leu	Leu		Gln	Gly	Ile	His		Asp	Val	Val	Phe	
		~1	•	110	51	•	**- 1	,,,,	115	~	17- 1	•	01	120
Val	His	GIA	Lys		Phe	Arg	Val	HIS	_	Cys	vaı	Leu	GIA	
X	C	31 0	m	125	21-	7	Met	Ť ou	130	ጥኩሎ	Tara	Т~ ~	Tvo	135
Arg	Set	AIA	ıyı	140	MIA	Wall	Mec	Dea	145	1111	цуs	ΙΙĐ	гув	150
ī.vs	Ser	Val	Val		Leu	Ara	His	Pro		T1e	Asn	Pro	Va1	
- 2, 0	001	•	,,,	155					160					165
Phe	Gly	Ala	Leu		Gln	Tyr	Leu	Tyr		Val	Ala	Ser	Lys	
	-			170		_		_	175				_	180
Gly	Thr	Сув	Val	Lys	Va1	Leu	Thr	Ile	Glu	Pro	Pro	Pro	Ala	Asp
				185					190					195
Pro	Arg	Leu	Arg	Glu	Asp	Met	Ala	Leu	Leu	Ala	Asp	Cys	Ala	Leu
				200					205					210
Pro	Pro	Glu	Leu	Arg	Gly	Asp	Leu	\mathtt{Trp}	Glu	Leu	Pro	Phe	Pro	Cys
				215					220					225
Pro	Asp	Gly	Phe		Ser	Суз	Pro	Aap		Суз	Phe	Arg	Val	
		_	_,	230	_		_		235	_,	_		_	240
Gly	Cys	Ser	Phe		Сув	His	Lys	Ala		Phe	Cys	GIĀ	Arg	
3		nh a	3	245	T	T	N ===	7	250	Dha	3	01	C	255
Asp	TAL	Pne	Arg	260	ren	Leu	Asp	иар	265	Pile	ALG	GIU	261	270
Glu	Pro	Δla	Thr		Glv	Glv	Pro	Pro		Va1	Thr	ī.eu	His	
			****	275	01,	O ₂			280					285
Ile	Ser	Pro	Asp		Phe	Thr	His	Val		Tyr	Tyr	Met	Tyr	
			•	290					295	-	•		-	300
Asp	His	Thr	Glu	Leu	Ser	Pro	Glu	Ala	Ala	Tyr	Asp	Val	Leu	Ser
				305					310					315
Val	Ala	Asp	Met	Tyr	Leu	Leu	Pro	Gly	Leu	ГЛЗ	Arg	Leu	Cys	_
				320					325					330
Arg	Ser	Leu	Ala		Met	Leu	Asp	Glu		Thr	Val	Val	Gly	
_	_	•		335	_		_	_	340	_	_		_	345
Trp	Arg	Val	Ala	_	Leu	Phe	Arg	Leu		Arg	Leu	GIu	Asp	
~	mh	C1	т	350	71 -	T	1707	T1.	355	T 1.00	T 011	1701	ω1	360
cys	inr	GIU	TAL	Met 365	wig	гÃ2	Val	116	370	гλа	rea	val	GIU	Arg 375
Gl v	Aen	Phe	Val		ء 1 ھ	Ve1	Lys	Glu		۵1ء	د ا ۵	ما لا	Va1	
SIU	นอน	1 116	141	380	u	141	د رپ		385	n_d	via	ura	V CI A	390
Ala	Ara	Gln	Glu		Asp	Ser	Ile	Pro		Va1	asa	Asp	Ile	
	5			395					400					405
Phe	His	Val	Ala		Thr	Val	Gln	Thr		Ser	Ala	Ile	Glu	
				410					415					420
Ala	Gln	Gln	Arg	Leu	Arg	Ala	Leu	G1u	Asp	Leu	Leu	Va1	Ser	Ile

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. 40/141

His Leu Thr His Glu Asn Val Gln Arg Lys Gln Ala Arg Thr Gly

Glu Glu Arg Glu Glu Glu Glu Glu Glu Gln Ile Ser Glu Ser Glu

275

290

305

280

295

310

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Ser Glu Asp Glu Glu Asn Glu Ile Ile Tyr Asn Pro Lys Asn Leu
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Pro Leu Gly Trp Asp Gly Lys Pro Ile Pro Tyr Trp Leu Tyr Lys
                                    340
Leu His Gly Leu Asn Ile Asn Tyr Asn Cys Glu Ile Cys Gly Asn
                                    355
                350
Tyr Thr Tyr Arg Gly Pro Lys Ala Phe Gln Arg His Phe Ala Glu
                                    370
                365
Trp Arg His Ala His Gly Met Arg Cys Leu Gly Ile Pro Asn Thr
                                    385
                380
Ala His Phe Ala Asn Val Thr Gln Ile Glu Asp Ala Val Ser Leu
                395
                                    400
Trp Ala Lys Leu Lys Leu Gln Lys Ala Ser Glu Arg Trp Gln Pro
                410
                                    415
Asp Thr Glu Glu Glu Tyr Glu Asp Ser Ser Gly Asn Val Val Asn
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                                   430
                                                         435
Lys Lys Thr Tyr Glu Asp Leu Lys Arg Gln Gly Leu Leu
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                                     40
Ala Lys Lys Gly Glu Lys Val Pro Lys Gly Lys Lys Gly Lys Ala
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Asp Ala Gly Lys Glu Gly Asn Asn Pro Ala Glu Asn Gly Asp Ala
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Leu	Arg	Pro	Ala	Gln 50	Arg	Ala	Leu	Tyr	Arg 55	Asp	Val	Met	Arg	Glu 60
Thr	Phe	Gly	His	Leu 65	Gly	Ala	Leu	Gly	Phe 70	Ser	Val	Pro	Lys	Pro 75
Ala	Phe	Ile	Ser	Trp 80	Val	Glu	Gly	Glu	Va1 85	Glu	Ala	Trp	Ser	Pro 90
Glu	Ala	Gln	Asp	Pro 95	Asp	Gly	Glu	Ser	Ser 100	Ala	Ala	Phe	Ser	Arg 105
Gly	Gln	Gly	Gln	Glu 110	Ala	Gly	Ser	Arg	Asp 115	Gly	Asn	Glu	Glu	Lys 120
Glu	Arg	Leu	Lys	Lys 125	Cys	Pro	Lys	Gln	Lys 130	Glu	Val	Ala	His	Glu 135
Val	Ala	Val	Lys	Glu 140	Trp	Trp	Pro	Ser	Val 145	Ala	Сув	Pro	Glu	Phe 150
Сув	Asn	Pro	Arg		Ser	Pro	Met	Asn		Trp	Leu	Lys	Asp	
Leu	Thr	Arg	Arg		Pro	His	Ser	Суѕ		Asp	Cys	Gly	Arg	
Phe	Ser	Tyr	Pro	_	Leu	Leu	Ala	Ser		G1n	Arg	Val	His	
Gly	Glu	Arg	Pro		Ser	Сув	Gly	Gln		G1n	Ala	Arg	Phe	
Gln	Arg	Arg	Tyr		Leu	Gln	His	Gln		Ile	His	Thr	Gly	
Lys	Pro	Tyr	Pro		Pro	Asp	Cys	Gly		Arg	Phe	Arg	Gln	
Gly	Ser	Leu	Ala	Ile 245	His	Arg	Arg	Ala		Thr	Gly	Glu	Lys	
Tyr	Ala	Суз	Ser	Asp 260	Сув	Lys	Ser	Arg	Phe 265	Thr	Tyr	Pro	Tyr	Leu 270
Leu	Ala	Ile	His	Gln 275	Arg	Lys	His	Thr	Gly 280	Glu	Lys	Pro	Tyr	Ser 285
Суз	Pro	Asp	Сув	Ser 290	Leu	Arg	Phe	Ala	Tyr 295	Thr	Ser	Leu	Leu	Ala 300
Ile	His	Arg	Arg	Ile 305	His	Thr	Gly	Glu	Lys 310	Pro	Tyr	Pro	Cys	
Asp	Суз	Gly	Arg	Arg 320	Phe	Thr	Tyr	Ser	Ser 325	Leu	Leu	Leu	Ser	
Arg	Arg	Ile	His		Asp	Ser	Arg	Pro		Pro	Сув	Val	Glu	
Gly	Lys	Gly	Phe	•	Arg	Lys	Thr	Ala	Leu 355	Glu	Ala	His	Arg	
Ile	His	Arg	Ser		Ser	Glu	Arg	Arg		Trp	Gln	Gln	Ala	
Val	Gly	Arg	Ser		Pro	Ile	Pro	Val		Gly	Gly	Lys	Asp	
Pro	Val	His	Phe		His	Phe	Pro	Asp		Phe	Gln	Glu	Phe	
Gln	Gln	Arg	Leu		Asp	Arg	Gly	Val		Ser	Asn	Ala	Pro	
Va1	Pro	Gly	Gln		Pro	Arg	Ser	Phe		Arg	Asp	Arg	Arg	

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Ser Gly Pro Tyr Ile Phe Leu Glu Gly Lys Lys Pro Leu Leu Tyr
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                                    460
Phe Pro Asp Thr Pro Pro Pro Pro Leu Glu Lys Ala Ala Glu Ala
                470
                                    475
Ala Leu Phe Lys Gly Lys Trp Asp Asp Glu Ala Arg Glu Met Ala
                485
                                    490
Pro Pro Pro Ala Pro Leu Leu Ala Pro Arg Pro Gly Glu Thr Arg
                500
                                    505
Pro Gly Cys Arg Lys Pro Gly Thr Val Ser Phe Ala Asp Val Ala
                515
                                    520
Val Tyr Phe Ser Pro Glu Glu Trp Gly Cys Leu Arg Pro Ala Gln
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Arg Ala Leu Tyr Arg Asp Val Met Gln Glu Thr Tyr Gly His Leu
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Gly Ala Leu Gly Phe Pro Gly Pro Lys Pro Ala Leu Ile Ser Trp
                                    565
                560
Met Glu Gln Glu Ser Glu Ala Trp Ser Pro Ala Ala Gln Asp Pro
                575
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Glu Lys Gly Glu Arg Leu Gly Gly Ala Arg Arg Gly Asp Val Pro
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Asn Arg Lys Glu Glu Glu Pro Glu Glu Val Pro Arg Ala Lys Gly
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Pro Arg Lys Ala Pro Val Lys Glu Ser Pro Glu Val Leu Val Glu
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Arg Asn Pro Asp Pro Ala Ile Ser Val Ala Pro Ala Arg Ala Gln
Pro Pro Lys Asn Ala Ala Trp Asp Pro Thr Thr Gly Ala Gln Pro
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                650
Pro Ala Pro Ile Pro Ser Met Asp Ala Gln Ala Gly Gln Arg Arg
                                    670
                665
His Val Cys Thr Asp Cys Gly Arg Arg Phe Thr Tyr Pro Ser Leu
                680
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Leu Val Ser His Arg Arg Met His Ser Gly Glu Arg Pro Phe Pro
                695
                                    700
Cys Pro Glu Cys Gly Met Arg Phe Lys Arg Lys Phe Ala Val Glu
                710
                                    715
Ala His Gln Trp Ile His Arg Ser Cys Ser Gly Gly Arg Arg Gly
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                                    730
Arg Arg Pro Gly Ile Arg Ala Val Pro Arg Ala Pro Val Arg Gly
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		•		410		_			415			•		420
Met	Lys	Val	His	G1n 425	His	Gln	Asp	Arg	G1y 430	Glu	Thr	Phe	Gln	Cys 435
Gln	Leu	Сув	Pro		Thr	Ser	Ser	Arg		Phe	Ser	Leu	Lys	
		_		440				_	445				-	450
His	Met	Arg	Cys		Gln	His	Phe	Leu	_	Thr	Glu	Ala	Lys	
Lve	G1v	G1.,	T16	455 Pro	7 an	Pro	λας	17n 1	460	61	502	Pro	ut.	465
ב עם	Giu	GIU	116	470	ASD	PIO	ASD	Val	475	GIY	261	PIO	nis	480
Ser	Asp	Ser	Ala	Cys	Leu	Gly	Gln	Gln	Arg	Glu	Gly	Gly	Gly	
01	•		01 .	485			1	_	490		_		_	495
GIU	Leu	Val	GIA	500	Met	Met	Thr	Ser	Asn 505	Thr	Pro	Glu	Arg	Thr 510
Ser	Gln	Gly	Gly		Gly	Val	Ser	Pro		Leu	Val	Lys	Glu	
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Pro	Lys	Glu	Asp	Asn 530	Gly	Leu	Pro	Thr		Phe	Thr	Leu	Asn	
Ala	Asp	Ara	Pro		Asn	His	Thr	Lvs	535 Leu	Lvs	Asp	Pro	Ser	540 Glu
				545					550					555
Tyr	Val	Ala	Asn		Ala	Ser	Ala	Leu		Ser	Gln	Asp	Ile	
Va1	Lvs	Met	λla	560 Ser	Asp	Phe	ī.eu	Met	565	Len	Sor	Ala	λl ₌	570
	-,-			575			204		580	Dea	561	nia	AIG	585
Gln	Lys	Glu	Pro		Asn	Leu	Asn	Phe		Val	Lys	Glu	Glu	Pro
Lve	Glu	Gly	Q1	590	Len	502	Th~		595	Dwa	7	Ser	C	600
nys	GIU	GIY	GIU	605	Dea	261	1111	1111	610	PIO	Arg	Ser	Ser	615
Val	Phe	Ser	Pro	Glu	Ser	Glu	Val	Ser	Ala	Pro	Gly	Val	Ser	
3		T	T	620	O1	a1	01		625			_	_	630
Asp	Ala	Leu	гуѕ	635	GIN	GIU	GIĀ	гλε	640	Ser	Val	Leu	Arg	Arg 645
Asp	Val	Ser	Val		Ala	Ala	Ser	Glu		Leu	Met	Lys	Leu	
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Ala	Glu	Ser	Tyr	Lys 665	Glu	Thr	Gln	Met	Val 670	Lys	Ile	Lys	Glu	Glu 675
Pro	Met	Glu	Val		Ile	Gln	Asp	Ser		Val	Ser	Ile	Ser	
_		_		680					685					690
Ser	Arg	Asn	Val	Gly 695	Tyr	Ser	Thr	Leu	11e	Gly	Arg	Glu	Lys	
Glu	Pro	Leu	Gln		Met	Pro	G1u	Gly		Val	Pro	Pro	Glu	705 Ara
				710					715					720
Asn	Leu	Phe	Ser		Asp	Ile	Ser	Val		Met	Ala	Ser	Glu	
Leu	Phe	Gln	Leu	725 Ser	Glu	Lvs	Val	Ser	730 Lvs	Glu	His	Asn	His	735
				740		-3-			745					750
Lys	Glu	Asn	Thr		Arg	Thr	Thr	Thr		Pro	Phe	Phe	Ser	Glu
λen	Th-	Dho	A = -	755	50-	Dwa	Dho	mb	760	3 ~~	C	Lys	01	765
veħ	1111	FIIG	ALG	770	ser	PIO	Pne	Inr	775	Asn	ser	гÀз	GIU	180
Leu	Pro	Ser	Asp	Ser	Val	Leu	His	Gly	Arg	Ile	Ser	Ala	Pro	
m\	01	•	T1 -	785				_,	790	~•	_	_		795
ınr	GIU	гЛЗ	TTE	800	ren	GIu	Ala	GTA	Asn 805	GIA	Leu	Pro	Ser	Trp 810
Lys	Phe	Asn	Asp		Leu	Phe	Pro	Суз		Val	Cys	Gly	Lys	
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Phe	Gly	Arg	Gln	Gln	Thr	Leu	Ser	Arg	His	Leu	Ser	Leu	His	Thr

835

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Cys Arg Ala Asn Leu Asn Gln His Leu Thr Val His Ser Val Lys
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Leu Val Ser Thr Asp Thr Glu Asp Ile Val Ser Ala Val Thr Ser
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Glu Gly Ser Asp Gly Lys Lys His Pro Tyr Tyr Tyr Ser Cys His
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                                    895
Val Cys Gly Phe Glu Thr Glu Leu Asn Val Gln Phe Val Ser His
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                                    910
Met Ser Leu His Val Asp Lys Glu Gln Trp Met Phe Ser Ile Cys
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                                    925
Cys Thr Ala Cys Asp Phe Val Thr Met Glu Glu Ala Glu Ile Lys
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Thr His Ile Gly Thr Lys His Thr Gly Glu Asp Arg Lys Thr Pro
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                950
Ser Glu Ser Asn Ser Pro Ser Ser Ser Ser Leu Ser Ala Leu Ser
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Asp Ser Ala Asn Ser Lys Asp Asp Ser Asp Gly Ser Gln Lys Asn
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Lys Gly Gly Asn Asn Leu Leu Val Ile Ser Val Met Pro Gly Ser
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Gln Pro Ser Leu Asn Ser Glu Glu Lys Pro Glu Lys Gly Phe Glu
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                                   1015
Cys Val Phe Cys Asn Phe Val Cys Lys Thr Lys Asn Met Phe Glu
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Arg His Leu Gln Ile His Leu Ile Thr Arg Met Phe Glu Cys Asp
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                                   1045
Val Cys His Lys Phe Met Lys Thr Pro Glu Gln Leu Leu Glu His
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                                     40
Val Leu Ile Asp Cys Pro His Pro Asn Cys Asn Lys Lys Tyr Lys
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His Ile Asn Gly Leu Arg Tyr His Gln Ala His Ala His Leu Asp
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                                     70
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Ser	Asp	Сув	Glu	Glu 95	Gly	Leu	Ser	Asn	Val 100	Ala	Leu	Glu	Сув	Ser 105
Glu	Pro	Ser	Thr	Ser 110	Val	Ser	Ala	Tyr	Asp 115	Gln	Leu	Lys	Ala	Pro 120
Ala	Ser	Pro	Gly	Ala 125	Gly	Asn	Pro	Pro	Gly 130	Thr	Pro	Lys	Gly	Lys 135
Arg	Glu	Leu	Met	Ser 140	Asn	Gly	Pro	Gly	Ser 145	Ile	Ile	Gly	Ala	Lys 150
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				170					175				qaA	180
				185					190				Lys	195
				200					205				Lys	210
_	_	-	_	215			_	_	220				Leu	225
				230					235				Pro	240
				245					250				Thr	255
		_		260					265				Thr	270
				275					280				Gln	285
				290					295				Ala	300
				305					310				Leu	315
				320					325				Ala	330
	_	_		335	_				340				Leu	345
				350					355				Leu	360
	_			365					370				Ile	375
			_	380		_			385	_			Pro	390
				395					400				Leu	405
				410					415				Gln	420
				425			-		430				_	Ser 435
				440					445				Arg	450
				455					460				Ile	465
				470					475				Ala	480
Ser	ser	GIN	Leu	Lys 485	GIu	ser	HIS	ser	Pro 490	туr	чуr	Hls	ser	Tyr 495

Asp	Pro	Tyr	Tyr	Ser 500	Pro	Ser	Tyr	Met	His 505	Pro	Gly	Gln	Val	Gly 510
Ala	Pro	Ala	Ala	Gly 515	Asn	Ser	Gly	Ser	Thr 520	Gln	Gly	Met	Lys	Ile 525
Lys	Lys	Glu	Ser	Glu 530	Glu	Asp	Ala	Glu	Lys 535	Lys	Asp	Lys	Ala	Glu 540
Gln	Leu	Asp	Ser	Lys 545	Lys	Val	Asp	His	Asn 550	Ser	Ala	Ser	Leu	Gln 555
Pro	Gln	His	Gln	Ser 560	Val	Ile	Thr	Gln	Arg 565	His	Pro	Ala	Leu	Ala 570
Gln	Ser	Leu	Tyr	Tyr 575	Gly	Gln	Tyr	Ala	Tyr 580	Gly	Leu	Tyr	Met	Asp 585
Gln	Lys	Ser	Leu	Met 590	Ala	Thr	Ser	Pro	Ala 595	Tyr	Arg	Gln	Gln	Tyr 600
Glu	Lys	Tyr	Tyr	Glu 605	Asp	Gln	Arg	Leu	Ala 610	Glu	Gln	Lys	Met	Ala 615
Gln	Thr	Gly	Arg	Gly 620	Asp	Cys	Glu	Arg	Lys 625	Ser	Glu	Leu	Pro	Leu 630
Lys	Glu	Leu	Gly	Lys 635	Glu	Glu	Thr	Lys	Gln 640	Lys	Asn	Met	Pro	Ser 645
Ala	Thr	Ile	Ser	Lys 650	Ala	Pro	Ser	Thr	Pro 655	Glu	Pro	Asn	Lys	Asn 660
His	Ser	Lys	Leu	Gly 665	Pro	Ser	Val	Pro	Asn 670	Ľув	Thr	Glu	Glu	Thr 675
Gly	Lys	Ser	Gln	Leu 680	Leu	Ser	Asn	His	Gln 685	Gln	Gln	Leu	Gln	Ala 690
Asp	Ser	Phe	Lys	Ala 695	Lys	Gln	Met	Glu	Asn 700	His	Gln	Leu	Ile	Lys 705
Glu	Ala	Val	Glu	Met 710	Lys	Ser	Val	Met	Asp 715	Ser	Met	Lys	Gln	Thr 720
Gly	Val	Asp	Pro	Thr 725	Ser	Arg	Phe	Lys	Gln 730	Asp	Pro	Asp	Ser	Arg 735
Thr	Trp	His	His	Tyr 740	Val	Tyr	Gln	Pro	Lys 745	Tyr	Leu	Ąsp	Gln	Gln 750
Lys	Ser	Glu	Glu	Leu 755	Asp	Arg	Glu	Lys	Lys 760	Leu	Lys	Glu	Asp	Ser 765
Pro	Arg	Lys	Thr	Pro 770	Asn	Lys	Glu	Ser	Gly 775	Val	Pro	Ser	Leu	Pro 780
Val	Ser	Leu	Thr	Ser 785	Ile	Lys	Glu	Glu	Pro 790	Lys	Glu	Ala	Lys	His 795
Pro	Asp	Ser	Gln	Ser 800	Met	Glu	Glu	Ser	Lys 805	Leu	Lys	Asn	Asp	Asp 810
Arg	Lys	Thr	Pro	Val 815	Asn	Trp	Lys	Asp	Ser 820	Arg	Gly	Thr	Arg	Val 825
Ala	Val	Ser	Ser	Pro 830	Met	Ser	Gln	His	Gln 835	Ser	Tyr	Ile	Gln	Tyr 840
Leu	His	Ala	Tyr	Pro 845	Tyr	Pro	Gln	Met	Tyr 850	Asp	Pro	Ser	His	Pro 855
Ala	Tyr	Arg	Ala	Val 860	Ser	Pro	Val	Leu	Met 865	His	Ser	Tyr	Pro	Gly 870
Ala	Tyr	Leu	Ser	Pro 875	Gly	Phe	His	Tyr	Pro 880	Val	Tyr	Gly	Lys	Met 885
Ser	Gly	Arg	Glu	Glu 890	Thr	Glu	Lys	Val		Thr	Ser	Pro	Ser	Val 900
Asn	Thr	Lys	Thr	Thr 905	Thr	G1u	Ser	Lys	Ala 910	Leu	Asp	Leu	Leu	Gln 915

Gln His Ala Asn Gln Tyr Arg Ser Lys Ser Pro Ala Pro Val Glu 925 920 Lys Ala Thr Ala Glu Arg Glu Arg Glu Ala Glu Arg Glu Arg Asp 940 935 Arg His Ser Pro Phe Gly Gln Arg His Leu His Thr His His His 955 950 Thr His Val Gly Met Gly Tyr Pro Leu Ile Pro Gly Gln Tyr Asp 970 965 Pro Phe Gln Gly Leu Thr Ser Ala Ala Leu Val Ala Ser Gln Gln 980 985 Val Ala Ala Gln Ala Ser Ala Ser Gly Met Phe Pro Gly Gln Arg 1000 1005 Arg Glu <210> 32 <211> 511 <212> PRT <213> Homo sapiens <220> <221> misc_feature <223> Incyte ID No: 4753527CD1 <400> 32 Met Ala Ala Ala Ile Thr Arg His Gly Arg Pro Gly Gly Ala Leu Pro Pro Glu Pro Ser Ala Pro Arg Gln Pro Gly Phe Gly Gly Arg Gly Arg Ala Glu Pro Pro Glu Ala Glu Ala Glu Ala Val Ala Ala Leu Leu Leu Asn Gly Gly Ser Gly Gly Gly Gly Gly Gly Gly Gly Gly Val Gly 65 Gly Gly Glu Ala Glu Thr Met Ser Glu Pro Ser Pro Glu Ser Ala 80 85 Ser Gln Ala Gly Glu Asp Glu Asp Glu Glu Glu Asp Asp Glu Glu 100 Glu Glu Asp Glu Ser Ser Ser Gly Gly Glu Glu Glu Ser 110 115 Ser Ala Glu Ser Leu Val Gly Ser Ser Gly Gly Ser Ser Ser Asp 130 125 Glu Thr Arg Ser Leu Ser Pro Gly Ala Ala Ser Ser Ser Ser Gly 140 145 Asp Gly Asp Gly Lys Glu Gly Leu Glu Glu Pro Lys Gly Pro Arg 155 160 Gly Ser Gln Gly Gly Gly Gly Gly Ser Ser Ser Ser Val 170 175 Val Ser Ser Gly Gly Asp Glu Gly Tyr Gly Thr Gly Gly Gly 190 185 Ser Ser Ala Thr Ser Gly Gly Arg Arg Gly Ser Leu Glu Met Ser 200 205 Ser Asp Gly Glu Pro Leu Ser Arg Met Asp Ser Glu Asp Ser Ile 220

Ser Ser Thr Ile Met Asp Val Asp Ser Thr Ile Ser Ser Gly Arg

235

Ser Thr Pro Ala Met Met Asn Gly Gln Gly Ser Thr Thr Ser Ser

230

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Cys Phe Asn Ser Ser Pro Asp Leu Ala Asp His Ile Arg Ser Ile
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His Val Asp Gly Gln Arg Gly Gly Val Phe Val Cys Leu Trp Lys
                290
                                    295
                                                        300
Gly Cys Lys Val Tyr Asn Thr Pro Ser Thr Ser Gln Ser Trp Leu
                305
                                    310
Gln Arg His Met Leu Thr His Ser Gly Asp Lys Pro Phe Lys Cys
                320
                                    325
Val Val Gly Gly Cys Asn Ala Ser Phe Ala Ser Gln Gly Gly Leu
                335
                                    340
Ala Arg His Val Pro Thr His Phe Ser Gln Gln Asn Ser Ser Lys
                350
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Val Ser Ser Gln Pro Lys Ala Lys Glu Glu Ser Pro Ser Lys Ala
                365
                                    370
Gly Met Asn Lys Arg Arg Lys Leu Lys Asn Lys Arg Arg Arg Ser
                380
                                    385
Leu Pro Arg Pro His Asp Phe Phe Asp Ala Gln Thr Leu Asp Ala
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Ile Arg His Arg Ala Ile Cys Phe Asn Leu Ser Ala His Ile Glu
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Ser Leu Gly Lys Gly His Ser Val Val Phe His Ser Thr Val Ile
                425
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Ala Lys Arg Lys Glu Asp Ser Gly Lys Ile Lys Leu Leu His
                                    445
Trp Met Pro Glu Asp Ile Leu Pro Asp Val Trp Val Asn Glu Ser
                455
                                    460
Glu Arg His Gln Leu Lys Thr Lys Val Val His Leu Ser Lys Leu
                470
                                    475
Pro Lys Asp Thr Ala Leu Leu Leu Asp Pro Asn Ile Tyr Arg Thr
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50/141

Met Lys Asn Ala Thr Ile Val Met Ser Val Arg Arg Glu Gln Gly

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Gly Phe Thr Arg Glu Glu Trp Gln Phe Leu Asp Gln Ser Gln Lys

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35

10

25

40

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Glu	Gln	Gly	Glu	Pro 80	Pro	Gly	Ile	Ala	G1u 85	Gly	Ala	Ala	His	Ser 90
Gln	Ile	Cys	Pro	Gly 95	Phe	Va1	Ile	Gln	Ser 100	Arg	Arg	Tyr	Ala	Gly 105
				110		Gly			115					120
Ile	Lys	Arg	Asp	Lув 125	Thr	Leu	Thr	Gly	Val 130	Lys	Tyr	His	Arg	Cys 135
	_			140		Lys			145		_			150
	_		_	155	_	Pro			160				_	165
				170		Gln			175					180
				185		Gly			190					195
	_	_		200		Thr			205	_				210
				215		Ser			220					225
-				230		His			235			_		240
	-			245		Сув			250					255
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				275		Lys			280					285
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	_	_		305		Phe	_		310		_			315
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				335		Ser			340					345
				350		Arg			355					360
				365					370					Thr 375
				380		Ile			385				_	390
				395		Ser			400					405
				410		Thr			415					420
				425		Asn			430					435
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Val	val	ASN	GIN	G1u 455	rne	Glu	Gin	Arg	11e 460	Ser	Leu	Thr	Asn	G1u 465

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Lys	Gly	ГЛа	Lys		Ser	Ala	His	Ser		Gly	Gln	Arg	Gly	Arg
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гла	Pro	GIA	GIA	Gly	Arg	Asn	Pro	GIA	340	Thr	vaı	ser	Ala	
C	Dwa	Dha	D	335	~1	C	nh.	C		mb	Dwa	ω1	C	345
ser	PIO	Pne	Pro	Gln 350	GIY	ser	Pne	Ser	355	THE	PIO	сту	Ser	360
Tare	Sor	Sor	Sor	Gly	Sar	Sor	va1	aln		Dro	Gln.	Aen	Dha	
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Ser	Phe	Thr	αaA	Ser	qsA	Leu	Ara	Asn		Ser	Tvr	Ser	His	
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Gln	Gln	Ser	Ser	Ala	Thr	Lys	qeA	Val	His	Lys	Gly	Glu	Ser	Gly
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Ser	Gln	Glu	Gly	Gly	Val	Asn	Ser	Phe		Thr	Leu	Ile	Gly	Leu
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Pro	Ser	Thr	Ser	Ala 425	Val	Thr	Ser	GIn		Lys	Ser	Phe	GIu	
Ser	Pro	alv	Aan	Leu	alv	λan	Ser	Sar	430	Dro	Th-	71-	Gly	435
501	110	017	nap	440	Oly	asii	501	Der	445	110	1111	210	GLY	450
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Lys	Lys	Arg	Lys	Gly	Asn	Lys	Gln	Ser	Lys	His	Gly	Pro	Gly	Arg
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Pro	Lys	Gly	Asn	Lys	Asn	Gln	Glu	Asn		Ser	His	Leu	Ser	
	_		_	485	— 1	_	_		490	_				495
Ser	ser	Ala	Ser	Pro 500	Thr	Ser	Ser	Val	A1a 505	Ser	Ala	Ala	GIĀ	
Tle	ፖስተ	Ser	Ser	Ser	T.e.11	Gln	Lare	Ser		Thr	T.ou	Len	Ara	510
110	****	Jei	261	515	Беи	GIM	шуъ	261	520	1111	Leu	red	ALG	525
Gly	Ser	Leu	Gln	Ser	Leu	Ser	Val	Gly		Ser	Pro	Val	Gly	
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Glu	Ile	Ser	Met	Gln	Tyr	Arg	His	Asp	Gly	Ala	Cys	Pro	Thr	Thr
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Thr	Phe	Ser	Glu	Leu	Leu	Asn	Ala	Ile		Asn	Gly	Ile	Tyr	
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Ser	Asn	Авр	Val	Ala 575	vaı	ser	Pne	Pro	Asn 580	vaı	Val	Ser	GIÀ	585
G1v	Ser	Ser	Thr	Pro	Va1	Ser	Ser	Ser		I.eu	Pro	Gln	Gln	
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Ser	Ser	Ala	Ala	Pro	Ala	Val	Ala	Thr	Thr	Gln	Ala	Asn	Thr	Leu
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Ser	Gly	Ser	Ser	Leu	Ser	Gln	Ala	Pro		His	Met	Tyr	Gly	
N	C	3	C	635	W-5	.1.	31.	•	640		- 1-		~ 1	645
Arg	ser	ASI	ser	Ser 650	Mec	AIA	Ата	Leu	655	Ala	GIN	ser	GIU	660
Asn	Gln	ጥከታ	Δsp	Gln	Δen	T.em	Glv	Agn		Sar	λνα	y an	Lou	
			1105	665	,,,p	204	OLY	uab	670	Jei	ALY	NOIL	Dea	675
Gly	Arg	Gly	Ser	Ser	Pro	Arg	Gly	Ser		Ser	Pro	Arg	Ser	
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Val	Ser	Ser	Leu	Gln	Ile	Arg	Tyr	Asp		Pro	Gly	Asn	Ser	Ser
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Leu	Glu	Asn	Leu	Pro	Pro	Val	Ala	Ala		Ile	Glu	Gln	Leu	
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GIU	vra	GIII	ттb	Ser	GIU	GTA	GTU	GIU	rne	reu	ьeu	GIU	GID	GTA

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Ala Lys Lys Glu Arg Leu Gln Leu Leu Asn Ala Gln Leu Ser Val
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Pro Phe Pro Thr Ile Thr Ala Asn Pro Ser Pro Ser His Gln Ile
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His Thr Phe Ser Ala Gln Thr Ala Pro Thr Thr Asp Ser Leu Asn
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Thr Ser Thr Ile Pro Ala Val Ser Ala Val Gly Gly Ile Ile Gly
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Ala Leu Pro Gly Asn Gln Leu Ala Ile Asn Gly Ile Val Gly Ala
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Leu Asn Gly Val Met Gln Thr Pro Val Thr Met Ser Gln Asn Pro
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                                    925
Thr Pro Leu Thr His Thr Thr Val Pro Pro Asn Ala Thr His Pro
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Met Pro Ala Thr Leu Thr Asn Ser Ala Ser Gly Leu Gly Leu Leu
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Ser Asp Gln Gln Arg Gln Ile Leu Ile His Gln Gln Gln Phe Gln
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Gln Leu Leu Asn Ser Gln Gln Leu Thr Pro Val His Arg His Pro
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Asp	Thr	Gly	Gly	Ala 95	Asp	His	Ser	Сув	Ser 100	Ile	Сув	Gly	Lys	Ser 105
Leu	Ser	Ser	Ala	Ser 110	Ser	Leu	Asp	Arg	His 115	Met	Leu	Va1	His	Ser 120
Gly	Glu	Arg	Pro	Tyr 125	Lys	Cys	Thr	Val	Cys 130	Gly	Gln	Ser	Phe	Thr 135
Thr	Asn	Gly	Asn	Met 140	His	Arg	His	Met	Lys 145	Ile	His	Glu	Lys	Asp 150
Pro	Asn	Ser	Ala	Thr 155	Ala	Thr	Ala	Pro	Pro 160	Ser	Pro	Leu	Lys	Arg 165
Arg	Arg	Leu	Ser	Ser 170	Lys	Arg	Lys	Leu	Ser 175	His	Asp	Ala	Glu	Ser 180
Glu	Arg	Glu	Asp	Pro 185	Ala	Pro	Ala	Lys	Lys 190	Met	Val	Glu	Asp	Gly 195
Gln	Ser	Gly	Asp	Leu 200	Glu	Lys	Lys	Ala	Asp 205	Glu	Val	Phe	His	Суs 210
Pro	Val	Сув	Phe	Lys 215	Glu	Phe	Val	Суз	Lys 220	Tyr	Gly	Leu	Glu	Thr 225
His	Met	Glu	Thr	His 230	Ser	Asp	Asn	Pro	Leu 235	Arg	СЛа	Asp	Ile	Cys 240
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qaƙ	Ser	Ser	Ile	Va1 455	Val	Lys	Pro	Ile	Ser 460	Gly	Glu	Ser	Ala	11e 465

Glu	Leu	Ala	Asp	Ile 470	Gln	Gln	Ile	Leu	Lys 475	Met	Ala	Ala	Ser	Ala 480
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	Leu			515	•				520					525
	Ser			530					535		-			540
	Ala			545					550					555
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	Arg			575					580					585
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	Ile			605					610					615
	Glu			620					625					630
	Gly			635					640					645
	Phe			650					655					660
	Val			665					670					675
	Asp			680					685					690
	Thr			695					700					705
	Pro			710					715					720
	His		_	725				_	730					735
	Val			740					745					750
	Asp			755					760					765
	Ala			770					775					780
	Gly	GIV	His	T.va	വിച	7 ~~	•						01	Cve
Ser				785					Phe 790					795
	Ala	Ala	Phe	785 Ala 800	Ala	Lys	Arg	Asn	790 Cys 805	Ile	His	His	Ile	795 Leu 810
Lys	Ala Gln	Ala His	Phe Leu	785 Ala 800 His 815	Ala Val	Lys Pro	Arg Glu	Asn Gln	790 Cys 805 Asp 820	Ile Ile	His Glu	His Ser	Ile Tyr	795 Leu 810 Val 825
Lys Leu	Ala Gln Ala	Ala His Ala	Phe Leu Asp	785 Ala 800 His 815 Gly 830	Ala Val Leu	Lys Pro Gly	Arg Glu Pro	Asn Gln Ala	790 Cys 805 Asp 820 Glu 835	Ile Ile Ala	His Glu Pro	His Ser Ala	Ile Tyr Ala	795 Leu 810 Val 825 Glu 840
Lys Leu Ala	Ala Gln Ala Ser	Ala His Ala Gly	Phe Leu Asp Arg	785 Ala 800 His 815 Gly 830 Gly 845	Ala Val Leu Glu	Lys Pro Gly Asp	Arg Glu Pro Ser	Asn Gln Ala Gly	790 Cys 805 Asp 820 Glu 835 Cys 850	Ile Ile Ala Ala	His Glu Pro Ala	His Ser Ala Leu	Ile Tyr Ala Gly	795 Leu 810 Val 825 Glu 840 Asp 855
Lys Leu Ala Cys	Ala Gln Ala	Ala His Ala Gly Pro	Phe Leu Asp Arg Leu	785 Ala 800 His 815 Gly 830 Gly 845 Thr 860	Ala Val Leu Glu Ala	Lys Pro Gly Asp Phe	Arg Glu Pro Ser Leu	Asn Gln Ala Gly Glu	790 Cys 805 Asp 820 Glu 835 Cys 850 Pro 865	Ile Ile Ala Ala Gln	His Glu Pro Ala Asn	His Ser Ala Leu Gly	Ile Tyr Ala Gly Phe	795 Leu 810 Val 825 Glu 840 Asp 855 Leu 870

Leu	Glu	Pro	Ala Sei		Phe	Ala	Val	Asp 895	Phe	Asn	Glu	Pro Leu 900
Asp	Phe	Ser		Gly	Leu	Ala	Leu	Val 910	Gln	Val	Lya	Gln Glu 915
Asn	Ile	Ser	Phe Let		Pro	Ser	Ser	Leu 925	Val	Pro	Tyr	Asp Cys 930
Ser	Met	Glu	Pro I16		Leu	Ser	Ile	Pro 940	Lys	Asn	Phe	Arg Lys 945
Gly	Asp	Lys	Asp Let 950		Thr	Pro	Ser	G1u 955	Ala	Lys	Lys	Pro Glu 960
Glu	Glu	Ala	Gly Ser		Glu	Gln	Pro	Ser 970	Pro	Сув	Pro	Ala Pro 975
Gly	Pro	Ser	Leu Pro		Thr	Leu	Gly	Pro 985	Ser	Gly	Ile	Leu Glu 990
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			101)			1	1015				Tyr Ser 1020
			1029	5			1	1030				Ala Leu 1035
			104)			1	1045				Pro Pro 1050
			105	5			1	L060				Pro Leu 1065
Ala	Ser	Ile	Ala Gli 1070		Ile	Ser		Val 1075	Ser	Ser	Ala	Pro Thr 1080
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			110)			1	105				Pro Lys 1110
			111	5			1	120				Lys Glu 1125
			1130)			1	135				Ala Ser 1140
			114	5			1	150				Gly Arg 1155
			116)			1	165				Gly Val 1170
			117	5			1	180				Met Leu 1185
			1190)			1	1195				Thr Ala 1200
			120	5			1	1210				Asp His 1215
			122)			1	L 225				Asp Lys 1230
			123	5			1	240				Gln Lys 1245
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			126	5			1	L270				Asp Ala 1275
			128)			1	L285				Thr Ser 1290
Arg	Asp	Arg	Glu Gl: 129		Ser	Glu		Ala 300	Thr	Glu	Leu	Arg Gln 1305

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Arg Lys Arg Thr Arg Arg Ala Thr Arg Ala Trp Thr Trp Thr Ser
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Pro Pro Ser Ser Trp Thr Ser Ser Trp Arg Arg Ala Thr Arg Arg
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Arg Lys Ala His Gly Arg Pro Gly Gly Pro Arg Thr Arg Arg Glu
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              1475
Glu Lys Ser Asp Asp Lys Lys Pro Lys Thr Asp Ser Pro Lys
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              1490
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Ser Gly Asn Asn Ala Val Ser Glu Asn Glu Ala Glu Leu Ala Pro
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Thr Glu Lys Leu Gly Val Ser Ala Pro Thr Ile Pro Val Arg Arg
                                    445
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Arg Leu Ala Trp Asp Thr Glu Asn Thr Ser Glu Asp Val Gln Lys
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Gln Pro Gly Glu Lys Glu Glu Glu Asp Asp Asn Glu Glu Glu Gly
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Asp Arg Lys Thr Gly Lys Gln Ala Phe Met Gly Glu Gln Glu Lys
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Leu Asp Val His Glu Lys Ser Lys Ala Asp Lys Met Lys Glu Gly
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Ser Asp Ser Ser Val Ser Ser Glu Lys Gly Gly Arg Leu Pro Thr
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Leu Gln Leu Pro Gln His Glu Ala Phe Asn Asp Glu Asp Glu Asp
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Arg Leu Ser Glu Ile Ser Ala Arg Ser Ala Ala Ser Ser Leu Arg
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				125					130				Ser	135
				140					145				Asp	150
				155					160				Glu	165
				170					175				Asp	180
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				200					205				Arg	210
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Gly	Val	His	Leu	Asn 230	Arg	Leu	Gln	Phe	Gln 235	Gln	Gln	Gln	Asn	Ser 240
Ile	His	Ala	Ala	Lys 245	Gln	Leu	Asp	Met	Gln 250	Ser	Ser	Trp	Val	Tyr 255
G1u	Thr	Gly	Arg	Leu 260	Cys	Glu	Pro	Glu	Val 265	Leu	Asn	Ser	Leu	Glu 270
Glu	Thr	Tyr	Ser	Pro 275	Phe	Phe	Arg	Asn	Asn 280	Ser	Glu	Lys	Met	Ser 285
Met	Glu	qaA	Glu	Asn 290	Phe	Arg	Lys	Arg	Lys 295	Leu	Pro	Val	Val	Ser 300
Ser	Val	Val	Lys	Val 305	Lys	Lys	Phe	Asn	His 310	Asp	Gly	Glu	Glu	Glu 315
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Pro	Ser	Lys	Gln	Ala 350	Asn	Lys	Asn	Leu	Ile 355	Leu	Lys	Ala	Ile	Ser 360
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Pro	Gln	Lys	Gln	Thr 380	Leu	Pro	Val	Ala	Pro 385	Arg	Thr	Arg	Thr	Ser 390
Gln	Glu	Glu	Leu	Leu 395	Ala	Glu	Val	Va1	Gln 400	Gly	Gln	Ser	Arg	Thr 405
Pro	Arg	Ile	Ser	Pro	Pro	Ile	Lys	G1u	Glu	Glu	Thr	Lys	Gly	Asp

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415
                410
Ser Val Glu Lys Asn Gln Gly Thr Gln Gln Arg Gln Leu Leu Ser
                                    430
Arg Leu Gln Ile Asp Pro Val Met Ala Glu Thr Leu Gln Met Ser
                                    445
                440
Gln Ala Glu Met Ser Glu Leu Ser Val Ala Gln Lys Pro Glu Lys
                                    460
                455
Leu Leu Glu Arg Cys Lys Tyr Trp Pro Ala Cys Lys Asn Gly Asp
                                    475
                470
Glu Cys Ala Tyr His His Pro Ile Ser Pro Cys Lys Ala Phe Pro
                485
                                    490
Asn Cys Lys Phe Ala Glu Lys Cys Leu Phe Val His Pro Asn Cys
                                    505
                500
Lys Tyr Asp Ala Lys Cys Thr Lys Pro Asp Cys Pro Phe Thr His
                                    520
                515
Val Ser Arg Arg Ile Pro Val Leu Ser Pro Lys Pro Ala Val Ala
                530
                                    535
Pro Pro Ala Pro Pro Ser Ser Ser Gln Leu Cys Arg Tyr Phe Pro
                                    550
                545
Ala Cys Lys Lys Met Glu Cys Pro Phe Tyr His Pro Lys His Cys
                560
                                    565
Arg Phe Asn Thr Gln Cys Thr Arg Pro Asp Cys Thr Phe Tyr His
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Pro Thr Ile Asn Val Pro Pro Arg His Ala Leu Lys Trp Ile Arg
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Pro Gln Thr Ser Glu
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Gln Glu Val Ile Trp Gln Gly Glu Ala Lys Glu Glu Lys Lys Ala
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Val Ser Lys Asp Gly Thr Ser Asp Val Pro Ala Glu Ile Cys Val
                                      40
                 35
Val Ile Gly Gly Val Arg Asn Gln Gln Thr Leu Gly Ser Tyr Glu
                 50
                                     55
Cys Gly Ile Cys Gly Lys Lys Tyr Lys Tyr Tyr Asn Cys Phe Gln
                                      70
                 65
Thr His Val Arg Ala His Arg Asp Thr Glu Ala Thr Ser Gly Glu
                                      85
                 80
Gly Ala Ser Gln Ser Asn Asn Phe Arg Tyr Thr Cys Asp Ile Cys
                                     100
                 95
Gly Lys Lys Tyr Lys Tyr Tyr Ser Cys Phe Gln Glu His Arg Asp
                                     115
Leu His Ala Val Asp Val Phe Ser Val Glu Gly Ala Pro Glu Asn
                 125
                                     130
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Arg	Ala	Asp	Pro	Phe 140	Asp	Gln	Gly	Val	Val 145	Ala	Thr	Asp	Glu	Val 150
Гуз	Glu	Glu	Pro	Pro 155	Glu	Pro	Phe	Gln	Lys 160	Ile	Gly	Pro	Lys	Thr 165
_		-		170					175				Tyr	180
		_		185					190				Ser	195
				200					205				Ser	210
_				215					220				Ala	225
				230			•		235				Ser	240
	_			245					250				Thr	255
				260					265				Ser	270
				275					280				Lys	285
				290					295				Ser	300
				305					310				Phe	315
				320					325				Asn	330
				335					340				Val	345
				350					355				Asn	360
				365					370				Glu	375
				380					385				Сув	390
				395					400				Leu	405
				410					415				Ala	420
				425					430				Leu	435
				440					445				Thr	450
				455					460					His 465
				470					475				Ala	480
				485			-		490					Lys 495
				500					505				Trp	510
				515					520				Thr	525
				530					535				Lys	540
ЭŢΫ	nec	cys	rea	Arg 545	cys	ser	WIG	cys	550	ьeu	NEN	ser	AGT	Val 555

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Gln Ser Val Leu Ser Lys Arg Thr Leu Gln Ala Arg Ser Met His
Glu Val Ile Glu Leu Leu Asn Val Cys Glu Asp Leu Ala Gly Ser
                575
Thr Gly Leu Ala Lys Glu Thr Phe Gly Ser Leu Glu Glu Thr Ser
                                    595
                590
Pro Pro Pro Cys Trp Asn Ser Val Thr Asp Ser Leu Leu Val
                                    610
                                                         615
                605
His Glu Arg Tyr Glu Gln Ile Cys Glu Phe Tyr Ser Arg Ala Lys
                                    625
                620
Lys Met Asn Leu Ile Gln Ser Leu Asn Lys His Leu Leu Ser Asn
                                    640
                635
Leu Ala Ala Ile Leu Thr Pro Val Lys Gln Ala Val Ile Glu Leu
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                                    655
Ser Asn Glu Ser Gln Pro Thr Leu Gln Leu Val Leu Pro Thr Tyr
                                    670
                665
Val Arg Leu Glu Lys Leu Phe Thr Ala Lys Ala Asn Asp Ala Gly
                                    685
                680
Thr Val Ser Lys Leu Cys His Leu Phe Leu Glu Ala Leu Lys Glu
                                    700
                695
Asn Phe Lys Val His Pro Ala His Lys Val Ala Met Ile Leu Asp
                                    715
                710
Pro Gln Gln Lys Leu Arg Pro Val Pro Pro Tyr Gln His Glu Glu
                725
                                    730
Ile Ile Gly Lys Val Cys Glu Leu Ile Asn Glu Val Lys Glu Ser
                740
                                    745
Trp Ala Glu Glu Ala Asp Phe Glu Pro Ala Ala Lys Lys Pro Arg
                755
                                     760
Ser Ala Ala Val Glu Asn Pro Ala Ala Gln Glu Asp Asp Arg Leu
                770
                                     775
Gly Lys Asn Glu Val Tyr Asp Tyr Leu Gln Glu Pro Leu Phe Gln
                                    790
                                                         795
                785
Ala Thr Pro Asp Leu Phe Gln Tyr Trp Ser Cys Val Thr Gln Lys
                800
                                    805
His Thr Lys Leu Ala Lys Leu Ala Phe Trp Leu Leu Ala Val Pro
                815
                                    820
Ala Val Gly Ala Arg Ser Gly Cys Val Asn Met Cys Glu Gln Ala
                830
                                    835
Leu Leu Ile Lys Arg Arg Leu Leu Ser Pro Glu Asp Met Asn
                                    850
                845
Lys Leu Met Phe Leu Lys Ser Asn Met Leu
                860
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Ile Arg Val Ile Phe Asn Leu Ile Phe Leu Ile Leu Gly Lys Tyr

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Ile Ala Ser Thr Gln Arg Pro Asp Gly Thr Trp Arg Lys Gln Arg
                                     40
Arg Val Lys Glu Gly Tyr Val Pro Gln Glu Glu Val Pro Val Tyr
Glu Asn Lys Tyr Val Lys Phe Phe Lys Ser Lys Pro Glu Leu Pro
                                    70
                65
Pro Gly Leu Ser Pro Glu Ala Thr Ala Pro Val Thr Pro Ser Arg
                80
                                    85
Pro Glu Gly Glu Pro Gly Leu Ser Lys Thr Ala Lys Arg Asn
                95
                                   100
Leu Lys Arg Lys Glu Lys Arg Arg Gln Gln Glu Lys Gly Glu
               110
                                   115
Ala Glu Ala Leu Ser Arg Thr Leu Asp Lys Val Ser Leu Glu Glu
                125
                                   130
                                                        135
Thr Ala Gln Leu Pro Ser Ala Pro Gln Gly Ser Arg Ala Ala Pro
                140
                                   145
Thr Ala Ala Ser Asp Gln Pro Asp Ser Ala Ala Thr Thr Glu Lys
                155
                                   160
Ala Lys Lys Ile Lys Asn Leu Lys Lys Lys Leu Arg Gln Val Glu
                170
                                   175
Glu Leu Gln Gln Arg Ile Gln Ala Gly Glu Val Ser Gln Pro Ser
                                   190
                185
Lys Glu Gln Leu Glu Lys Leu Ala Arg Arg Arg Ala Leu Glu Glu
                                    205
Glu Leu Glu Asp Leu Glu Leu Gly Leu
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Arg Ala Phe Arg Arg Gln Arg Gly Arg Arg Pro Ser Gln Asp Asp
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                                     25
Val Glu Ala Ala Pro Glu Glu Thr Arg Ala Leu Tyr Arg Glu Tyr
                 35
                                     40
Arg Thr Leu Lys Arg Thr Thr Gly Gln Ala Gly Gly Gly Leu Arg
                                     55
                 50
Ser Ser Glu Ser Leu Pro Ala Ala Ala Glu Glu Ala Pro Glu Pro
                 65
                                     70
Arg Cys Trp Gly Pro His Leu Asn Arg Ala Ala Thr Lys Ser Pro
                 80
                                     85
Gln Pro Thr Pro Gly Arg Ser Arg Gln Gly Ser Val Pro Asp Tyr
                 95
                                   100
Gly Gln Arg Leu Lys Ala Asn Leu Lys Gly Thr Leu Gln Ala Gly
                                   115
Pro Ala Leu Gly Arg Arg Pro Trp Pro Leu Gly Arg Ala Ser Ser
                125
                                    130
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Lys	Ala	Ser	Thr	Pro	Lys	Pro	Pro	Gly	Thr 145	Gly	Pro	Val	Pro	Ser 150
Phe	Ala	Glu	Lys		Ser	Asp	Glu	Pro	Pro 160	Gln	Leu	Pro	Glu	
Gln	Pro	Arg	Pro	Gly 170	Arg	Leu	Gln	His	Leu 175	Gln	Ala	Ser	Leu	Ser 180
Gln	Arg	Leu	Gly	Ser 185	Leu	Asp	Pro	Gly	Trp 190	Leu	Gln	Arg	Суз	His 195
Ser	Glu	Val	Pro	Asp 200	Phe	Leu	Gly	Ala	Pro 205	Lys	Ala	Cys	Arg	Pro 210
Asp	Leu	Gly	Ser	Glu 215	Glu	Ser	Gln	Leu	Leu 220	Ile	Pro	Gly	Glu	Ser 225
			Gly	230	_				235	_				240
			Glu	245					250					255
		_	Gly	260	_	_	_	_	265					270
			Gln	275					280			_		285
		_	Ala	290					295		_			300
			Gln	305					310	_				315
			Tyr	320					325					330
			Glu	335					340					345
			Asp	350	_		_		355				_	360
_		_	Val	365	_	_			370					375
			Trp	380					385					390
			Gly	395					400					405
			Phe	410		_			415	-				420
			Asp	425	,			_	430					435
			Pro	440					445					450
			Tyr	455		_			460					465
			Val	470					475					480
	_		Gly	485					490					495
			Leu	500					505					510
			Leu	515					520		_			525
			Val	530					535			_	-	540
AST	ser	отА	Leu	Pro 545	Pro	суз	ren	гуз	550	Ala	cys	TTE	nls	555

		_,	_		~1 .	•	~1		1	•	01	T	T1.	3
Gly	Met	Thr	Arg	Lys 560	GIn	Arg	GIu	Ser	565	ren	GIN	гÀа	TTE	Arg 570
Ala	Ala	Gln	Val	His 575	Val	Leu	Met	Leu	Thr 580	Pro	Glu	Ala	Leu	Val 585
Gly	Ala	Gly	Gly	Leu 590	Pro	Pro	Ala	Ala	G1n 595	Leu	Pro	Pro	Val	Ala 600
Phe	Ala	Сув	Ile	Asp 605	Glu	Ala	His	Сув	Leu 610	Ser	Gln	Trp	Ser	His 615
Asn	Phe	Arg	Pro		Tyr	Leu	Arg	Val		Lys	Val	Leu	Arg	
Arg	Met	Gly	Val		Сув	Phe	Leu	Gly		Thr	Ala	Thr	Ala	
Arg	Arg	Thr	Ala		Asp	Val	Ala	Gln		Leu	Ala	Val	Ala	
Glu	Pro	Asp	Leu		Gly	Pro	Ala	Pro		Pro	Thr	Asn	Leu	
Leu	Ser	Val	Ser		Asp	Arg	Asp	Thr		Gln	Ala	Leu	Leu	
Leu	Leu	Gln	Gly		Arg	Phe	Gln	Asn		Asp	Ser	Ile	Ile	
Tyr	Cys	Asn	Arg		Glu	Asp	Thr	Glu		Ile	Ala	Ala	Leu	
Arg	Thr	Сув	Leu		Ala	Ala	Trp	Val		Gly	Ser	Gly	Gly	
Ala	Pro	Lys	Thr		Ala	Glu	Ala	Tyr		Ala	Gly	Met	Сув	
Arg	Glu	Arg	Arg		Pro	Gln	Gly	Glu		Leu	Arg	Glu	Leu	
Arg	His	Val	His		Asp	Ser	Thr	Asp		Leu	Ala	Va1	Lys	
Leu	Val	Gln	Arg		Phe	Pro	Ala	Сув	–	Сув	Thr	Сув	Thr	
Pro	Pro	Ser	Glu		Glu	Gly	Ala	Val		Gly	Glu	Arg	Pro	Val 810
Pro	Lys	Tyr	Pro		Gln	Glu	Ala	Glu		Leu	Ser	His	Gln	Ala 825
Ala	Pro	Gly	Pro		Arg	Val	Суз	Met	Gly 835	His	Glu	Arg	Ala	Leu 840
Pro	Ile	Gln	Leu	Thr 845	Val	Gln	Ala	Leu	Asp 850	Met	Pro	Glu	Glu	Ala 855
Ile	Glu	Thr	Leu		Суз	Tyr	Leu	Glu	Leu 865	His	Pro	His	His	Trp 870
Leu	Glu	Leu	Leu		Thr	Thr	Tyr	Thr	His 880	Сув	Arg	Leu	Asn	Cys 885
Pro	Gly	Gly	Pro		Gln	Leu	Gln	Ala	Leu 895	Ala	His	Arg	Суз	Pro 900
Pro	Leu	Ala	Val	Cys 905	Leu	Ala	Gln	Gln	Leu 910	Pro	G1u	Asp	Pro	Gly 915
Gln	Gly	Ser	Ser		Va1	Glu	Phe	Asp	Met 925		Lys	Leu	Val	Asp 930
Ser	Met	Gly	Trp		Leu	Ala	Ser	Val		Arg	Ala	Leu	Сув	
Leu	Gln	Trp	Asp		Glu	Pro	Arg	Thr		Val	Arg	Arg	Gly	Thr 960
Gly	Val	Leu	Val		Phe	Ser	Glu	Leu		Phe	His	Leu	Arg	Ser 975

Pro Gly Asp Leu Thr Ala Glu Glu Lys Asp Gln Ile Cys Asp Phe 985 Leu Tyr Gly Arg Val Gln Ala Arg Glu Arg Gln Ala Leu Ala Arg 1000 995 1005 Leu Arg Arg Thr Phe Gln Ala Phe His Ser Val Ala Phe Pro Ser 1015 Cys Gly Pro Cys Leu Glu Gln Gln Asp Glu Glu Arg Ser Thr Arg 1025 1030 1035 Leu Lys Asp Leu Leu Gly Arg Tyr Phe Glu Glu Glu Glu Gln Gln 1040 1045 1050 Glu Pro Gly Gly Met Glu Asp Ala Gln Gly Pro Glu Pro Gly Gln 1060 1055 Ala Arg Leu Gln Asp Trp Glu Asp Gln Val Arg Cys Asp Ile Arg 1070 1075 Gln Phe Leu Ser Leu Arg Pro Glu Glu Lys Phe Ser Ser Arg Ala 1090 1085 Val Ala Arg Ile Phe His Gly Ile Gly Ser Pro Cys Tyr Pro Ala 1100 1105 1110 Gln Val Tyr Gly Gln Asp Arg Arg Phe Trp Arg Lys Tyr Leu His 1115 1120 1125 Leu Ser Phe His Ala Leu Val Gly Leu Ala Thr Glu Glu Leu Leu 1130 1135 Gln Val Ala Arg

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160
Pro His Pro Cys Asp Thr Ala Cys Pro Ala Pro Leu Pro Val Val
                170
                                    175
Leu Val Ala Pro Arg Ser Thr Ile Leu Ser Met Ser Arg Thr Trp
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Thr Cys Arg Arg Trp Ala Val Ala Pro Cys Arg Ala Glu Lys Leu
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Met Cys Ser Ser Ser Arg Ser
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Asn Met Glu Thr Thr Arg Thr Arg Cys Trp Lys Asp Tyr Val Ser
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                                     25
Leu Arg Ile Glu Ala Ile Arg Ala Glu Tyr Gln Lys Met Pro Ala
                 35
                                     40
Phe Leu His Glu Glu Glu Gln His His Leu Glu Arg Leu Arg Lys
Glu Gly Glu Asp Ile Phe Gln Gln Leu Asn Glu Ser Lys Ala Arg
                                     70
Met Glu His Ser Arg Glu Leu Leu Arg Gly Met Tyr Glu Asp Leu
                 80
                                     85
Lys Gln Met Cys His Lys Ala Asp Val Glu Leu Leu Gln Ala Phe
Gly Asp Ile Leu His Arg Tyr Glu Ser Leu Leu Gln Val Ser
                110
                                    115
Glu Pro Val Asn Pro Glu Leu Ser Ala Gly Pro Ile Thr Gly Leu
                125
                                    130
Leu Asp Ser Leu Ser Gly Phe Arg Val Asp Phe Thr Leu Gln Pro
               140
                                    145
Glu Arg Ala Asn Ser His Ile Phe Leu Cys Gly Asp Leu Arg Ser
               155
                                    160
Met Asn Val Gly Cys Asp Pro Gln Asp Asp Pro Asp Ile Thr Gly
               170
                                   175
Lys Ser Glu Cys Phe Leu Val Trp Gly Ala Gln Ala Phe Thr Ser
               185
                                    190
Gly Lys Tyr Tyr Trp Glu Val His Met Gly Asp Ser Trp Asn Trp
                200
                                    205
Ala Phe Gly Val Cys Asn Asn Tyr Trp Lys Glu Lys Arg Gln Asn
                215
                                    220
Asp Lys Ile Asp Gly Glu Glu Gly Leu Phe Leu Leu Gly Cys Val
               230
                                   235
Lys Glu Asp Thr His Cys Ser Leu Phe Thr Thr Pro Leu Val
               245
                                   250
Val Gln Tyr Val Pro Arg Pro Thr Ser Thr Val Gly Leu Phe Leu
               260
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Asp Cys Glu Gly Arg Ile Met Ser Phe Val Asp Val Asp Gln Ser 275 280 Phe Leu Ile Tyr Ile Ile Pro Asn Cys Ser Phe Ser Pro Pro Leu 290 Arg Pro Ile Phe Cys Cys Ser His Phe <210> 43 <211> 483 <212> PRT <213> Homo sapiens <220> <221> misc_feature <223> Incyte ID No: 368741CD1 <400> 43 Met Trp Leu Gly Thr Ser Gly Lys Ser Gly Leu Pro Gly His Cys 10 Leu Glu Asn Pro Leu Gln Glu Cys His Pro Ala Gln Leu Glu Glu 20 25 Trp Ala Leu Lys Gly Ile Ser Arg Pro Ser Val Ile Ser Gln Pro 35 40 Glu Gln Lys Glu Glu Pro Trp Val Leu Pro Leu Gln Asn Phe Glu 50 55 Ala Arg Lys Ile Pro Arg Glu Ser His Thr Asp Cys Glu His Gln 70 Val Ala Lys Leu Asn Gln Asp Asn Ser Glu Thr Ala Glu Gln Cys 85 Gly Thr Ser Ser Glu Arg Thr Asn Lys Asp Leu Ser His Thr Leu 95 100 Ser Trp Gly Gly Asn Trp Glu Gln Gly Leu Glu Leu Glu Gly Gln 115 Tyr Gly Thr Leu Pro Gly Glu Gly Gln Leu Glu Ser Phe Ser Gln 125 130 Glu Arg Asp Leu Asn Lys Leu Leu Asp Gly Tyr Val Gly Glu Lys 140 145 Pro Met Cys Ala Glu Cys Gly Lys Ser Phe Asn Gln Ser Ser Tyr 155 160 Leu Ile Arg His Leu Arg Thr His Thr Gly Glu Arg Pro Tyr Thr 170 175 Cys Ile Glu Cys Gly Lys Gly Phe Lys Gln Ser Ser Asp Leu Val 185 190 Thr His Arg Arg Thr His Thr Gly Glu Lys Pro Tyr Gln Cys Lys 200 205 Gly Cys Glu Lys Lys Phe Ser Asp Ser Ser Thr Leu Ile Lys His 215 220 Gln Arg Thr His Thr Gly Glu Arg Pro Tyr Glu Cys Pro Glu Cys 230 235 Gly Lys Thr Phe Gly Arg Lys Pro His Leu Ile Met His Gln Arg 245 250 Thr His Thr Gly Glu Lys Pro Tyr Ala Cys Leu Glu Cys His Lys 260 265 Ser Phe Ser Arg Ser Ser Asn Phe Ile Thr His Gln Arg Thr His 275 280 Thr Gly Val Lys Pro Tyr Arg Cys Asn Asp Cys Gly Glu Ser Phe

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290
                                     295
Ser Gln Ser Ser Asp Leu Ile Lys His Gln Arg Thr His Thr Gly
                                     310
Glu Arg Pro Phe Lys Cys Pro Glu Cys Gly Lys Gly Phe Arg Asp
                320
                                     325
Ser Ser His Phe Val Ala His Met Ser Thr His Ser Gly Glu Arg
                335
                                     340
Pro Phe Ser Cys Pro Asp Cys His Lys Ser Phe Ser Gln Ser Ser
                350
                                     355
His Leu Val Thr His Gln Arg Thr His Thr Gly Glu Arg Pro Phe
                365
                                     370
Lys Cys Glu Asn Cys Gly Lys Gly Phe Ala Asp Ser Ser Ala Leu
                380
                                     385
Ile Lys His Gln Arg Ile His Thr Gly Glu Arg Pro Tyr Lys Cys
                395
                                     400
Gly Glu Cys Gly Lys Ser Phe Asn Gln Ser Ser His Phe Ile Thr
                410
                                     415
His Gln Arg Ile His Leu Gly Asp Arg Pro Tyr Arg Cys Pro Glu
                425
                                     430
Cys Gly Lys Thr Phe Asn Gln Arg Ser His Phe Leu Thr His Gln
                440
                                     445
Arg Thr His Thr Gly Glu Lys Pro Phe His Cys Ser Lys Cys Asn
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Lys Ser Phe Arg Gln Lys Ala His Leu Leu Cys His Gln Asn Thr
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His Leu Ile
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Phe Phe <210> 45 <211> 200 <212> PRT <213> Homo sapiens <220> <221> misc_feature <223> Incyte ID No: 7506253CD1 <400> 45 Met Asn Asn Lys Phe Asp Ala Leu Lys Asp Asp Asp Ser Gly Asp 5 10 His Asp Gln Asn Glu Glu Asn Ser Thr Gln Lys Asp Gly Glu Lys 20 25 Glu Lys Thr Glu Arg Asp Lys Asn Gln Ser Ser Lys Arg Lys 40 Val Glu Gln Phe Trp Arg Phe Tyr Ser His Met Val Arg Pro Gly 50 55 Asp Leu Thr Gly His Ser Asp Phe His Leu Phe Lys Glu Gly Ile 65 70 Lys Pro Met Trp Glu Asp Asp Ala Asn Lys Asn Gly Gly Lys Trp 85 Ile Ile Arg Leu Arg Lys Gly Leu Ala Ser Arg Cys Trp Glu Asn 95 100 Leu Ile Leu Ala Met Leu Gly Glu Gln Phe Met Val Gly Glu Glu 115 110 Ile Cys Gly Ala Val Val Ser Val Arg Phe Gln Glu Asp Ile Ile 130 Ser Ile Trp Asn Lys Thr Ala Ser Asp Gln Ala Thr Thr Ala Arg 140 145 Ile Arg Asp Thr Leu Arg Arg Val Leu Asn Leu Pro Pro Asn Thr 160 155 Ile Met Glu Tyr Lys Thr His Thr Asp Ser Ile Lys Met Pro Gly 170 175 Arg Leu Gly Pro Gln Arg Leu Leu Phe Gln Asn Leu Trp Lys Pro 185 190 Arg Leu Asn Val Pro 200 <210> 46 <211> 123 <212> PRT <213> Homo sapiens <220> <221> misc_feature <223> Incyte ID No: 7506353CD1

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<211> 874

<212> PRT

<213> Homo sapiens

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<221> misc_feature

<223> Incyte ID No: 7506372CD1

<400> 47

Met Tyr Gly Ala Ser Gly Gly Arg Ala Lys Pro Glu Arg Lys Ser 10 Gly Ala Lys Glu Glu Ala Gly Pro Gly Gly Ala Gly Gly Gly Ser Arg Val Glu Leu Leu Val Phe Gly Tyr Ala Cys Lys Leu Phe Arg Asp Asp Glu Arg Ala Leu Ala Gln Glu Gln Gly Gln His Leu Ile Pro Trp Met Gly Asp His Lys Ile Leu Ile Asp Arg Tyr Asp Gly Arg Gly His Leu His Asp Leu Ser Glu Tyr Asp Ala Glu Tyr 80 Ser Thr Trp Asn Arg Asp Tyr Gln Leu Ser Glu Glu Glu Ala Arg 100 95 Ile Glu Ala Leu Cys Asp Glu Glu Arg Tyr Leu Ala Leu His Thr 110 115 Asp Leu Leu Glu Glu Glu Ala Arg Gln Glu Glu Glu Tyr Lys Arg 125 130 Leu Ser Glu Ala Leu Ala Glu Asp Gly Ser Tyr Asn Ala Val Gly 140 145 Phe Thr Tyr Gly Ser Asp Tyr Tyr Asp Pro Ser Glu Pro Thr Glu 155 160 Glu Glu Glu Pro Ser Lys Gln Arg Glu Lys Asn Glu Ala Glu Asn 170 175 Leu Glu Glu Asn Glu Glu Pro Phe Val Ala Pro Leu Gly Leu Ser 185 190 Val Pro Ser Asp Val Glu Leu Pro Pro Thr Ala Lys Met His Ala 200 205 Ile Ile Glu Arg Thr Ala Ser Phe Val Cys Arg Gln Gly Ala Gln 215 220 Phe Glu Ile Met Leu Lys Ala Lys Gln Ala Arg Asn Ser Gln Phe

				230					235					240
Yab	Phe	Leu	Arg		Asp	His	Tyr	Leu	Asn 250	Pro	Tyr	Tyr	Lys	Phe 255
Ile	Gln	Lys	Ala		Lys	G1u	Gly	Arg	Tyr 265	Thr	Val	Leu	Ala	Glu 270
Asn	Lys	Ser	Asp		Lys	Lys	Lys	Ser	Gly 280	Val	Ser	Ser	Asp	Asn 285
Glu	Asp	Asp	Asp		Glu	Glu	Asp	Gly	Asn 295	Tyr	Leu	His	Pro	Ser 300
Leu	Phe	Ala	Ser	Lys 305	Lys	Суз	Asn	Arg	Leu 310	Glu	Glu	Leu	Met	Lys 315
Pro	Leu	Lys	Val	Val 320	Asp	Pro	Asp	His	Pro 325	Leu	Ala	Ala	Leu	Val 330
Arg	Lys	Ala	Gln	Ala 335	qaA	Ser	Ser	Thr	Pro 340	Thr	Pro	His	Asn	Ala 345
Asp	Gly	Ala	Pro	Val 350	Gln	Pro	Ser	Gln	Val 355	Glu	Tyr	Thr	Ala	Asp 360
Ser	Thr	Val	Ala	Ala 365	Met	Tyr	Tyr	Ser	Tyr 370	Tyr	Met	Leu	Pro	Asp 375
Gly	Thr	Tyr	Сув	Leu 380	Ala	Pro	Pro	Pro	Pro 385	Gly	Ile	Asp	Val	Thr 390
Thr	Tyr	Tyr	Ser	Thr 395	Leu	Pro	Ala	Gly	Val 400	Thr	Val	Ser	Asn	Ser 405
Pro	Gly	Val	Thr	Thr 410	Thr	Ala	Pro	Pro	Pro 415	Pro	Gly	Thr	Thr	Pro 420
Pro	Pro	Pro	Pro	Thr 425	Thr	Ala	Glu	Thr	Ser 430	Ser	Gly	Ala	Thr	Ser 435
Thr	Thr	Thr	Thr	Thr 440	Ser	Ala	Leu	Ala	Pro 445	Val	Ala	Ala	Ile	Ile 450
Pro	Pro	Pro	Pro	Asp 455	Val	Gln	Pro	Val	Ile 460	Asp	Lys	Leu	Ala	Glu 465
Tyr	Val	Ala	Arg	Asn 470	Gly	Leu	Lys	Phe	Glu 475	Thr	Ser	Val	Arg	Ala 480
Lys	Asn	Asp	Gln	Arg 485	Phe	Glu	Phe	Leu	Gln 490	Pro	Trp	His	Gln	Tyr 495
Asn	Ala	Tyr	Tyr	Glu 500	Phe	Lys	Lys	Gln	Phe 505	Phe	Leu	Gln	Lys	Glu 510
-	Gly	•		515					520					525
	Asp			530					535					540
Ala	Pro	Glu	Asp	Ala 545	Ala	Glu	Val	Gly	Ala 550	Arg	Ala	Gly	Ser	Gly 555
_	Lys			560					565					570
Leu	Val	Lys	Ala	Lys 575	Gln	Lys	Leu	Glu	Asp 580	Arg	Leu	Ala	Ala	Ala 585
Ala	Arg	Glu	Lys	Leu 590	Ala	Gln	Ala	Ser	Lys 595	Glu	Ser	Lys	Glu	600 Lys
	Leu			605					610					615
	Leu			620					625					630
	Ser			635					640					645
Leu	Leu	Thr	Gly	Gly	Arg	Pro	Leu	Pro	Thr	Leu	Glu	Val	Lys	Pro

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655
Pro Asp Arg Pro Ser Ser Lys Ser Lys Asp Pro Pro Arg Glu Glu
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                665
Glu Lys Glu Lys Lys Lys Lys His Lys Lys Arg Ser Arg Thr
                                    685
                680
Arg Ser Arg Ser Pro Lys Tyr His Ser Ser Ser Lys Ser Arg Ser
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Arg Ser His Ser Lys Ala Lys His Ser Leu Pro Ser Ala Tyr Arg
                                   715
               710
Thr Val Arg Arg Ser Arg Ser Arg Ser Arg Ser Pro Arg Arg
                                                       735
               725
                                   730
Ala His Ser Pro Glu Arg Arg Glu Glu Arg Ser Val Pro Thr
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                                   745
Ala Tyr Arg Val Ser Arg Ser Pro Gly Ala Ser Arg Lys Arg Thr
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                                   760
Arg Ser Arg Ser Pro His Glu Lys Lys Lys Lys Arg Arg Ser Arg
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                                   775
Ser Arg Thr Lys Ser Lys Ala Arg Ser Gln Ser Val Ser Pro Ser
                                   790.
               785
Lys Gln Ala Ala Pro Arg Pro Ala Ala Pro Ala Ala His Ser Ala
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                                   805
His Ser Ala Ser Val Ser Pro Val Glu Ser Arg Gly Ser Ser Gln
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               815
Glu Arg Ser Arg Gly Val Ser Gln Glu Lys Glu Ala Gln Ile Ser
               830
                                    835
Ser Ala Ile Val Ser Ser Val Gln Ser Lys Ile Thr Gln Asp Leu
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Met Ala Lys Val Arg Ala Met Leu Ala Ala Ser Lys Asn Leu Gln
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Thr Ser Ala Ser
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Arg Leu Leu Gly His Leu Ala Ala Cys Leu Arg Gln Leu Gly Pro
Ser Arg Arg Pro Ala Ser Leu Ser Pro Ala Ala Pro Ala Glu Ala
                125
                                    130
Pro Ala Pro Glu Val Tyr Ala Gly Arg Pro Leu Leu Pro Ser Leu
                140
                                    145
Gly Gly Pro Phe Pro Leu Leu Ala Pro Pro Leu Leu Pro Gly Leu
                155
                                    160
Thr Arg Ala Leu Pro Ala Ala Pro Arg Ala Gly Pro Gln Gly Pro
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                                    175
Gly Gly Pro Trp Arg Pro Trp Leu Arg
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Glu Arg Glu Leu Ile Cys Pro Ala Cys Lys Glu Leu Phe Thr His
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                                     25
Pro Leu Ile Leu Pro Cys Gln His Ser Ile Cys His Lys Cys Val
                                     40
Lys Glu Leu Leu Thr Leu Asp Asp Ser Phe Asn Asp Val Gly
                                     55
Ser Asp Asn Ser Asn Gln Ser Ser Pro Arg Leu Arg Leu Pro Ser
                                     70
                 65
Pro Ser Met Asp Lys Ile Asp Arg Ile Asn Arg Pro Gly Trp Lys
                 80
                                     85
Arg Asn Ser Leu Thr Pro Arg Thr Thr Val Phe Pro Cys Pro Gly
                                    100
Cys Glu His Asp Val Asp Leu Gly Glu Arg Gly Ile Asn Gly Leu
                110
                                    115
Phe Arg Asn Phe Thr Leu Glu Thr Ile Val Glu Arg Tyr Arg Gln
                125
                                    130
Ala Ala Arg Ala Ala Thr Ala Ile Met Cys Asp Leu Cys Lys Pro
                140
                                    145
Pro Pro Gln Glu Ser Thr Lys Ser Cys Met Asp Cys Ser Ala Ser
                                    160
                155
Tyr Cys Asn Glu Cys Phe Lys Ile His His Pro Trp Gly Thr Ile
                                    175
                170
Lys Ala Gln His Glu Tyr Val Gly Pro Thr Thr Asn Phe Arg Pro
                185
                                    190
Lys Ile Leu Met Cys Pro Glu His Glu Thr Glu Arg Ile Asn Met
                200
                                    205
Tyr Cys Glu Leu Cys Arg Arg Pro Val Cys His Leu Cys Lys Leu
                                    220
                215
Gly Gly Asn His Ala Asn His Arg Val Thr Thr Met Ser Ser Ala
                230
                                    235
Tyr Lys Thr Leu Lys Glu Lys Leu Ser Lys Asp Ile Asp Tyr Leu
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				245					250					255
Tla	Glv	Lve	Glu	245 Ser	Gln	Val	Lva	Ser		Tle	Ser	Glu	Leu	
110	Cly	D , 3	014	260	· · · ·		2,5	50-	265					270
Leu	Leu	Met	Lys	Glu	Thr	Glu	Сув	Asn	Gly	Glu	Arg	Ala	Lys	Glu
				275					280					285
Glu	Ala	Ile	Thr	His	Phe	Glu	Lys	Leu		Glu	Val	Leu	Glu	Glu
				290			_	_	295		_	_	_	300
Arg	Lys	Ser	Ser		Leu	Lys	Ala	Ile	-	Ser	Ser	Lys	Lys	
λ	Tan	3	T	305	~1 m	Thr	015	Mot	310	01	Th. 12	Gl n	Gly	315
Arg	Leu	Asp	гÀв	320	GIN	ing	GIN	met	325	GIU	ıyı	GIII	GIY	330
Leu	Glu	Asn	Asn		Leu	Val	Glv	Tvr		Gln	Glu	Va1	Leu	
				335			-	-	340					345
G1u	Thr	Asp	Gln	Ser	Cys	Phe	Val	Gln	Thr	Ala	Lys	Gln	Leu	His
				350					355					360
Leu	Arg	Ile	Gln	_	Ala	Thr	Glu	Ser		Arg	Ser	Phe	Arg	
31 -	21-	01-	mh	365	Dh.	G1	3.00	Th	370	7701	3 an	Th~	Co.	375
Ala	Ala	GIN	Thr	380	Pne	Glu	wab	ıyı	385	vai	ASII	1111	per	390
Gln	Thr	Glu	Leu		Glv	Glu	Leu	Ser		Phe	Ser	Ser	Glv	
				395					400				-	405
Asp	Val	Pro	Glu	Ile	Asn	Glu	Glu	Gln	Ser	Lys	Va1	Tyr	Asn	Asn
				410					415					420
Ala	Leu	Ile	Asn		His	His	Pro	Glu		Asp	Lys	Ala	Asp	
Пь	1701	7	01	425	3	T 140	T1.	λan	430	7	7 an	61	Wor	435
туг	vaı	Leu	GIU	440	Arg	ГЛЗ	TTE	ASN	445	Авр	Авр	GIU	Mec	450
Trp	Asn	Glu	Ile		Val	Cys	Glv	Thr		Lvs	Ile	Ile	Gln	
•				455			•		460	-				465
Leu	Glu	Asn	Ser	Ser	Thr	Tyr	Ala	Phe	Arg	Val	Arg	Ala	Tyr	Lys
				470					475					480
Gly	Ser	Ile	Сув		Pro	Cys	Ser	Arg		Leu	Ile	Leu	His	
B	B	31-	D	485	DL-	Ser	DL -	*	490	3	01	T	~ ~	495
Pro	PIO	Ala	Pro	500	Pne	Set	Pne	Leu	505	Asp	GIU	гàя	Cys	510
Tvr	Asn	Asn	Glu		Leu	Leu	Leu	Asn		Lys	Arg	Asp	Arg	
-				515					520	_	_	-	_	525
Glu	Ser	Arg	Ala	Gly	Phe	Asn	Leu	Leu	Leu	Ala	Ala	Glu	Arg	Ile
				530					535				_	540
Gln	Val	Gly	Tyr	Tyr 545	Thr	Ser	Leu	-	_		Ile	Gly	Asp	
G1v	Tle	ጥኮሎ	Lve		Lva	His	Dho		550		Δra	17a1	Glu	555 Pro
GTĀ	116	1111	гур	560	Lys	nis	File	Пр	565	FIIG	ary	Vai	Giu	570
Tvr	Ser	Tyr	Leu		Lys	Val	Gly	Val		Ser	Ser	Asp	Lys	
•		•		575	-		-		580			_	-	585
Gln	Glu	Trp	Leu	Arg	Ser	Pro	Arg	Asp	Ala	Val	Ser	Pro	Arg	Tyr
				590					595			_		600
Glu	Gln	Asp	Ser		His	Asp	Ser	Gly		Glu	Asp	Ala	Cys	
N	C	C	01=	605	Dha	Thr	Lou	17-1	610	T10	01	Mot	015	615
Asp	ser	Ser	GIII	620	Pne	Int	Leu	Val	625	116	GTA	Mer	GIII	630
Phe	Phe	Ile	Pro		Ser	Pro	Thr	Ser		Asn	Glu	Pro	Glu	
				635					640	_				645
Arg	Val	Leu	Pro	Met	Pro	Thr	Ser	Ile		Ile	Phe	Leu	Asp	
				650	_				655	_			_	660
Авр	Lys	Gly	Lys	Val	Asn	Phe	Tyr	Asp	Met	Asp	Gln	Met	Lys	Cys

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670
Leu Tyr Glu Arg Gln Val Asp Cys Ser His Thr Leu Tyr Pro Ala
                                    685
                680
Phe Ala Leu Met Gly Ser Gly Gly Ile Gln Leu Glu Glu Pro Ile
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                                    700
Thr Ala Lys Tyr Leu Glu Tyr Gln Glu Asp Met
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Met Arg Glu Val Arg Ala Arg Leu Ala Ala Thr Gln Val Glu Tyr
                35
                                    40
Ile Ser Gly Lys Val Phe Phe Thr Thr Cys Ser Asp Leu Asn Met
                50
                                    55
Leu Lys Lys Leu Lys Ser Ala Glu Arg Leu Phe Leu Leu Ile Lys
                                    70
Lys Gln Phe Pro Leu Ile Ile Ser Ser Val Ser Lys Gly Lys Ile
Phe Asn Glu Met Gln Arg Leu Ile Asn Glu Asp Pro Gly Ser Trp
                 95
                                    100
Leu Asn Ala Ile Ser Ile Trp Lys Asn Leu Leu Glu Leu Asp Ala
                110
                                   115
Lys Lys Glu Lys Leu Ser Gln Arg Asp Asp Asn Gln Leu Lys Arg
                                    130
Lys Val Gly Glu Asn Glu Ile Ile Ala Lys Lys Leu Lys Ile Glu
                                   145
                140
Gln Met Gln Lys Ile Glu Glu Asn Arg Asp Cys Gln Leu Glu Lys
                                   160
                155
Gln Ile Lys Glu Glu Thr Leu Glu Gln Arg Asp Phe Thr Thr Lys
                170
                                   175
Ser Glu Lys Phe Gln Glu Glu Glu Phe Gln Asn Asp Ile Glu Lys
                                   190
                185
Ala Ile Asp Thr His Asn Gln Asn Asp Leu Thr Phe Arg Val Ser
                200
                                    205
Cys Arg Cys Ser Gly Thr Ile Gly Lys Ala Phe Thr Ala Gln Glu
                                    220
                215
Val Gly Lys Val Ile Gly Ile Ala Ile Met Lys His Phe Gly Trp
                230
                                   235
Lys Ala Asp Leu Arg Asn Pro Gln Leu Glu Ile Phe Ile His Leu
                245
                                    250
Asn Asp Ile Tyr Ser Val Val Gly Ile Pro Val Phe Arg Val Ser
                260
                                    265
                                                        270
Leu Ala Ser Arg Ala Tyr Ile Lys Thr Ala Gly Leu Arg Ser Thr
                275
                                    280
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Ile Ala Trp Ala Met Ala Ser Leu Ala Asp Ile Lys Ala Gly Ala
                                    295
Phe Val Leu Asp Pro Met Cys Gly Leu Gly Thr Ile Leu Leu Glu
               305
Ala Ala Lys Glu Trp Pro Asp Val Tyr Tyr Val Gly Ala Asp Val
                                    325
               320
Ser Asp Ser Gln Leu Leu Gly Thr Trp Asp Asn Leu Lys Ala Ala
                                    340
               335
Gly Leu Glu Asp Lys Ile Glu Leu Leu Lys Ile Ser Val Ile Glu
                                    355
               350
Leu Pro Leu Pro Ser Glu Ser Val Asp Ile Ile Ile Ser Asp Ile
               365
                                    370
Pro Phe Gly Lys Lys Phe Lys Leu Gly Lys Asp Ile Lys Ser Ile
                380
                                    385
Leu Gln Glu Met Glu Arg Val Leu His Val Gly Gly Thr Ile Val
               395
                                    400
Leu Leu Ser Glu Asp His His Arg Arg Leu Thr Asp Cys Lys
                                    415
                410
Glu Ser Asn Ile Pro Phe Asn Ser Lys Asp Ser His Thr Asp Glu
                425
                                    430
Pro Gly Ile Lys Lys Cys Leu Asn Pro Glu Glu Lys Thr Gly Ala
                440
                                    445
Phe Lys Thr Ala Ser Thr Ser Phe Glu Ala Ser Asn His Lys Phe
                455
                                    460
Leu Asp Arg Met Ser Pro Phe Gly Ser Leu Val Pro Val Glu Cys
                                    475
Tyr Lys Val Ser Leu Gly Lys Thr Asp Ala Phe Ile Cys Lys Tyr
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                485
Lys Lys Ser His Ser Ser Gly Leu
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Ala Leu Glu Asp Pro His Gln Gly Gln Arg Ser Arg Glu Lys Ser
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                                     25
Lys Ala Thr Glu Val Met Ala Asp Met Phe Asp Gly Arg Leu Glu
                 35
                                     40
Pro Ile Val Phe Pro Pro Pro Arg Leu Pro Glu Glu Gly Val Ala
                 50
                                     55
Pro Gln Asp Pro Ala Asp Gly Gly His Thr Phe His Ile Leu Val
                 65
                                     70
Asp Ala Gly Arg Ser His Gly Ala Ile Lys Ala Gly Gln Glu Val
                                     85
Thr Pro Pro Pro Ala Glu Gly Leu Glu Ala Ala Ser Ala Ser Leu
                                    100
Thr Thr Asp Gly Ser Leu Lys Asn Gly Phe Pro Gly Glu Glu Thr
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His Gly Leu Gly Gly Glu Lys Ala Leu Glu Thr Cys Gly Ala Gly
                125
                                    130
Arg Ser Glu Ser Glu Val Ile Ala Glu Gly Lys Ala Glu Asp Val
                                    145
Lys Pro Glu Glu Cys Ala Met Phe Ser Ala Pro Val Asp Glu Lys
                                    160
Pro Gly Glu Glu Met Asp Val Ala Glu Glu Asn Arg Ala Ile
                170
                                    175
Asp Glu Val Asn Arg Glu Ala Gly Pro Gly Pro Gly Pro Gly Pro
                185
                                    190
                                                        195
Leu Asn Val Gly Leu His Leu Asn Pro Leu Glu Ser Ile Gln Leu
                200
                                    205
Glu Leu Asp Ser Val Asn Ala Glu Ala Asp Arg Ala Leu Leu Gln
                215
                                    220
Val Glu Arg Arg Phe Gly Gln Ile His Glu Tyr Tyr Leu Glu Gln
                230
                                    235
Arg Asn Asp Ile Ile Arg Asn Ile Pro Gly Phe Trp Val Thr Ala
                245
                                   250
Phe Arg His His Pro Gln Leu Ser Ala Met Ile Arg Gly Gln Asp
                260
                                    265
Ala Glu Met Leu Ser Tyr Leu Thr Asn Leu Glu Val Lys Glu Leu
                275
                                    280
Arg His Pro Arg Thr Gly Cys Lys Phe Lys Phe Phe Phe Gln Arg
                290
                                    295
Asn Pro Tyr Phe Arg Asn Lys Leu Ile Val Lys Val Tyr Glu Val
                305
                                    310
Arg Ser Phe Gly Gln Val Val Ser Phe Ser Thr Leu Ile Met Trp
                                    325
Arg Arg Gly His Gly Pro Gln Ser Phe Ile His Arg Asn Arg His
                335
                                    340
Val Ile Cys Ser Phe Phe Thr Trp Phe Ser Asp His Ser Leu Pro
                350
Glu Ser Asp Arg Ile Ala Gln Ile Ile Lys Glu Asp Leu Trp Ser
                365
                                    370
Asn Pro Leu Gln Tyr Tyr Leu Leu Gly Glu Asp Ala His Arg Ala
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Arg Arg Arg Leu Val Arg Glu Pro Val Glu Ile Pro Arg Pro Phe
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                                   400
Gly Phe Gln Cys Gly
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Val Phe Glu Pro Ser Trp Ala Glu Phe Gln Asp Pro Leu Gly Tyr
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Ile	Arg	Pro	Pro	Ala 50	Asp	Trp	Gln	Pro	Pro 55	Phe	Ala	Val	Glu	Val 60
Asp	Asn	Phe	Arg	Phe 65	Thr	Pro	Arg	Val	Gln 70	Arg	Leu	Asn	Glu	Leu 75
Glu	Ala	Gln	Thr	Arg 80	Val	Lys	Leu	Asn	Tyr 85	Leu	qaA	Gln	Ile	Ala 90
Lys	Phe	Trp	Glu	Ile 95	Gln	Gly	Ser	Ser	Leu 100	Lys	Ile	Pro	Asn	Val 105
Glu	Arg	Lys	Ile	Leu 110	Asp	Leu	Tyr	Ser	Leu 115	Ser	Lys	Gln	Суз	Asn 120
Thr	His	Pro	Phe	Asp 125	Asn	Glu	Val	Lys	Asp	Lys	Glu	Tyr	Lys	
His	Ser	Ile	Pro		Arg	Gln	Ser	Val		Pro	Ser	Lys	Phe	
Ser	Tyr	Ser	Arg		Ala	Lys	Arg	Leu		Pro	Asp	Pro	Glu	
Thr	Glu	Glu	Asp		G1u	Lys	His	Pro		Leu	Lys	Lys	Leu	
Ile	Tyr	Gly	Pro		Pro	Lys	Met	Met		Leu	Gly	Leu	Met	
Lys	Asp	Lys	Asp		Thr	Val	His	Lys		Val	Thr	Cys	Pro	
Thr	Va1	Thr	Val		Asp	Glu	Gln	Ser		Gly	Gly	Asn	Val	
Ser	Thr	Leu	Leu		Gln	His	Leu	Ser		Glu	Pro	Сув	Thr	
Thr	Thr	Met	Gln		Arg	Lys	Asn	His		Ser	Ala	Gln	Phe	
Asp	Ser	Tyr	Ile		Gln	Val	Cys	Ser		Gly	Asp	Glu	Asp	
Lys	Leu	Leu	Phe	Cys 275	Asp	Gly	Сув	Asp	Asp 280	Asn	Tyr	His	Ile	
Cys	Leu	Leu	Pro	Pro 290	Leu	Pro	Glu	Ile	Pro 295	Arg	Gly	Ile	Trp	Arg 300
Cys	Pro	Lys	Сув	Ile 305	Leu	Ala	Glu	Cys	Lys 310	Gln	Pro	Pro	Glu	
Phe	Gly	Phe	Glu	Gln 320	Ala	Thr	Gln	Glu	Tyr 325	Ser	Leu	Gln	Ser	
Gly	Glu	Met	Ala	Asp 335	Ser	Phe	Lys	Ser	Asp 340	Tyr	Phe	Asn	Met	
Val	His	Met	Val	Pro 350	Thr	Glu	Leu	Val		Lys	G1u	Phe	Trp	
Leu	Val	Ser	Ser	Ile 365	Glu	Glu	Asp	Val	Thr 370	Val	Glu	Tyr	Gly	
yab	Ile	His	Ser		Glu	Pḥe	Gly	Ser		Phe	Pro	Val	Ser	
Ser	Lys	Gln	Asn		Ser	Pro	Glu	Glu		G1u	Tyr	Ala	Thr	
Gly	Trp	Asn	Leu		Val	Met	Pro	Val		Asp	Gln	Ser	Val	
Сув	His	Ile	Asn		Asp	Ile	Ser	Gly		Lys	Val	Pro	Trp	
Tyr	Val	Gly	Met		Phe	Ser	Ala	Phe		Trp	His	Ile	Glu	

His	Trp	Ser	Tyr	Ser 455	Ile	Asn	Tyr	Leu	His 460	Trp	Gly	Glu	Pro	Lys 465
Thr	Trp	Tyr	Gly		Pro	Ser	·Leu	Ala	Ala 475	Glu	His	Leu	Glu	Glu 480
Val	Met	Lys	Met	Leu 485	Thr	Pro	Glu	Leu	Phe 490	Asp	Ser	Gln	Pro	Asp 495
			Gln	500					505					510
		_	Val	515					520					525
			Thr	530					535					540
			Phe	545					550					555
			Gly	560		_			565	_				570
			Val	575					580					585
			Glu	590					595					600
			Phe	605					610					615
			Glu	620					625					630
			Pro	635					640					645
			Leu	650					655					660
			Leu	665					670					675
			Tyr	680					685					690
			His	695					700					705
			Lys	710					715					720
•			Phe	725					730					735
			Phe	740					745					750
				755					760					165 765
				770					775					Leu 780
			Arg	785					790					795
			Gln	800					805					810
			Gln	815					820					825
			Gly	830					835					840
			Glu	845					850					855
Glu	Gln	Ala	Gln	Trp 860	Leu	Asp	Glu	Val	Lys 865	Gln	Ala	Leu	Ala	Pro 870

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Ser Ala His Arg Gly Ser Leu Val Ile Met Gln Gly Leu Leu Val
                875
                                    880
Met Gly Ala Lys Ile Ala Ser Ser Pro Ser Val Asp Lys Ala Arg
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Ala Glu Leu Gln Glu Leu Leu Thr Ile Ala Glu Arg Trp Glu Glu
                                    910
                                                        915
Lys Ala His Phe Cys Leu Glu Ala Arg Gln Lys His Pro Pro Ala
                920
                                    925
Thr Leu Glu Ala Ile Ile Arg Glu Thr Glu Asn Ile Pro Val His
                935
                                    940
Leu Pro Asn Ile Gln Ala Leu Lys Glu Ala Leu Thr Lys Ala Gln
                950
                                    955
Ala Trp Ile Ala Asp Val Asp Glu Ile Gln Asn Gly Asp His Tyr
                965
                                    970
Pro Cys Leu Asp Asp Leu Glu Gly Leu Val Ala Val Gly Arg Asp
                980
                                   985
Leu Pro Val Gly Leu Glu Glu Leu Arg Gln Leu Glu Leu Gln Val
                995
                                  1000
Leu Thr Ala His Ser Trp Arg Glu Lys Ala Ser Lys Thr Phe Leu
              1010
                                  1015
Lys Lys Asn Ser Cys Tyr Thr Leu Leu Glu Val Arg Ser Glu Thr
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Leu Thr His Ser Leu Phe Phe Ile Trp Pro Gly Cys Cys Glu Met
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Ala His Ile Met Arg Thr
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Ile Ser Glu Leu Asn Gly Lys Asn Ile Glu Asp Val Ile Ala Gln
                35
                                     40
Gly Ile Gly Lys Leu Ala Ser Val Pro Ala Gly Gly Ala Val Ala
                 50
                                     55
Val Ser Ala Ala Pro Gly Ser Ala Ala Pro Ala Ala Gly Ser Ala
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Pro Ala Ala Ala Glu Glu Lys Lys Asp Glu Lys Lys Glu Glu Ser
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Glu Glu Ser Asp Asp Met Gly Phe Gly Leu Phe Asp
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